RESEARCH LETTERS

leading hypothesis. However, *H. cinaedi* is an enterohepatic bacterium, and the patient had undergone surgery for an adhesive small intestinal obstruction 1 month before. Her medical and surgical history might have increased the intraintestinal pressure and induced the hepatic cyst infection through biliary reflux.

The patient had schizophrenia but was not immunocompromised beyond her surgical history. Many cases of *H. cinaedi* infection have been reported in immunocompromised hosts (2), but reports of *H. cinaedi* infections in immunocompetent patients have been increasing (3). Matsumoto et al. showed that *H. cinaedi* bacteremia was found in only 0.06% of total blood samples (4); none of the patients in their study were HIV-positive, but many were immunocompromised by other conditions. Kiehlbauch et al. also conducted a retrospective study of *H. cinaedi* bacteremia and found that 45% of patients were HIV-positive (5). *H. cinaedi* infection can occur regardless of a patient's immunologic or environmental status.

H. cinaedi infections are often reported in Japan. Miyake et al. reported that the *H. cinaedi* detection rate has increased after introduction of the BACTEC system (6). We also used BACTEC bottles. The widespread use of this type of blood culture bottle throughout Japan might contribute to the positivity rate of *H. cinaedi*.

We report a case of *H. cinaedi* hepatic cyst infection with bacteremia. *H. cinaedi* infection can occur in both nosocomial and community-acquired situations and in both immunocompromised and immunocompetent patients; its manifestations vary quite widely. Although the positivity rate of *H. cinaedi* is very low, it might still be overlooked. Further research is warranted to identify the epidemiologic and clinical features of *H. cinaedi* infection.

About the Author

Dr. Suzuki is a medical doctor at the National Center for Global Health and Medicine in Shinjuku-ku, Tokyo, Japan. His main research interest is nosocomial infectious diseases.

References

- Kawamura Y, Tomida J, Morita Y, Fujii S, Okamoto T, Akaike T. Clinical and bacteriological characteristics of *Helicobacter cinaedi* infection. J Infect Chemother. 2014;20:517–26. http://dx.doi.org/ 10.1016/j.jiac.2014.06.007
- Kamimura K, Kumaki D, Arita M, Kobayashi Y, Mizuno K, Kusama F, et al. First case of bacteremia caused by *Helicobacter cinaedi* in a patient with liver cirrhosis: a case report and literature review. Clin J Gastroenterol. 2015;8:306–17. http://dx.doi.org/10.1007/s12328-015-0600-0
- Uwamino Y, Muranaka K, Hase R, Otsuka Y, Hosokawa N. Clinical features of community-acquired *Helicobacter cinaedi* bacteremia. Helicobacter. 2016;21:24–8. http://dx.doi.org/10.1111/ hel.12236
- Matsumoto T, Goto M, Murakami H, Tanaka T, Nishiyama H, Ono E, et al. Multicenter study to evaluate bloodstream infection

by *Helicobacter cinaedi* in Japan. J Clin Microbiol. 2007;45:2853– 7. http://dx.doi.org/10.1128/JCM.00465-07

- Kiehlbauch JA, Tauxe RV, Baker CN, Wachsmuth IK. *Helicobacter cinaedi*–associated bacteremia and cellulitis in immunocompromised patients. Ann Intern Med. 1994;121:90–3. http://dx.doi.org/1 0.7326/0003-4819-121-2-199407150-00002
- Miyake N, Chong Y, Nishida R, Nagasaki Y, Kibe Y, Kiyosuke M, et al. A dramatic increase in the positive blood culture rates of *Helicobacter cinaedi*: the evidence of differential detection abilities between the Bactec and BacT/Alert systems. Diagn Microbiol Infect Dis. 2015;83:232–3. http://dx.doi.org/ 10.1016/j.diagmicrobio.2015.07.017

Address for correspondence: Satoshi Kutsuna, National Center for Global Health and Medicine, Disease Control and Prevention Center, 1-21-1, Toyama, Shinjuku-ku, Tokyo, 162-8655, Japan; email: sonare.since1192@gmail.com

Mycobacterium tuberculosis RD-Rio Strain in Kazakhstan

Yuriy Skiba, Igor Mokrousov, Dilyara Nabirova, Anna Vyazovaya, Elina Maltseva, Natalya Malakhova, Gulnara Ismagulova, Ilva Pole, Renate Ranka, Zhanar Sapiyeva, Shakhimurat Ismailov, Daphne Moffett

Author affiliations: Almaty Branch of National Center for Biotechnology at Central Reference Laboratory, Almaty, Kazakhstan (Y. Skiba); Aitkhozhin Institute of Molecular Biology and Biochemistry, Almaty (Y. Skiba, E. Maltseva, N. Malakhova, G. Ismagulova); St. Petersburg Pasteur Institute, St. Petersburg, Russia (I. Mokrousov, A. Vyazovaya); Centers for Disease Control and Prevention, Central Asia Regional Office, Almaty (D. Nabirova, D. Moffett); Latvian Biomedical Research and Study Centre, Riga, Latvia (I. Pole, R. Ranka); Ministry of Health, Almaty (Z. Sapiyeva, S. Ismailov); The Global Fund, Geneva, Switzerland (S. Ismailov)

DOI: https://doi.org/10.3201/eid2503.181179

Mycobacterium tuberculosis RD-Rio strains are still rare in the former Soviet Union countries and Asia. We describe a strain in Kazakhstan that belongs to the RD-Rio secondary branch, which is endemic to northwest Russia and eastern Europe. Although RD-Rio strains are frequently multidrug resistant, this heterogeneous branch included only drugsusceptible isolates. The RD-Rio strain of *Mycobacterium tuberculosis* was initially described in Rio de Janeiro, Brazil, and was demonstrated to be spread beyond South America (1). However, RD-Rio isolates are rare in northern Eurasia (i.e., in Baltic and former Soviet Union countries) (2). Phylogenetically, RD-Rio is part of the Latin-American-Mediterranean (LAM) genetic family and is marked by 2 large genomic deletions, RD174 and RD-Rio (3); RD-Rio is speculatively associated with particular pathogenic properties (1).

We describe a strain from Kazakhstan with confirmed RD-Rio deletion. Molecular analysis and comparison with the global LAM dataset showed that it belongs to the particular secondary branch described in the north of European Russia and Eastern Europe. Although RD-Rio isolates have been associated with multidrug resistance (MDR) (4), this branch on a dendrogram (Figure, panel A) included only drug-susceptible isolates.

We conducted this study as part of an ongoing molecular epidemiologic surveillance study of M. tuberculosis in Kazakhstan implemented in collaboration with the Centers for Disease Control and Prevention office in Kazakhstan (CDC-Central Asia Region Office). M. tuberculosis strain #127 was isolated in Almaty, Kazakhstan, in 2015 from a 52-year-old man with pulmonary tuberculosis (TB). He received anti-TB treatment at the TB hospital of the Interdistrict TB Dispensary in Almaty for 2 months and was discharged. We tested the strain for drug susceptibility to the first- and second-line drugs (streptomycin, isoniazid, rifampin, ethambutol, prothionamide, ofloxacin, kanamycin, capreomycin, and cycloserine). We tested DNA for drug-resistance mutations (in *rpoB*, *katG*, *inhA*, ahpC, embB, gyrA, gyrB, rrs, eis); spoligotyping; 24-locus variable-number tandem-repeat (VNTR) typing; detection of genome deletions RD174, RD-Rio, and RD115; and

LAM family–specific single-nucleotide polymorphisms (SNPs) in *Rv0129c* (Appendix, https://wwwnc.cdc.gov/ EID/article/25/3/18-1179-App.pdf). The strain was susceptible to all tested drugs and did not bear drug resistance mutations in the tested gene targets. It was assigned to the RD-Rio sublineage, spoligotype SIT20 (according to SIT-VIT2 database, http://www.pasteur-guadeloupe.fr:8081/ SITVIT2/), and Mlva15-9 code #16113-830 (according to https://www.MIRU-VNTRplus.org). We conducted phylogenetic analysis on the 24-MIRU-VNTR profile of this strain along with 357 isolates of the global LAM dataset (*5*) (Figure; Appendix Figure 1, 2).

A recent global LAM study demonstrated that SIT20 is one of the major RD-Rio spoligotypes and is subdivided into 2 branches on the basis of the ETRB locus alleles (5). In this study, we showed that, on the global LAM tree, the strain from Kazakhstan clustered within the branch that included only drug-susceptible isolates from northwestern Russia and Latvia with SIT20 and derived SIT1321 spoligoprofiles (Figure, panel B; Appendix Figure 1).

The case-patient's medical record contained no information about his contacts and travel before hospitalization. As a man of working age, he could have traveled to Russia as a migrant worker. It has been estimated that \approx 1.9 million Kazakhstan citizens lived in Russia dur-1989-2007 (http://focus-migration.hwwi.de/Rusing sian-Federation.6337.0.html?&L=1). In 2015, a total of 2,560,000 persons, including seasonal labor migrants (62% of all outgoing migrants), were known to have migrated from Kazakhstan to Russia (https://www.iom.int/ world-migration). Russian law requires that migrants with TB be deported, which may explain why this case-patient preferred to disappear or remain unavailable. Thus, a hypothesis about Russian origin of this strain is based solely on the fact that Kazakhstan is the most common country



Figure. Mycobacterium tuberculosis RD-Rio strain in Kazakhstan. A) Section of the variable-number tandem-repeatbased dendrogram of the Latin-American-Mediterranean family of Mycobacterium tuberculosis RD-Rio strain with enlarged branch including SIT20 strain from Kazakhstan. All isolates were drug susceptible. The complete dendrogram with VNTR profiles is provided in Appendix Figures 1, 2 (https://wwwnc.cdc.gov/EID/ article/25/3/18-1179-App.pdf). B) Binary spoligoprofiles of the studied strains.

Emerging Infectious Diseases • www.cdc.gov/eid • Vol. 25, No. 3, March 2019

RESEARCH LETTERS

of origin of immigrants to Russia. However, phylogenetic analysis based on high-resolution VNTR loci placed this strain within the branch exclusively made of the isolates from different and neighboring regions in northwestern Russia and Latvia (Figure). We consider this clustering to be evidence that this strain is related to the *M. tuberculosis* population in the European part of Russia. Another example of cross-country *M. tuberculosis* transmission is the "successful Russian strain" Beijing B0/W148-cluster; its overall prevalence in Kazakhstan is low, at 3%, and its isolates were identified in the northern part of the country that is close to Russia (6).

Previously, strains of the LAM RD-Rio or SIT20 spoligotype were not described in several countrywide studies in Kazakhstan during 1997–2014 (2,6–8). In neighboring Kyrgyzstan, SIT20 was not described either in the civilian or penitentiary settings (9,10). In view of the rarity of RD-Rio isolates in northern Eurasia and their previous absence in Kazakhstan, the isolation of such a strain in Kazakhstan, especially in the most distant southern region, deserves attention. That no other isolates have been found through our ongoing surveillance strongly suggests the strain was imported and not acquired in Kazakhstan. The isolates in this SIT20 Russian branch were sufficiently heterogeneous in terms of VNTR locus diversity; they were isolated in different years, and all isolates were drug susceptible (Figure). RD-Rio is known to be associated with MDR, and even by chance, some of these isolates in former Soviet Union countries could have acquired drug resistance under the current adverse conditions of TB control in this region. Nevertheless, these strains have remained drug susceptible. Further surveillance will be needed to determine if additional strains appear and, if so, whether they remain drug susceptible or acquire drug resistance.

Acknowledgments

Funding for Y.S. is through Ministry of Education and Science of Republic of Kazakhstan grant 3732/GF4, and Ministry of Health of Republic of Kazakhstan program 49019/PCF-MZSR-OT-18. Funding for I.M. and A.V. is through Russian Science Foundation grant 14-14-00292. Funding for I.P. and R.R. is through Latvian National Research program VPP 5.7 "Biomedicine."

About the Author

Dr. Skiba is Group Leader in the Genome Laboratory at Aitkhozhin Institute of Molecular Biology and Biochemistry, and Head of Laboratory of Molecular Biology, Almaty Branch, National Center for Biotechnology in the Central Reference Laboratory, Almaty, Kazakhstan. His research interests include molecular epidemiology of tuberculosis, nosocomial tuberculosis, and database development.

References

- Gibson AL, Huard RC, Gey van Pittius NC, Lazzarini LC, Driscoll J, Kurepina N, et al. Application of sensitive and specific molecular methods to uncover global dissemination of the major RD-Rio sublineage of the Latin American–Mediterranean *Mycobacterium tuberculosis* spoligotype family. J Clin Microbiol. 2008;46:1259–67. http://dx.doi.org/10.1128/JCM.02231-07
- Mokrousov I, Vyazovaya A, Narvskaya O. *Mycobacterium* tuberculosis Latin American–Mediterranean family and its sublineages in the light of robust evolutionary markers. J Bacteriol. 2014;196:1833–41. http://dx.doi.org/10.1128/JB.01485-13
- Lazzarini LC, Huard RC, Boechat NL, Gomes HM, Oelemann MC, Kurepina N, et al. Discovery of a novel *Mycobacterium tuberculosis* lineage that is a major cause of tuberculosis in Rio de Janeiro, Brazil. J Clin Microbiol. 2007;45:3891–902. http://dx.doi.org/10.1128/JCM.01394-07
- Dalla Costa ER, Lazzarini LC, Perizzolo PF, Díaz CA, Spies FS, Costa LL, et al. *Mycobacterium tuberculosis* of the RD-Rio genotype is the predominant cause of tuberculosis and associated with multidrug resistance in Porto Alegre City, South Brazil. J Clin Microbiol. 2013;51:1071–7. http://dx.doi.org/10.1128/ JCM.01511-12
- Mokrousov I, Vyazovaya A, Iwamoto T, Skiba Y, Pole I, Zhdanova S, et al. Latin American–Mediterranean lineage of *Mycobacterium tuberculosis*: human traces across pathogen's phylogeography. Mol Phylogenet Evol. 2016;99:133–43. http://dx.doi.org/10.1016/j.ympev.2016.03.020
- Skiba Y, Mokrousov I, Ismagulova G, Maltseva E, Yurkevich N, Bismilda V, et al. Molecular snapshot of *Mycobacterium tuberculosis* population in Kazakhstan: a country-wide study. Tuberculosis (Edinb). 2015;95:538–46. http://dx.doi.org/10.1016/ j.tube.2015.04.012
- Kubica T, Agzamova R, Wright A, Aziz MA, Rakishev G, Bismilda V, et al. The Beijing genotype is a major cause of drug-resistant tuberculosis in Kazakhstan. Int J Tuberc Lung Dis. 2005;9:646–53.
- Ibrayeva A, Kozhamkulov U, Raiymbek D, Alenova A, Igilikova S, Zholdybayeva E, et al. Molecular epidemiology of *Mycobacterium tuberculosis* strains circulating in the penitentiary system of Kazakhstan [short communication]. Int J Tuberc Lung Dis. 2014;18:298–301. http://dx.doi.org/10.5588/ijtld.13.0558
- Mokrousov I, Valcheva V, Sovhozova N, Aldashev A, Rastogi N, Isakova J. Penitentiary population of *Mycobacterium tuberculosis* in Kyrgyzstan: exceptionally high prevalence of the Beijing genotype and its Russia-specific subtype. Infect Genet Evol. 2009;9:1400–5. http://dx.doi.org/10.1016/j.meegid.2009.07.007
- Mokrousov I, Isakova J, Valcheva V, Aldashev A, Rastogi N. Molecular snapshot of *Mycobacterium tuberculosis* population structure and drug resistance in Kyrgyzstan. Tuberculosis (Edinb). 2013;93:501–7. http://dx.doi.org/10.1016/j.tube.2013.05.008

Address for correspondence: Igor Mokrousov, St. Petersburg Pasteur Institute, 14 Mira str, St. Petersburg 197101, Russia; email: imokrousov@mail.ru; Yuriy Skiba, Aitkhozhin Institute of Molecular Biology and Biochemistry, 86 Dosmuhamedova str, Almaty, 480012, Kazakhstan; email: yuriy.skiba@gmail.com

Mycobacterium tuberculosis RD-Rio Strain in Kazakhstan

Appendix



Appendix Figure 1. *Mycobacterium tuberculosis* RD-Rio strain in Kazakhstan. A) Section of the variablenumber tandem-repeat–based dendrogram of the Latin-American-Mediterranean family of *Mycobacterium tuberculosis* RD-Rio strain with enlarged branch including SIT20 strain from Kazakhstan. All isolates were drug-susceptible. B) Binary spoligoprofiles of the studied strains. C) Locations of isolation of the studied strains.



Appendix Figure 2. The UPGMA dendrogram of 259 VNTR types (357 LAM isolates) (Mokrousov et al., 2016) with SIT20 strain from Kazakhstan. Asterisks indicate RD-Rio sublineage and its branch with SIT20 strain from Kazakhstan ("SIT20 Russian branch"). The column "Type, number and origin" refers to the LAM types in Mokrousov et al. (2016). Mlva15–9 refers to nomenclature of MIRU-VNTRplus.org. Twenty-four VNTR loci are listed in the order in which they are found on chromosome, clockwise: 424, 577, 580, 802, 960, 1644, 1955, 2163b, 2165, 2401, 2996, 3192, 3690, 4052, 4156, 154, 2059, 2347, 2461, 2531, 2687, 3007, 3171, 4348.

Supplementary References

- Allix-Béguec C, Harmsen D, Weniger T, Supply P, Niemann S. Evaluation and strategy for use of MIRU-VNTRplus, a multifunctional database for online analysis of genotyping data and phylogenetic identification of *Mycobacterium tuberculosis* complex isolates. J Clin Microbiol. 2008;46:2692–9. <u>PubMed http://dx.doi.org/10.1128/JCM.00540-08</u>
- Demay C, Liens B, Burguière T, Hill V, Couvin D, Millet J, et al. SITVITWEB—a publicly available international multimarker database for studying *Mycobacterium tuberculosis* genetic diversity and molecular epidemiology. Infect Genet Evol. 2012;12:755–66. <u>PubMed</u> <u>http://dx.doi.org/10.1016/j.meegid.2012.02.004</u>
- Gibson AL, Huard RC, Gey van Pittius NC, Lazzarini LC, Driscoll J, Kurepina N, et al. Application of sensitive and specific molecular methods to uncover global dissemination of the major RDRio sublineage of the Latin American–Mediterranean *Mycobacterium tuberculosis* spoligotype family. J Clin Microbiol. 2008;46:1259–67. <u>PubMed http://dx.doi.org/10.1128/JCM.02231-07</u>
- Kamerbeek J, Schouls L, Kolk A, van Agterveld M, van Soolingen D, Kuijper S, et al. Simultaneous detection and strain differentiation of *Mycobacterium tuberculosis* for diagnosis and epidemiology. J Clin Microbiol. 1997;35:907–14. <u>PubMed</u>
- 5. Lazzarini LC, Huard RC, Boechat NL, Gomes HM, Oelemann MC, Kurepina N, et al. Discovery of a novel *Mycobacterium tuberculosis* lineage that is a major cause of tuberculosis in Rio de Janeiro, Brazil. J Clin Microbiol. 2007;45:3891–902. <u>PubMed http://dx.doi.org/10.1128/JCM.01394-07</u>
- 6. Mokrousov I, Rastogi N. Spacer-based macroarrays for CRISPR genotyping. Methods Mol Biol. 2015;1311:111–31. PubMed http://dx.doi.org/10.1007/978-1-4939-2687-9_7
- Mokrousov I, Vyazovaya A, Narvskaya O. *Mycobacterium tuberculosis* Latin American– Mediterranean family and its sublineages in the light of robust evolutionary markers. J Bacteriol. 2014;196:1833–41. <u>PubMed http://dx.doi.org/10.1128/JB.01485-13</u>
- Mokrousov I, Vyazovaya A, Iwamoto T, Skiba Y, Pole I, Zhdanova S, et al. Latin American– Mediterranean lineage of *Mycobacterium tuberculosis*: human traces across pathogen's phylogeography. Mol Phylogenet Evol. 2016;99:133–43. <u>PubMed</u> <u>http://dx.doi.org/10.1016/j.ympev.2016.03.020</u>
- 9. Rüsch-Gerdes S, Pfyffer GE, Casal M, Chadwick M, Siddiqi S. Multicenter laboratory validation of the BACTEC MGIT 960 technique for testing susceptibilities of *Mycobacterium tuberculosis* to

classical second-line drugs and newer antimicrobials. J Clin Microbiol. 2006;44:688–92. PubMed http://dx.doi.org/10.1128/JCM.44.3.688-692.2006

- 10. Supply P, Allix C, Lesjean S, Cardoso-Oelemann M, Rüsch-Gerdes S, Willery E, et al. Proposal for standardization of optimized mycobacterial interspersed repetitive unit-variable-number tandem repeat typing of *Mycobacterium tuberculosis*. J Clin Microbiol. 2006;44:4498–510. <u>PubMed</u> http://dx.doi.org/10.1128/JCM.01392-06
- 11. World Health Organization. Policy guidance on drug-susceptibility testing (DST) of second-line antituberculosis drugs. WHO/HTM/TB/2008.392. Geneva: The Organization; 2008.

<jrn>12. Zimenkov DV, Kulagina EV, Antonova OV, Zhuravlev VY, Gryadunov DA. Simultaneous

drug resistance detection and genotyping of Mycobacterium tuberculosis using a low-density hydrogel

microarray. J Antimicrob Chemother. 2016;71:1520-31. PubMed

http://dx.doi.org/10.1093/jac/dkw015</jrn>