J Clin Microbiol. 2011;49:945–54. http://dx.doi.org/10.1128/JCM.01689-10

- Lane RS, Manweiler SA, Stubbs HA, Lennette ET, Madigan JE, Lavoie PE. Risk factors for Lyme disease in a small rural community in northern California. Am J Epidemiol. 1992;136:1358–68.
- Girard YA, Travinsky B, Schotthoefer A, Fedorova N, Eisen RJ, Eisen L, et al. Population structure of the Lyme borreliosis spirochete *Borrelia burgdorferi* in the Western black-legged tick (*Ixodes pacificus*) in northern California. Appl Environ Microbiol. 2009;75:7243–52. http://dx.doi.org/10.1128/AEM.01704-09
- Travinsky B, Bunikis J, Barbour AG. Geographic differences in genetic locus linkages for *Borrelia burgdorferi*. Emerg Infect Dis. 2010;16:1147–50. http://dx. doi.org/10.3201/eid1607.091452
- Seinost G, Dykhuizen DE, Dattwyler RJ, Golde WT, Dunn JJ, Wang IN, et al. Four clones of *Borrelia burgdorferi* sensu stricto cause invasive infection in humans. Infect Immun. 1999;67:3518–24.
- Barbour AG, Travinsky B. Evolution and distribution of the *ospC* gene, a transferable serotype determinant of *Borrelia burgdorferi*. MBio. 2010;1:10.1128/mBio. 00153-10.
- Barbour AG, Bunikis J, Travinsky B, Gatewood HA, Diuk-Wasser M, Fish D, et al. Niche partitioning of *Borrelia burgdorferi* and *Borrelia miyamotoi* in the same reservoir host and arthropod vector. Am J Trop Med Hyg. 2009;81:1120–31. http://dx.doi.org/10.4269/ajtmh.2009. 09-0208
- Strle K, Jones KL, Drouin EE, Li X, Steere AC. *Borrelia burgdorferi* RST 1 (OspC Type A) Genotype is associated with greater inflammation and more severe Lyme disease. Am J Pathol. 2011;178:2726–39. http://dx.doi.org/10. 1016/j.ajpath.2011.02.018

Address for correspondence: Anne M. Kjemtrup, California Department of Public Health, 1616 Capitol Ave, MS 7307, PO Box 997377, Sacramento, CA 95899-9730. USA; email: Anne.Kjemtrup@cdph.ca.gov



# Zoonotic Baylisascaris procyonis Roundworms in Raccoons, China

To the Editor: Baylisascaris procyonis, an intestinal roundworm that infects raccoons (Procyon lotor), causes fatal or severe neural larva migrans in animals and humans (1,2). Globally,  $\approx 130$  species of wild and domesticated animals are susceptible (2). Infections in humans typically occur in children who have the disorders pica or geophagia and ingest B. procyonis eggs in items contaminated with raccoon feces (3). Clinical manifestations include ocular disease, eosinophilic encephalitis, and eosinophilic cardiac pseudotumors; severe infection can lead to death. Since 1984,  $\approx 24$  cases of B. procyonis-related human neural larva migrans have been reported, mainly in the United States (1,3-5;K.R. Kazacos, pers. comm.). Despite few cases among humans, lack of effective treatment and widespread distribution of infected raccoons in close association with humans make B. procyonis a potentially serious public health threat (2,6). The current distribution of *B. procyonis* is poorly recorded in Asia (2,7), except for Japan (8). We describe B. procvonis infections among raccoons in China as part of a series of ongoing surveys of helminthic zoonoses linked to captive exotic animals in zoologic gardens (ZGs) in China.

More than 90% of raccoons in China (n >320) are raised as exotic ornamental animals in 18 ZGs. During 2011–2013, we collected  $2\times308$  fecal samples (i.e., 1 repeat within each sampling) from 277 raccoons in 12 randomly selected ZGs (online Technical Appendix Figure 1, wwwnc. cdc.gov/EID/article/20/12/14-0970-Techapp1.pdf). Samples were stored in individual plastic bags at  $-20^{\circ}$ C until use. We examined raccoons (n =

31) at the Sichuan ZGs twice, in June 2012 and May 2013. We identified B. procyonis eggs in feces using morphologic and molecular analyses (1,2,9). The nuclear first internal transcribed spacer (428 bp) and mitochondrial cytochrome c oxidase subunit 1 (cox-1, 938 bp) genes in each sample were PCR-amplified and sequenced. B. procyonis infection was confirmed by sequencing and phylogenetic analyses of both genes (7,9). We reexamined  $\approx 60\%$  of fecal samples to validate results. Prevalence (95% CI) was calculated for the overall population and independently for female, male, juvenile, and adult raccoons. We determined differences between the tested ZG prevalence and prevalence by sex or age of raccoons using  $\chi^2$  or Fisher exact tests in SAS (SAS Institute, Cary, NC, USA); p values <0.05 were considered significant.

Building on egg-based morphologic characterization and internal transcribed spacer 1 and cox-1 genebased phylogenies using neighborjoining trees (online Technical Appendix Figure 2), we found B. procyonis in raccoon feces from 5/12 ZGs (42%; 95% CI 14%–70%), including 2 in the most densely populated provinces, Henan and Sichuan. More infections were found in western than central and eastern ZGs (4/6 and 1/6, respectively; Table, online Technical Appendix Figure 1) (p = 0.079). Fecal samples of 35 raccoons (13%; 95% CI 9%–17%) tested positive for *B. procy*onis. The mean intensity of egg shedding was 5,000 eggs per gram (range 800-11,200 eggs per gram; data not shown). No significant difference was observed in the intensity of shedding by comparing sex and age of animals, and no significant differences were noted in the mean prevalence between female and male raccoons (12% versus 14%; p = 0.677) or between adult and juvenile animals (13% versus 10%; p = 0.536).

This investigation documents the presence and prevalence of *B. procyonis* 

Table. Prevalence of Baylisascaris procyonis roundworm in	nfectio	ns amon	g captive	e raccoons	China, 2011-20	)13*

	No. <i>B. procyonis</i> -positive samples/total no. samples (%)						
	Sex		Age group				
Location, zoological gardens	М	F	Adult	Juvenile	Total		
Western region					33/146 (23)		
Chongqing	-	0/4	0/4	_	0/4		
Bifengxia	0/8	0/14	0/13	0/9	0/22, 0/22		
Chengdu	0/4	1/5 (20)	0/6	1/3 (33)	1/9 (11), 0/9		
Xi'an Wildlife	0/9	1/27 (4)	0/28	1/8 (13)	1/36 (3)		
Kunming	12/12 (100)	15/15 (100)	22/22 (100)	5/5 (100)	27/27 (100)		
Kunming Wildlife	1/24 (4)	3/24 (13)	4/48 (8)	_	4/48 (8)		
Central region					2/56 (4)		
Harbin Northern Forest	0/12	0/18	0/21	0/9	0/30		
Zhengzhou	1/5 (20)	1/11 (9)	2/12 (17)	0/4	2/16 (13)		
Changsha	0/3	0/7	0/5	0/5	0/10		
Eastern region					0/75		
Beijing	0/16	0/24	0/22	0/18	0/40		
Guangzhou	0/5	0/15	0/20	_	0/20		
Shanghai Wildlife	0/4	0/11	0/9	0/6	0/15		
Total	14/102 (14)	21/175 (12)	28/210 (13)	7/67 (10)	35/277 (13)		
p value	0.677		0.536		-		

\*Raccoons, considered to be exotic ornamental animals, are mainly kept in 18 zoologic gardens (ZGs) in China; 12 ZGs were examined for *B. procyonis* prevalence during the study period. Sichuan ZGs, including Bifengxia ZG and Chengdu ZG, were tested twice during this surveillance period. –, no raccoons in the group or no data available.

among raccoons in China. The findings imply that raccoons harboring this parasite have the potential for spreading it to humans. One reason is that captive raccoons adapt readily to humans and easily take food offered by hand; another is that communal raccoon latrine sites in ZGs are usually close to areas where humans gather, so ZG visitors may be exposed to large numbers of eggs (online Technical Appendix Figure 3). These eggs can remain viable and infective for years (2), and latrines are recognized as primary sources of transmission of B. procyonis to humans (4). Current public health initiatives to prevent B. procyonis infections in humans rely on the education of veterinary and human health care professionals, who in turn inform the public (1,6,10). Thus, veterinarians, clinicians, and public health officials in China should be more informed about this pathogen, especially in regions with large raccoon populations.

Because of a lack of clinical awareness of this illness and subsequent lack of early diagnosis and effective treatment, prevention of *B. procyonis* infection by education is essential. In addition, a strategy for eradication is needed. Heat, in the form of boiling water, steam-cleaning, or fire, is the optimal tool for killing *B. procy*onis eggs (2) and therefore can be used to decontaminate areas surrounding latrines. Within heavily contaminated areas, removing and then sterilizing the top few inches of surface soil with heat would be effective and practical (1,2). Among captive raccoon populations, particularly in China, regular deworming is also likely to be helpful in reducing novel and existing sources of infection (1-3).

Finally, although no cases of human infection have been reported in China to our knowledge, physicians should consider including B. procyonis infections in their differential diagnoses of patients with indicative features: clinical (eosinophilic encephalitis, ocular disease), epidemiologic (raccoon exposure), radiologic (white matter disease), and laboratory results (blood and CNS eosinophilia) (1,10). This study lays the foundation for future steps to educate the population of China about B. procyonis infection and to create programs to prevent the spread of this disease to humans.

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### Yue Xie, Xuan Zhou, Mei Li, Tianyu Liu, Xiaobin Gu, Tao Wang, Weimin Lai, Xuerong Peng, and Guangyou Yang

Author affiliation: Sichuan Agricultural University, Ya'an, Sichuan, China

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#### References

- Gavin PJ, Kazacos KR, Shulman ST. Baylisascariasis. Clin Microbiol Rev. 2005;18:703–18. http://dx.doi.org/10.11 28/CMR.18.4.703-718.2005
- Kazacos K. Baylisascaris procyonis and related species. In: Samuel W, Pybus M, Kocan A, editors. Parasitic diseases of wild mammals. 2nd ed. Ames (IA): Iowa State University Press; 2001. p. 301–41.
- Murray WJ, Kazacos KR. Raccoon roundworm encephalitis. Clin Infect Dis. 2004;39:1484–92. http://dx.doi.org/10. 1086/425364

#### LETTERS

- Page LK, Anchor C, Luy E, Kron S, Larson G, Madsen L, et al. Backyard raccoon latrines and risk for *Baylisascaris* procyonis transmission to humans. Emerg Infect Dis. 2009;15:1530–1. http://dx.doi. org/10.3201/eid1509.090128
- Kelly TG, Madhavan VL, Peters JM, Kazacos KR, Silvera VM. Spinal cord involvement in a child with raccoon roundworm (*Baylisascaris procyonis*) meningoencephalitis. Pediatr Radiol. 2012;42:369–73. http://dx.doi.org/10. 1007/s00247-011-2151-y
- Wise ME, Sorvillo FJ, Shafir SC, Ash LR, Berlin OG. Severe and fatal central nervous system disease in humans caused by *Baylisascaris procyonis*, the common roundworm of raccoons: a review of current literature. Microbes Infect. 2005;7:317–23. http://dx.doi.org/ 10.1016/j.micinf.2004.12.005
- Blizzard EL, Yabsley MJ, Beck MF, Harsch S. Geographic expansion of *Bay-lisascaris procyonis* roundworms, Florida, USA. Emerg Infect Dis. 2010;16:1803–4. http://dx.doi.org/10.3201/eid1611.100549
- Miyashita M. Prevalence of *Baylisascaris* procyonis in raccoons in Japan and experimental infections of the worm in laboratory animals. Journal of Urban Living and Health Association. 1993;37:137–51.
- Xie Y, Zhang Z, Niu L, Wang Q, Wang C, Lan J, et al. The mitochondrial genome of *Baylisascaris procyonis*. PLoS ONE. 2011;6:e27066. http://dx.doi.org/10.1371/ journal.pone.0027066
- Sorvillo F, Ash LR, Berlin OGW, Tatabe J, Degiorgio C, Morse SA. *Baylisascaris* procyonis: an emerging helminthic zoonosis. Emerg Infect Dis. 2002;8:355–9. http://dx.doi.org/10.3201/eid0804.010273

Address for correspondence: Guangyou Yang, Department of Parasitology, College of Veterinary Medicine, Sichuan Agricultural University, 46 Xinkang Rd, Ya'an, Sichuan, 625014, People's Republic of China; email: guangyouyang@hotmail.com



## Novel Divergent Rhabdovirus in Feces of Red Fox, Spain

To the Editor: Rhabdoviruses (family Rhabdoviridae) are enveloped single-stranded negative-sense RNA viruses belonging to the Mononegavirales order. The International Committee on Taxonomy of Viruses recognizes 11 genera (Cytorhabdovirus, Ephemerovirus, Lyssavirus, Novirhabdovirus, Nucleorhabdovirus, Perhabdovirus, Sigmavirus, Sprivivirus, Tibrovirus, Tupavirus, Vesiculovirus) (1). In addition, many recently described rhabdoviruses remain unassigned. Rhabdoviruses contain 5 major genes, encoding for nucleoprotein (N), phosphoprotein (P), matrix (M), glycoprotein (G), and RNA-dependent RNA polymerase (L). The Rhabdoviridae family includes pathogens of various animal species, humans, and plants. Viruses of the genus Lyssavirus are the most relevant to public health because they can cause rabies. Bats are the driving force within this genus; foxes and various other species of wild carnivores also can be infected with lyssaviruses and transmit them to humans and dogs (2).

During a viral metagenomic survey, conducted as described previously(3), of fecal samples collected from 4 red foxes (Vulpes vulpes) that were found dead in Álava, Basque Country, Spain, we identified the complete coding sequence and the partial leader and trailer sequence of a novel rhabdovirus, tentatively called red fox fecal rhabdovirus (RFFRV; 15,541 nt, Gen-Bank accession no. KF823814; online Technical Appendix, http://wwwnc. cdc.gov/EID/article/20/12/14-0236-Techapp1.pdf) by mapping 8,287 of the 56,519 sequence reads in the sample of a red fox. A proportion of obtained reads contained sequences that were >99% identical to mitochondrial DNA of V. vulpes, which confirmed that the sample was collected from a red fox.

The obtained sequence of RFFRV was partially confirmed by specific primers and Sanger sequencing of PCR amplicons. Five major and 3 minor open reading frames (ORFs) were identified that had a genome organization similar to that of other rhabdoviruses (Figure, panel A). No significant hits were obtained by BLAST analysis (http://blast.ncbi.nlm.gov/Blast. cgi) of N, P, M, and G nucleotide and amino acid sequences, which was reported previously for novel divergent rhabdoviruses (4).

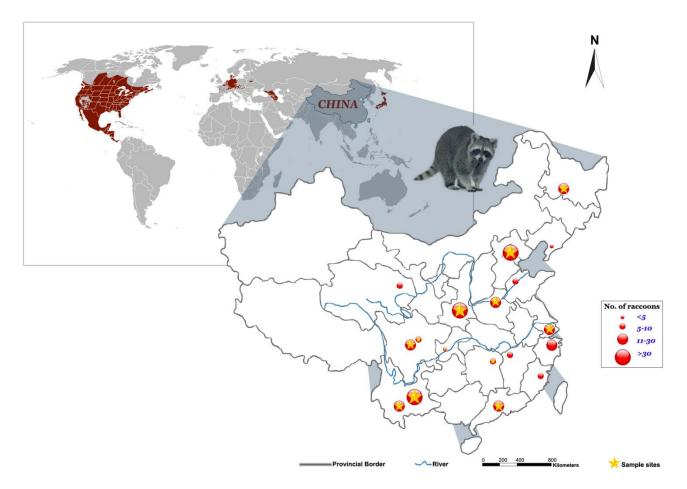
Predicted N, P, and M genes of RFFRV consist of 1,629, 2,490, and 813 nt, respectively, encoding for 543, 830, and 271 aa (online Technical Appendix Table 1). In addition to the absence of significant hits observed by BLAST analysis, no significant sequence homology was observed with known rhabdovirus proteins in pairwise alignments. Furthermore, no conserved motifs were detected in N, P, and M genes of RFFRV that are commonly observed in rhabdoviruses. However, intergenic regions between all major ORFs contained relatively conserved motifs that could be transcription termination/polyadenylation sequences (A/U) CU<sub>7</sub>, similar to other rhabdoviruses (5). Adjacent to this termination signal was a stretch of conserved nucleotides that might function as a transcription initiation signal (online Technical Appendix Table 1).

The amino acid sequence of the G protein consisted of 669 aa and contained an N terminal signal peptide (1-MYHLIVLLVMLGQRA-VA-17), a noncytoplasmic domain (aa 18–646), a transmembrane domain (647-ITAILMPLLSLAVVVGI-IMCC-667), and a cytoplasmic tail of 2 aa, similar to other rhabdovirus G proteins as predicted by using Phobius and TMHMM (http://www.cbs.dtu. dk/services/TMHMM) (6,7). We predicted 3 potential glycosylation sites in the ectodomain at positions 38–40

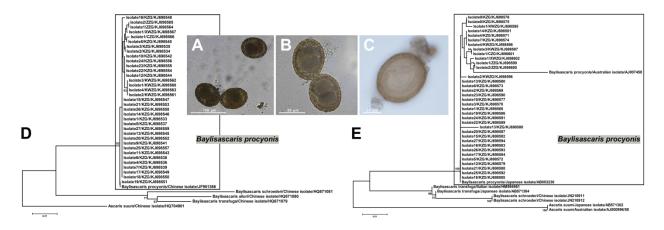
# Zoonotic *Baylisascaris procyonis* Roundworms in Raccoons, China

## **Technical Appendix**

Distribution of raccoons in China, morphological and molecular characterization of *Baylisascaris procyonis* parasitic roundworm eggs in captive raccoons in China, and the potential risk of human infection with *B. procyonis* in China.



Technical Appendix Figure 1. Distribution of raccoons in China, 2011–2013. As exotic ornamental animals, >320 raccoons are kept in 18 Zoological Gardens (ZGs) in China. The size of the red circles represents the density of raccoons in each ZG during the study period. Sampling sites are marked with yellow stars.



Technical Appendix Figure 2. Morphological and molecular characterization of *Baylisascaris procyonis* parasitic roundworm eggs in captive raccoons in China, 2011–2013. Eggs with 1–2 cells (A and B) or infective larvae (C) detected in the feces of raccoons by the flotation method with NaNO3 were identified as B. procyonis on the basis of their size and morphologic features (*1,2*). Phylogenetic relationships inferred by using GTR + I + G model-based neighbor-joining analysis on the basis of the partial sequences of mitochondrial cox-1 (D) and nuclear ITS-1 (E) from isolates of Baylisascaris species and related ascaridoids. Ascaris suum was used as an outgroup. Neighbor-joining trees were constructed using MEGA 5.0 (www.megasoftware.net). Bootstrapping with 1,000 replicates was performed to calculate the percentage reliability for each node in both genes; only values of ≥50% are shown. Horizontal branch lengths are proportional to genetic distances. Black boxes containing the 35 isolates from this study represent the B. procyonis clusters (sequences are in GenBank: KJ698533–KJ698567 for cox-1 and KJ698568–KJ698602 for ITS-1). Scale bars indicate the number of nucleotide substitutions per site. KZG, Kunming ZG; ZZG, Zhengzhou ZG; XWZG, Xi'an Wildlife ZG; CZG, Chengdu ZG; KWZG, Kunming Wildlife ZG.



Technical Appendix Figure 3. An image series showing the artificial "niche" created for raccoons in Chinese Zoological Gardens (ZGs) and depicting the potential risk of human infection with *Baylisascaris procyonis* parasitic roundworms. A) Different living environments of raccoons in the studied ZGs. B) Several typical raccoon latrines found in ZGs. C) Human-raccoon interactions, particularly between young children and raccoons. The red arrows within panel C) denote the locations of latrine sites and indicate that these sites are in close proximity to humans and therefore potentially put large numbers of infective *B. procyonis* eggs in the immediate environment of visitors.

## References

1. Gavin PJ, Kazacos KR, Shulman ST. Baylisascariasis. Clin Microbiol Rev. 2005;18:703–18. <u>PubMed</u> <u>http://dx.doi.org/10.1128/CMR.18.4.703-718.2005</u> Kazacos K. *Baylisascaris procyonis* and related species. In: Samuel W, Pybus M, Kocan A, editors.
Parasitic diseases of wild mammals. 2nd ed. Ames: Iowa State University Press; 2001. p. 301–41.