
Septicemia Caused by Tick-borne Bacterial Pathogen *Candidatus* *Neoehrlichia* *mikurensis*

Jan S. Fehr,¹ Guido V. Bloemberg,¹ Claudia Ritter,
Michael Hombach, Thomas F. Lüscher,
Rainer Weber, and Peter M. Keller

We have repeatedly detected *Candidatus Neoehrlichia mikurensis*, a bacterium first described in *Rattus norvegicus* rats and *Ixodes ovatus* ticks in Japan in 2004 in the blood of a 61-year-old man with signs of septicemia by 16S rRNA and *groEL* gene PCR. After 6 weeks of therapy with doxycycline and rifampin, the patient recovered.

Since the novel bacterial genus *Neoehrlichia* was first described in 2004, its pathogenic role in humans has remained unexplained (1). Related bacteria such as *Ehrlichia chaffeensis* and *Anaplasma phagocytophilum* are emerging tick-borne human pathogens that cause monocytic and granulocytic ehrlichiosis, respectively. These tick-borne diseases manifest themselves as febrile illness, mild transient hepatitis, transient thrombocytopenia, and occasionally as a rash. The family *Anaplasmataceae* comprises the genera *Ehrlichia*, *Anaplasma*, *Neorickettsia*, and *Aegyptianella*, and the proposed genus *Neoehrlichia*. These are all obligate intracellular bacteria, which currently are difficult or impossible to isolate and culture (2). Infections caused by agents of this bacterial family have been recognized as an emerging problem in the past 2 decades, possibly due to ecologic changes and the resulting expansion of tick populations (3).

Case Report

In August 2009, a 61-year-old Caucasian man who lived in Switzerland sought treatment at the emergency department of University Hospital in Zurich, reporting a 10-day history of malaise, temperature as high as 39.5°C, chills, and moderate dyspnea. Six weeks previously, he had undergone coronary artery bypass graft surgery and mi-

tral valve reconstruction for which prosthetic material was used. The patient had not noticed tick bites or a skin rash; neither did he recall a rodent bite. A pet dog and cat lived in his household.

Physical examination showed a reduced general health condition and a temperature of 38.5°C. Blood pressure was 109/68 mm Hg, heart rate was 86 beats/min, and oxygen saturation was 95% with 2 L nasal oxygen. No murmur was detected on cardiac auscultation. No skin or joint abnormalities were found. Laboratory tests showed elevated leukocytes (12.9×10^3 cells/ μ L), with a high fraction of neutrophils (10.1×10^3 cells/ μ L) and thrombocyte count within reference range (277×10^3 cells/ μ L); aminotransferase levels within reference ranges (aspartate aminotransferase 18 U/L, alanine aminotransferase 20 U/L); and an elevated C-reactive protein (CRP) of 68 mg/L (reference range <5 mg/L). Chest radiograph showed no signs of cardiac decompensation or of pulmonary infiltrates. Transthoracic echocardiograph showed only minor insufficiency of the aortic and tricuspid valves. In addition, degenerative alterations of aortic valve, but no vegetations, were noted with comparable findings in the follow-up echocardiograph 1 week later.

At the follow-up visit, no hints of infectious foci were found. Five sets of blood cultures were drawn with >12 h difference between the first and the last set. Antimicrobial drug treatment for endocarditis with prosthetic material, consisting of vancomycin, gentamicin, and rifampin, was initiated.

Blood cultures remained negative for microbial growth, even after extended incubation. Serologic tests for agents of culture-negative endocarditis and tick-borne diseases were performed. Enzyme immunoassays (EIAs) were positive for immunoglobulin (Ig) G antibodies reactive to *Bartonella henselae* (512) and *B. quintana* (1,024), *Coxiella burnetii* (phase II IgG titer 160), *Rickettsia rickettsii/conorii* (IgG 256), and *Rickettsia typhi* (IgG 128), *Mycoplasma pneumoniae* (index 2.7). IgM was positive only for *A. phagocytophilum* (512, atypical fluorescence pattern), presenting a low titer of IgG at this stage. Serologic test results for *Brucella* spp., *Chlamydia trachomatis*, *Chlamydia pneumoniae*, and *Borrelia burgdorferi* were negative. Species-specific PCRs for *A. phagocytophilum*, *Tropheryma whipplei*, *B. henselae*, *B. quintana*, *Legionella* spp., and *L. pneumophila* were negative.

Bacterial broad-spectrum 16S rRNA gene PCR, followed by sequence analysis, identified *Candidatus Neoehrlichia mikurensis* in 4 of 8 sequential blood samples; the 4 samples that tested positive were collected before (day 0) and during the initial phase (days 7 and 13) of an effective course of antimicrobial drug therapy (Figure 1). For 16S rRNA gene amplification, DNA was extracted and ampli-

Author affiliations: University Hospital, Zurich, Switzerland (J.S. Fehr, T.F. Lüscher, R. Weber); and University of Zurich, Zurich (G.V. Bloemberg, C. Ritter, M. Hombach, P.M. Keller)

DOI: 10.3201/eid1607.091907

¹These authors contributed equally to this article.

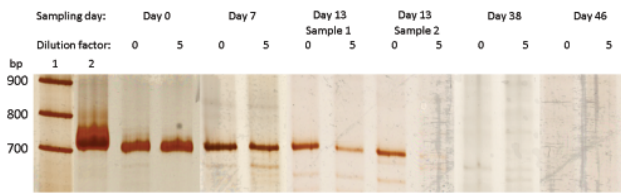


Figure 1. Polyacrylamide gel electrophoresis analysis of broad range 16S rRNA gene PCR products obtained from blood samples. Lane 1, marker, 100 bp DNA ladder (Roche DNA Marker XIV); lane 2, positive control, *Escherichia coli*; following lanes, PCR products obtained from blood specimens arranged by date of collection. For each specimen PCR products are shown obtained with undiluted (0) and 5 \times -diluted (5) DNA extracts. The 2 last negative samples are not shown.

fied from anticoagulated blood (4-mL EDTA tubes), uncoagulated blood from a BacT/ALERT SA aerobic blood culture flask (bioMérieux SA, Geneva, Switzerland), and coagulated blood (online Technical Appendix, www.cdc.gov/eid/content/16/7/1127-Techapp.pdf) as described (4). Sequences derived (GenBank accession nos. GQ501089-GQ501091) were analyzed by SmartGene IDNS software (Zug, Switzerland). We amplified and sequenced (1,150 bp; GenBank accession no. HM045824) an alternative target gene (*groEL*) with primers (online Technical Appendix) derived from published sequences (1), which showed 98% homology to previously published *Candidatus Neoehrlichia mikurensis groEL* sequences (Figure 2, panel B).

After being treated with vancomycin, gentamicin, and rifampin for 10 days, the patient became afebrile, and clinical symptoms improved. Leukocyte count was within reference range, and CRP dropped from 68 mg/L to 23 mg/L. At this time, *Candidatus Neoehrlichia mikurensis*

was detected in the first blood sample. Rifampin (450 mg 2 \times /d) was continued, and vancomycin and gentamicin were switched to oral doxycycline (100 mg 2 \times /d). Three weeks later, CRP was 1 mg/L, body temperature was within the normal range, and treatment was continued to finish a 6-week course. Two weeks after the end of treatment, the patient was seen for a follow-up visit. Neither clinical nor laboratory results raised any concern of relapse. Results of broad-range PCR of the 16S rRNA gene to detect *Candidatus Neoehrlichia mikurensis* were negative for the first time in 5 weeks since treatment began initiation and remained negative at the follow-up visit 2 weeks after the end of treatment.

Candidatus Neoehrlichia mikurensis was previously found in *Rattus norvegicus* rats and *Ixodes ovatus* ticks in Japan (1), in *R. norvegicus* rats in China (5), and in *I. ricinus* ticks in the Netherlands (6,7), Slovakia (8), and the Asian part of Russia (9). Closely related rickettsial bacteria (Figure 2) have been identified in *Procyon lotor* raccoons in the Piedmont region of Georgia, USA (10). Another closely related species (*Candidatus Ehrlichia walkeri*) has been detected in *I. ricinus* ticks collected from humans in northern Italy (11). The geographic distribution of the tick population has also been studied (12).

Our patient lives in a high-risk area for ticks in Switzerland. *I. ricinus* is the main tick species in this region. A tick-borne disease appears epidemiologically possible in this patient, who is a golfer and the owner of a large garden and thus is repeatedly exposed to the habitat of the potential vector, *I. ricinus*, even though he remembered no tick bites. Of note, only 50%–70% of patients with Lyme disease remember receiving a tick bite (13). Blood of the patient's pet animals (dog and cat) was examined by broad-range 16S rRNA gene PCR to exclude presence of bacterial pathogens.

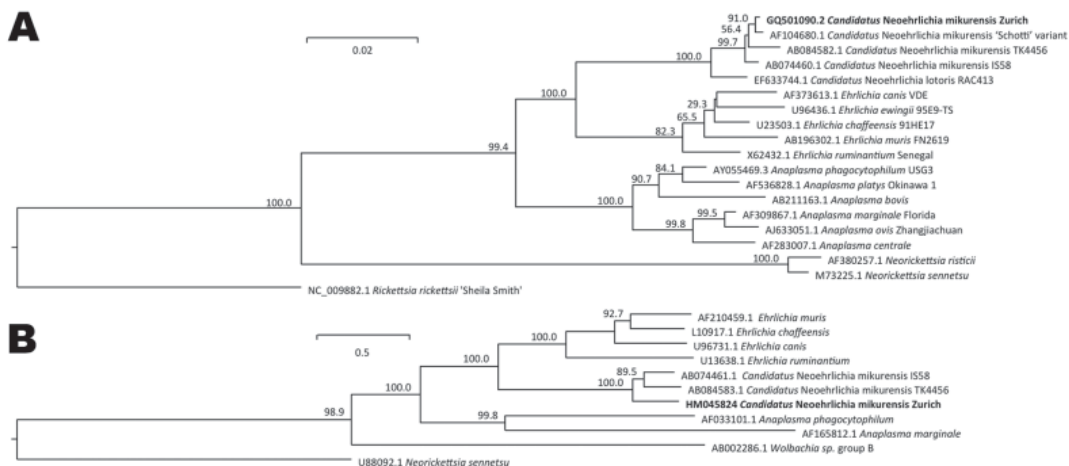


Figure 2. A) Phylogenetic tree based on the 16S rRNA gene sequences of *Candidatus Neoehrlichia mikurensis* GQ501090.2 (our patient's isolate, herein termed Zurich and indicated in **boldface**) and related organisms. The number at nodes indicates percentages of bootstrap support based on 10,000 replicates. Scale bar indicates 0.02 substitutions per nucleotide position. B) Phylogenetic tree based on the *groEL* sequences. Scale bars indicate 0.05 substitutions per nucleotide position.

In Wister rats, *Candidatus Neoehrlichia mikurensis* has been shown to infect spleen sinus endothelial cells, forming intracellular inclusions on the side of the endosome (by electron microscopy 60 days after infection) (1). Accordingly, we assumed that in human hosts, valvular endothelial cells are likely involved. The initial antimicrobial drug therapy, which contained rifampin, may already have contributed to the reduction of the bacterial load but was not completely effective (Figure 1). Thus, following the recommended guidelines for treatment of intracellular rickettsial bacteria with endocardial involvement, we changed to a 6-week course of treatment consisting of rifampin combined with doxycycline (14). After the end of the course, we observed a successful response.

We detected *Candidatus Neoehrlichia mikurensis* in 4 of 8 consecutive blood specimens, and repeated analysis showed the disappearance of the pathogen's DNA during the course of treatment (Figure 1). Laboratory diagnosis of ehrlichiosis is severely hampered because the relevant pathogens cannot be cultured on routine media. Serologic tests depend on samples collected during acute phase of illness, obtaining comparative samples in the course of the disease, and demonstrating a >4-fold increase in antibody titers.

Conclusions

We have identified *Candidatus Neoehrlichia mikurensis* in multiple blood samples of a patient who sought treatment for septicemia. Therapeutic success has been shown over time by the fact that the suggested pathogen's DNA was no longer detectable and by a favorable clinical outcome. Surveys of arthropod populations should be conducted to examine the geographic distribution of *Candidatus Neoehrlichia mikurensis*, and species-specific assays could determine the relevance of this organism in human ehrlichial diseases.

Acknowledgments

We thank Erik C. Böttger for continuous support, advice, and comments on the manuscript. We also thank the Institute of Clinical Microbiology and Immunology, St. Gallen, for help with *Mycoplasma* serology; the Institut Central des Hôpitaux du Valais, Sion, for help with *Coxiella* serology; Logolab AG, Kilchberg, Switzerland, for help with *Anaplasma* serology; and C. Silaghi for help with *A. phagocytophilum* species-specific PCR.

Dr Fehr is an infectious disease specialist and attending physician at the University Hospital of Zurich. His research interests include bacterial pathogens and HIV/AIDS. He has investigated surgical site infections and emerging pathogens in Tanzania.

Dr Bloemberg is head of the Department of Molecular Diagnostics at the Institute of Medical Microbiology, University

of Zurich. He is a specialist in molecular genetics and microbial biochemistry. His research interests include bacterial pathogens, biofilms, antibacterial drugs, and antibiotic resistance.

References

1. Kawahara M, Rikihisa Y, Isogai E, Takahashi M, Misumi H, Suto C, et al. Ultrastructure and phylogenetic analysis of '*Candidatus Neoehrlichia mikurensis*' in the family *Anaplasmataceae*, isolated from wild rats and found in *Ixodes ovatus* ticks. *Int J Syst Evol Microbiol*. 2004;54:1837–43. DOI: 10.1099/ijs.0.63260-0
2. Dumler JS, Madigan JE, Pusterla N, Bakken JS. Ehrlichioses in humans: epidemiology, clinical presentation, diagnosis, and treatment. *Clin Infect Dis*. 2007;45(Suppl 1):S45–51. DOI: 10.1086/518146
3. Estrada-Pena A. Tick-borne pathogens, transmission rates and climate change. *Front Biosci*. 2009;14:2674–87. DOI: 10.2741/3405
4. Bosshard PP, Kronenberg A, Zbinden R, Ruef C, Böttger EC, Altwegg M. Etiologic diagnosis of infective endocarditis by broad-range polymerase chain reaction: a 3-year experience. *Clin Infect Dis*. 2003;37:167–72. DOI: 10.1086/375592
5. Pan H, Liu S, Ma Y, Tong S, Sun Y. Ehrlichia-like organism gene found in small mammals in the suburban district of Guangzhou of China. *Ann N Y Acad Sci*. 2003;990:107–11. DOI: 10.1111/j.1749-6632.2003.tb07346.x
6. Schouls LM, Van De Pol I, Rijpkema SG, Schot CS. Detection and identification of *Ehrlichia*, *Borrelia burgdorferi* sensu lato, and *Bartonella* species in Dutch *Ixodes ricinus* ticks. *J Clin Microbiol*. 1999;37:2215–22.
7. van Overbeek L, Gassner F, van der Plas CL, Kastelein P, Nunes-da Rocha U, Takken W. Diversity of *Ixodes ricinus* tick-associated bacterial communities from different forests. *FEMS Microbiol Ecol*. 2008;66:72–84. DOI: 10.1111/j.1574-6941.2008.00468.x
8. Spitaliska E, Boldis V, Kostanova Z, Kocianova E, Stefanidesova K. Incidence of various tick-borne microorganisms in rodents and ticks of central Slovakia. *Acta Virol*. 2008;52:175–9.
9. Rar VA, Fomenko NV, Dobrotvorsky AK, Livanova NN, Rudakova SA, Fedorov EG, et al. Tickborne pathogen detection, Western Siberia, Russia. *Emerg Infect Dis*. 2005;11:1708–15.
10. Dugan VG, Gaydos JK, Stallknecht DE, Little SE, Beall AD, Mead DG, et al. Detection of *Ehrlichia* spp. in raccoons (*Procyon lotor*) from Georgia. *Vector Borne Zoonotic Dis*. 2005;5:162–71. DOI: 10.1089/vbz.2005.5.162
11. Brouqui P, Sanogo YO, Caruso G, Merola F, Raoult D. '*Candidatus Ehrlichia walkeri*': a new *Ehrlichia* detected in *Ixodes ricinus* tick collected from asymptomatic humans in northern Italy. *Ann N Y Acad Sci*. 2003;990:134–40. DOI: 10.1111/j.1749-6632.2003.tb07352.x
12. Koutaro M, Santos AS, Dumler JS, Brouqui P. Distribution of '*Ehrlichia walkeri*' in *Ixodes ricinus* (Acari: Ixodidae) from the northern part of Italy. *J Med Entomol*. 2005;42:82–5. DOI: 10.1603/0022-2585(2005)042[0082:DOEWII]2.0.CO;2
13. Hengge UR, Tannappel A, Tyring SK, Erbel R, Arendt G, Ruzicka T. Lyme borreliosis. *Lancet Infect Dis*. 2003;3:489–500. DOI: 10.1016/S1473-3099(03)00722-9
14. Parola P, Davoust B, Raoult D. Tick- and flea-borne rickettsial emerging zoonoses. *Vet Res*. 2005;36:469–92. DOI: 10.1051/vetres:2005004

Address for correspondence: Guido V. Bloemberg, University of Zurich, Institute of Medical Microbiology, Gloriastrasse 32, 8006 Zurich, Switzerland; email: bloemberg@imm.uzh.ch

Septicemia Caused by Tick-borne Bacterial Pathogen *Candidatus Neoehrlichia mikurensis*

Technical Appendix

Supplementary Methods

Bacterial DNA Isolation

For the EDTA-blood mixture, the following procedure was applied: 1.5 mL of anticoagulated blood was mixed with 5 mL of EC lysis buffer (155 mmol/L NH₄Cl, 10 mmol/L KHCO₃, 2 mmol/L EDTA) in a 50-mL centrifugation tube and cooled on ice for 30 min. The tubes were inverted every 5 min and centrifuged at 16,000 × *g* for 15 min at 4°C. The supernatant was discarded, 5 mL EC lysis buffer was added, and the tube was centrifuged at 16,000 × *g* for 15 min. All but 1 mL of the supernatant was discarded; the pellet was gently resuspended and transferred into a 1.5-mL screw-cap tube (Eppendorf, Hamburg, Germany). The tube was centrifuged at 16,000 × *g* for 10 min. Subsequently, the supernatant was removed, and the pellet was mixed with 200 µL of digest buffer (50 mmol/L Tris-HCl pH 8.0, 1 mmol/L EDTA, 0.5% sodium dodecyl sulfate, 400 µg/mL proteinase K [recombinant, PCR-grade; Roche Diagnostics, Rotkreuz, Switzerland]). The suspension was vortexed until the pellet had completely dissolved and then incubated in a thermomixer at 55°C at 1,000 rpm for 2 h.

DNA from a uncoagulated (heparinized) blood sample was prepared as follows: buffy-coat blood was frozen and thawed; 1.5 mL of fluid was then transferred to a 50-mL centrifugation tube, together with 5 mL of EC lysis buffer. The sample was kept on ice for 30 min, and afterwards the tube was centrifuged at 16,000 × *g* for 10 min. Further steps were performed as mentioned for EDTA-blood samples.

To extract DNA from blood culture material, the following protocol was used: 500 µL of blood culture material was added to 1 mL of 50 mmol/L NaOH, 50 mmol/L sodium citrate in a 1.5-mL Eppendorf reaction tube and inverted gently for 10 min. After centrifugation at 16,000 ×

g for 5 min, the supernatant was disposed and the pellet was resuspended in 500 μ L 0.5 mol/L Tris-HCl (pH 8.0). Centrifugation was repeated, the supernatant was discarded, and again 500 μ L Tris-HCl solution was added and centrifugation was repeated. The supernatant was discarded, and the pellet was resuspended in 200 μ L of Tris-EDTA buffer. The solution was incubated in a thermomixer at 95°C at 600 rpm for 1 h. The tube was then centrifuged at 9,300 \times g for 15 min, and the DNA-containing supernatant was used for further purification steps. The DNA of all samples was finally purified and enriched by using the QIAamp DNA Blood Kit (QIAGEN, Hombrechtikon, Switzerland) with a slightly modified protocol: 200 μ L buffer AL was added, the mixture was heated to 70°C for 10 min. Then 200 μ L ethanol was added, and the solution was pipetted onto the purification column. The column was centrifuged at 6,000 \times g for 1 min. The column was transferred to a new collection tube and washed by using 500 μ L of buffer AW1, centrifuged for 1 min at 6,000 \times g, washed with 500 μ L of AW2, and centrifuged for 3 min at 16,000 \times g. Finally, DNA was eluted by using 50 μ L of buffer AE. The column was incubated for 5 min and then centrifuged at 6,000 \times g for 1 min.

Bacterial Broad-Range 16S rDNA and *groEL* PCR

Five microliters of the resulting DNA-containing eluate and 5 μ L of a 1:5 diluted eluate were used as template in a 50- μ L amplification reaction essentially as described (1). Five microliters of *Escherichia coli* chromosomal DNA (\approx 50 ng/ μ L) served as a positive control. Briefly, the PCR used 1.25 U low DNA AmpliTaq DNA polymerase (Applied Biosystems, Rotkreuz, Switzerland) in a reaction mixture containing 1 \times PCR buffer (10 mmol/L Tris-HCl pH 9.0, 50 mmol/L KCl, 1.5 mmol/L MgCl₂, 0.1% Triton X), 0.2 mmol/L dNTPs (Roche Diagnostics), and primers at a concentration of 0.1 mmol/L (BAK11w, 5'-AGT TTG ATC MTG GCT CAG-3'; and BAK2 5'-GGA CTA CHA GGG TAT CTA AT-3'). All reagents (e.g., *Limulus* amoebocyte lysate water) and buffers used were included as negative controls. The primary amplicons were visualized by polyacrylamide gel electrophoresis and silver staining and reamplified in a seminested PCR with primers BAK11w and BAK553r (5'-TTA CCG CGG CTG CTG GCA C-3'). Automated cycle sequencing was performed by using primer BAK11w on an ABI Prism 3100-Avant Genetic Analyzer (Applied Biosystems). *groEL* was amplified by using primers groEL_F 5'-TAT AGC TAA GGA AGC ATA GTC TA-3' and groEL_R 5'-AGC TCT

AGT AGC ATG TAA AGC-3' (this study). Sequencing was conducted with the amplification primers and additional internal *groEL* primers (1).

Reference

1. Takano A, Ando S, Kishimoto T, Fujita H, Kadosaka T, Nitta Y, et al. Presence of a novel *Ehrlichia* sp. in *Ixodes granulatus* found in Okinawa, Japan. *Microbiol Immunol.* 2009;53:101–6. [Medline](#)
[DOI: 10.1111/j.1348-0421.2008.00093.x](https://doi.org/10.1111/j.1348-0421.2008.00093.x)