
Highly Pathogenic Avian Influenza A(H5N8) Virus, Cameroon, 2017

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Highly pathogenic avian influenza A(H5N8) viruses of clade 2.3.4.4 spread into West Africa in late 2016 during the autumn bird migration. Genetic characterization of the complete genome of these viruses detected in wild and domestic birds in Cameroon in January 2017 demonstrated the occurrence of multiple virus introductions.

Since the first detection in China in 1996, highly pathogenic avian influenza (HPAI) viruses of the H5 subtype descendent of the H5N1 virus A/goose/Guangdong/1/1996 (Gs/GD/96) have evolved into multiple genetic clades (1, 2) that have been threatening poultry worldwide. Since 2010, clade 2.3.4 has demonstrated an unusual propensity to replace its N1 subtype and acquire different neuraminidase (NA) genes from unrelated avian influenza viruses through reassortment, which has resulted in the emergence of new viral subtypes within the Gs/GD/96 H5 lineage (e.g., N2, N5, N6, N8). In late spring 2016, reassortant HPAIA(H5N8) clade 2.3.4.4 (group B) virus was detected in migratory wild birds in Qinghai Lake, China (3), and in the salt lake system of Uvs Nuur, on the border between Mongolia and the Russian Federation (4). Since then, this newly emerged virus has caused multiple outbreaks of the disease in poultry and wild birds across Europe, Asia, and the Middle

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East and has extended not only to countries in northern, central, and western Africa, as did the previous Gs/GD/96 H5 lineage outbreaks, but also to the eastern and southern parts of Africa (5). We investigated the epidemiology of the outbreaks in early 2017 of HPAI H5N8 virus in domestic and wild birds in Cameroon and determined the possible origin of this virus through whole-genome analyses.

The Study

On January 2, 2017, high death rates were reported in Indian peafowl (*Pavo cristatus*) in a backyard exotic poultry farm located in the town of Makilingaye (Tokombéré district of Mayo-Sava division), a village neighboring Nigeria in the far-north region of Cameroon. Samples were collected by the Cameroon Epidemio-Surveillance Network (Reseau d'Epidemio-Surveillance au Cameroun [RESCAM]) of the Ministry of Livestock, Fisheries and Animal Industries and were sent to the National Veterinary Laboratory (LANAVET) in Garoua, where the H5N8 subtype was diagnosed. Almost all the peafowl (103/107) died within ≈2 weeks. Death in chickens (*Gallus gallus domesticus*, 24/24) housed in the same compound was delayed and appeared 5 days later than in peafowl. Following the laboratory confirmation of the first outbreak, the Cameroon government, through the Ministry of Livestock, Fisheries, and Animal Industries, implemented prompt and strong control measures to stop the spread of the virus and reduce the risk of human infections. Stamping out was deployed together with movement restrictions and virological surveillance; disinfection of premises and contact materials was intensified. The RESCAM team conducted a routine avian influenza survey in the Maroua, Yagoua, and Guidiguis central poultry markets in the far-north region. All the samples were analyzed at LANAVET Garoua and Annex Yaounde; H5N8 virus was detected in 5 birds (1 pigeon, 1 chicken, 2 guinea fowls, and 1 duck) out of 122 birds.

We sequenced the hemagglutinin (HA) and NA gene segments of the virus A/Indian peafowl/Cameroon/17RS1661-6/2017, identified from an Indian peafowl in Makilingaye, at the Istituto Zooprofilattico Sperimentale delle Venezie in Legnaro, Italy, along with the complete genomes of 2 positive samples collected from domestic birds (chicken and duck) and the partial genome (all segments except for polymerase acidic protein [PA] and polymerase basic protein 1 [PB1]) of a sample from

Table. Epidemiologic information of viruses characterized in study of highly pathogenic avian influenza A(H5N8) virus, Cameroon, January 2017*

Virus	Sample type	Location	EpiFlu accession numbers for 8 gene segments
A/chicken/Cameroon/17RS1661-1/2017	Tracheal swab	Maroua market	HA, MG650619; MP, MG650622; NA, MG650626; NP, MG650630; NS, MG650632; PA, MG650635; PB1, MG650638; PB2, MG650641
A/duck/Cameroon/17RS1661-3/2017	Tracheal swab	Yagoua market	HA, MG650620; MP, MG650623; NA, MG650627; NP, MG650629; NS, MG650633; PA, MG650636; PB1, MG650637; PB2, MG650639
A/pigeon/Cameroon/17RS1661-4/2017	Cloacal swab	Maroua market	HA, MG650621; MP, MG650624; NA, MG650628; NP, MG650631; NS, MG650634; PA, NR; PB1, na; PB2, MG650640
A/Indian peafowl/Cameroon/17RS1661-6/2017	Tracheal swab	Makilingaye	HA, MG650619; MP, NR; NA, MG650626; NP, NR; NS, NR; PA, NR; PB1, NR; PB2, NR

*HA, hemagglutinin; MP, matrix protein, NA, neuraminidase; NR, not reported; NP, nucleoprotein; NS, nonstructural proteins; PA, polymerase acidic protein; PB1, polymerase basic protein 1; PB2, polymerase basic protein 2.

†From GISAID EpiFlu database (online Technical Appendix 2, <https://wwwnc.cdc.gov/EID/article/24/7/17-2120-Techapp2.xlsx>).

a wild pigeon, all identified in the Maroua and Yagoua markets (Table). A detailed description of the methods used for sequencing and genetic analyses is provided in online Technical Appendix 1 (<https://wwwnc.cdc.gov/EID/article/24/7/17-2120-Techapp1.pdf>) and details on the HA gene segments used for the analysis are given in

Technical Appendix 2 (<https://wwwnc.cdc.gov/EID/article/24/7/17-2120-Techapp2.xlsx>). We submitted consensus sequences to GenBank (accession nos. MG650618–41).

Topology of the phylogenetic tree based on the HA gene segment showed that the 4 H5N8 viruses detected in Cameroon in 2017 fell within genetic clade 2.3.4.4 group

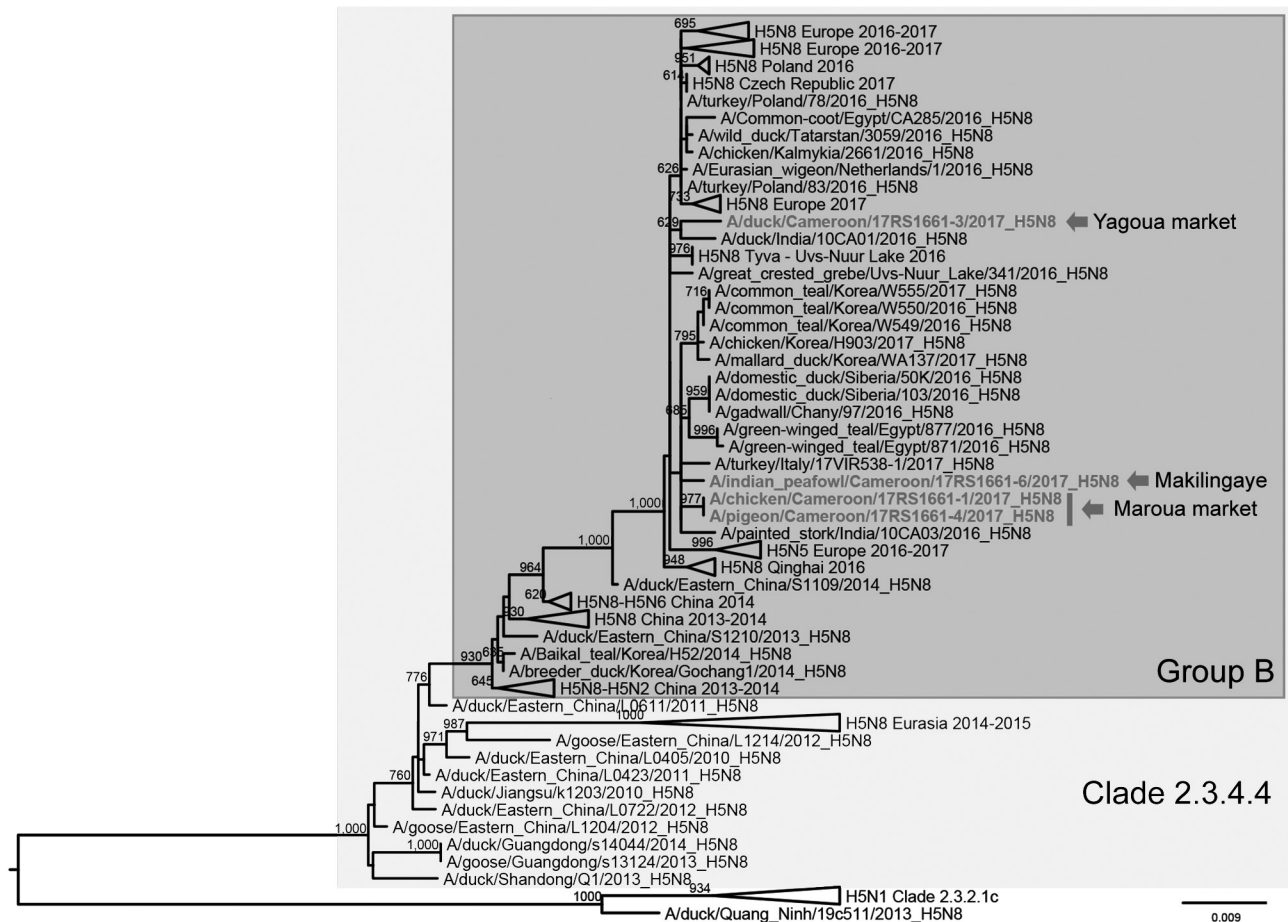


Figure. Maximum-likelihood phylogenetic tree of the HA gene of highly pathogenic avian influenza A(H5N8) virus from Cameroon (bold) and reference viruses. Arrows indicate location where Cameroon viruses were obtained. Bootstrap supports >600/1,000 are indicated next to the nodes. Scale bar indicates numbers of nucleotide substitutions per site.

B (Figure; online Technical Appendix 1 Figure 1). The HA gene segments of the 3 viruses from Makilingaye (A/Indian peafowl/Cameroon/17RS1661-6/2017) and Maroua market clustered together; similarity ranged from 99.5% (between the viruses collected from Makilingaye and Maroua markets) to 100% (between the 2 viruses from Maroua market). These segments grouped with H5N8 viruses collected during 2016–2017 in South Korea, Egypt, Italy, Siberia, and India (similarity between 99.3% and 99.6%). However, the virus A/duck/Cameroon/17RS1661-3/2017 identified in Yagoua market clustered separately and showed the highest similarity (99.3%) to a virus from India (A/duck/India/10CA01/2016) (Figure).

We confirmed the different clustering of the viruses collected from distinct markets in Cameroon by the analyses of the other gene segments, which suggests the occurrence of ≥ 2 distinct viral introductions in the country. Specifically, sequences of the 2 viruses from Maroua market were identical for all the available gene segments (Table) and clustered with viruses collected in Asia, Europe, and Egypt (online Technical Appendix 1 Figures 2–8). On the other hand, the virus A/duck/Cameroon/17RS1661-3/2017 grouped with the A/duck/India/10CA01/2016 virus (similarity 94.3–99.9%), except for the nucleoprotein (NP) gene, where it showed the highest identity (99.3%) with a virus collected in 2016 in the Russian Federation (Uvs Nuur Lake). The lack of genetic information on other H5N8 viruses detected in Africa makes it impossible to pinpoint how these viruses entered Cameroon and spread, nor is it feasible to determine where the 2 introductions occurred. The time to the most recent common ancestor estimated for the HA gene (online Technical Appendix 1) suggested that 2 introductions into Africa may have occurred almost simultaneously during March–December 2016 (online Technical Appendix 1 Figure 9). However, analyses of the amino acid sequences show that A/duck/Cameroon/17RS1661-3/2017 possesses the N319K mutation in the NP protein, which has been reported to enhance polymerase activity and stimulate vRNA synthesis in mammalian cells (6). In addition, A/chicken/Cameroon/17RS1661-1/2017 contains the V100A mutation in the PA protein, which is an amino acid signature typical of human influenza viruses (7).

Conclusions

Nigeria was the first country in West Africa to report the presence of the HPAI H5N8 virus in November 2016 (8,9). Less than 2 months later, in January 2017, the virus was detected in the far-north region of Cameroon in Makilingaye, close to the Nigerian border. Considering the extensive and porous frontier between Cameroon and Nigeria, trade and movement of poultry might have played a key role in the spread of the virus. However, the involvement of wild birds cannot be excluded. The region contains several wetlands

(Domayo River, Mayo Kani River, and Maga Lake) where different wild birds congregate, in particular during the dry season (December–April), when the virus was first detected. The almost simultaneous detection (early January 2017) of the H5N8 virus in poultry and wild birds in distant locations in Africa, such as Tunisia and Uganda, suggests that the role of wild birds in the introduction and/or dissemination of the virus in the region should not be overlooked. Of note, the first outbreak caused by the Gs/GD/96 H5 lineage in Cameroon, in 2006, also occurred in the far-north region of Cameroon at about the same time. This finding might suggest a common pathway for introduction of the virus into this area and highlights the need to improve surveillance in this region.

Although the epidemiologic and genetic data are insufficient to establish definite pathways and time of introduction of H5N8 virus into West Africa, this study demonstrates that ≥ 2 distinct H5N8 viruses entered Cameroon. This finding, together with the evidence that this event represents the third incursion of a Gs/GD/96-lineage H5 HPAI virus into Cameroon, again underlines the need to perform avian influenza surveillance on an ongoing basis for rapid identification and response to outbreaks in this area.

Acknowledgments

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About the Author

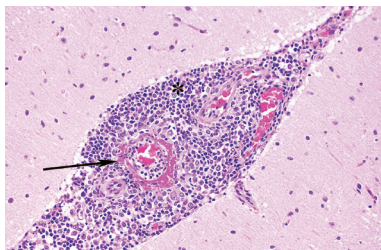
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December 2015: Zoonotic Infections



- Identifying and Reducing Remaining Stocks of Rinderpest Virus
- Opportunistic Pulmonary *Bordetella hinzii* Infection after Avian Exposure
- Zoonotic Leprosy in the Southeastern United States
- Infection Risk for Persons Exposed to Highly Pathogenic Avian Influenza A H5 Virus–Infected Birds, United States, December 2014–2015
- High Prevalence of Intermediate *Leptospira* spp. DNA in Febrile Humans from Urban and Rural Ecuador

- Biological Warfare Plan in the 17th Century—the Siege of Candia, 1648–1669
- Influenza A(H6N1) Virus in Dogs, Taiwan
- Methicillin-Resistant *Staphylococcus aureus* Prevalence among Captive Chimpanzees, Texas, USA, 2012



- Novel *Waddlia* Intracellular Bacterium in *Artibeus intermedius* Fruit Bats, Mexico
- Tembusu-Related Flavivirus in Ducks, Thailand
- Japanese Macaques (*Macaca fuscata*) as Natural Reservoir of *Bartonella quintana*

- *Onchocerca lupi* Nematode in a Cat, Europe
- Increased Number of Human Cases of Influenza Virus A(H5N1) Infection, Egypt, 2014–15
- Replication Capacity of Avian Influenza A(H9N2) Virus in Pet Birds, Chickens, and Mammals, Bangladesh
- Hendra Virus Infection in Dog, Australia, 2013
- No Evidence of Gouléako and Herbert Virus Infections in Pigs, Côte d'Ivoire and Ghana
- Aquatic Bird Bornavirus 1 in Wild Geese, Denmark
- Vectorborne Transmission of *Leishmania infantum* from Hounds, United States



Highly Pathogenic Avian Influenza A(H5N8) Virus, Cameroon, 2017

Technical Appendix

Materials and Methods

Sample Collection and Laboratory Analysis

The Cameroon Epidemio-Surveillance Network (RESCAM) of the Ministry of Livestock, Fisheries and Animal Industries (MINEPIA) collected 1 dead peacock during the outbreak in Makilingaye township and sent it to the National Veterinary Laboratory (LANAVET) in Garoua for analysis. In addition, we collected tracheal and cloacal swabs from 50 chickens, 55 ducks, 11 guinea fowls, and 6 pigeons during avian influenza field investigations in Maroua, Yagoua, and Guidiguis' central poultry markets (Technical Appendix Table 1). We sent these samples to the same laboratory for analysis. We extracted viral RNA from all the samples using the QIamp RNA mini kit (QIAGEN, Hilden, Germany) according to the manufacturer's protocol. We conducted RNA detection of the H5 gene using real-time reverse transcription PCR (RT-PCR) (1) followed by RNA detection of the N8 gene (2).

Genome Sequencing

At the World Organization for Animal Health (OIE)/Food and Agriculture Organization of the United Nations (UN-FAO) Reference Laboratory for Avian Influenza (Istituto Zooprofilattico Sperimentale delle Venezie, Legnaro, PD) in Italy, total RNA was purified from 4 HPAI H5N8-positive clinical samples using the Nucleospin RNA kit (Macherey–Nagel, Düren, Germany). Complete influenza A virus genomes were amplified with the SuperScript III One-Step RT-PCR system with Platinum Taq High Fidelity (Invitrogen, Carlsbad, CA, USA) using 1 pair of primers complementary to the conserved elements of the influenza A virus promoter (3). Sequencing libraries were obtained using a Nextera DNA XT Sample preparation kit (Illumina, San Diego, CA, USA) following the manufacturer's instructions and quantified using the Qubit dsDNA High Sensitivity kit (Invitrogen). We determined the average fragment

length using the Agilent High Sensitivity Bioanalyzer Kit (Agilent Technologies, Santa Clara, CA, USA). The indexed libraries were pooled in equimolar concentrations and sequenced in multiplex for 250 bp paired-end on Illumina MiSeq, according to the manufacturer's instructions.

High-Throughput Sequencing Data Analysis

We assessed the Illumina reads' quality using FastQC v0.11.2 (<https://www.bioinformatics.babraham.ac.uk/projects/fastqc>). We filtered the raw data by removing reads with more than 10% of undetermined (N) bases, reads with more than 100 bases with Q score below 7, and duplicated paired-end reads. We clipped the remaining reads from Illumina Nextera XT adaptors with scythe v0.991 (<https://github.com/vsbuffalo/scythe>) and trimmed them with sickle v1.33 (<https://github.com/najoshi/sickle>). We discarded reads shorter than 80 bases or unpaired after previous filters. We then aligned the high-quality reads against a reference genome using BWA v0.7.12 (4). We processed the alignments with Picard-tools v2.1.0 (<http://picard.sourceforge.net>) and GATK v3.5 (5,6,7) to correct potential errors, realign reads around indels, and recalibrate base quality. We called single nucleotide polymorphisms using LoFreq v2.1.2 (8) and used the outputs to generate the consensus sequences. We obtained the complete genome of viruses A/chicken/Cameroon/17RS1661-1/2017 and A/duck/Cameroon/17RS1661-3/2017; the HA, NA, MA, NP, NS, and PB2 gene segments of A/pigeon/Cameroon/17RS1661-4/2017; and the HA and NA gene segments of the virus A/Indian_peafowl/Cameroon/17RS1661-6/2017.

Phylogenetic Analyses

We compared the sequences of the 8 gene segments of the H5N8 viruses from Cameroon with all the H5 sequences belonging to clade 2.3.4.4 group B available in GISAD and with the 50 most related sequences resulted from the BLAST search. We aligned sequences of each gene segment using MAFFT v. 7 (9). We obtained maximum likelihood phylogenetic trees for each gene segment using the best-fit general time-reversible model of nucleotide substitution with gamma-distributed rate variation among sites (with 4 rate categories, Γ_4) and a heuristic SPR branch-swapping search (10) available in the PhyML program v3.1. To assess the robustness of individual nodes of the phylogeny, we performed 1,000 bootstrap replicates. Phylogenetic trees were visualized with the program FigTree v1.4.2 (<http://tree.bio.ed.ac.uk/software/figtree/>).

Estimation of Time to the Most Recent Common Ancestor

We estimated the tMRCAs of the HPAI H5N8 identified in Cameroon for the HA gene segment using BEAST v1.8.4 software (11). We selected a HKY85 + Γ_4 nucleotide substitution model with two data partitions reflecting codon positions (1st + 2nd positions, 3rd position) and with base frequency unlinked across all codon positions (SRD06 substitution model). In addition, we used a relaxed (uncorrelated lognormal) molecular clock and constant population size coalescent as the tree prior. We used Markov chain Monte Carlo (MCMC) and chain lengths of 100 million iterations to achieve convergence as assessed using Tracer v1.6 (<http://beast.bio.ed.ac.uk/Tracer>). The maximum clade credibility (MCC) phylogenetic tree was summarized from the posterior distribution of trees using TreeAnnotator v1.8.4 (11) after the removal of an appropriate burn-in (10% of the samples). The MCC tree was visualized using FigTree v1.4.2 (<http://tree.bio.ed.ac.uk/software/figtree/>) (Technical Appendix Figure 9).

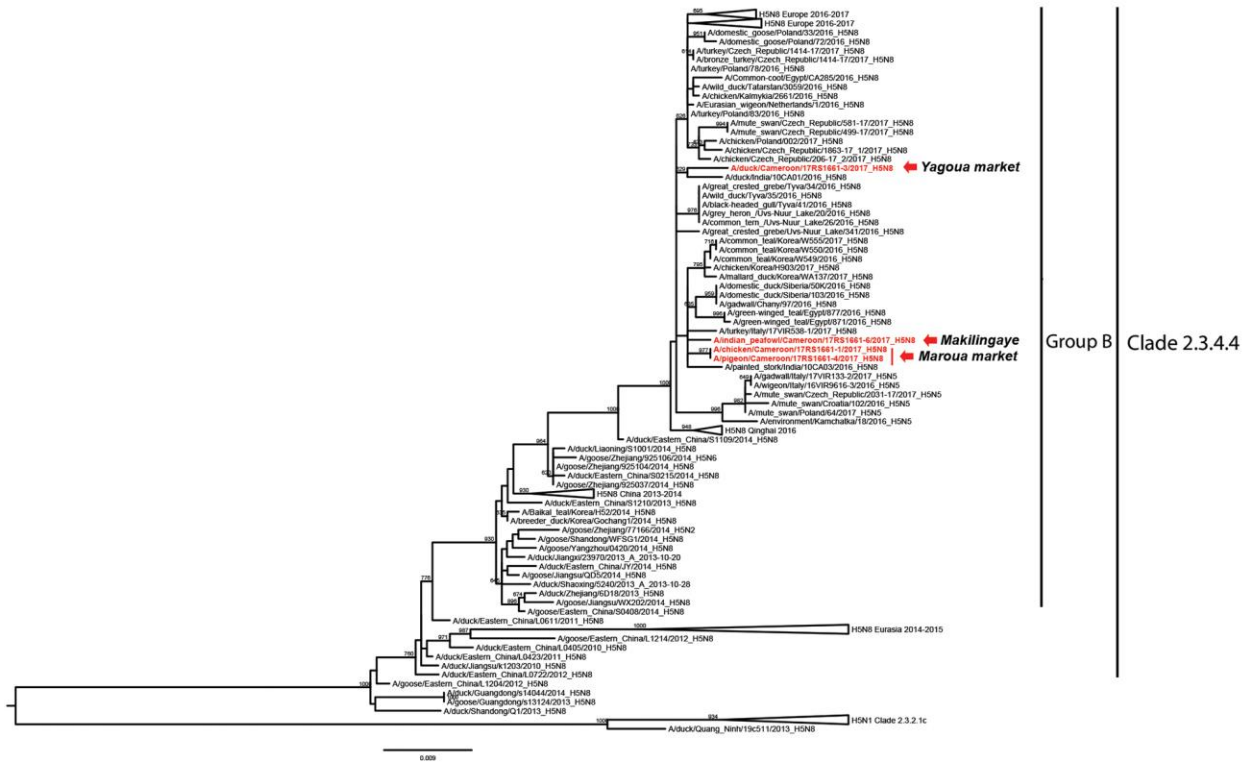
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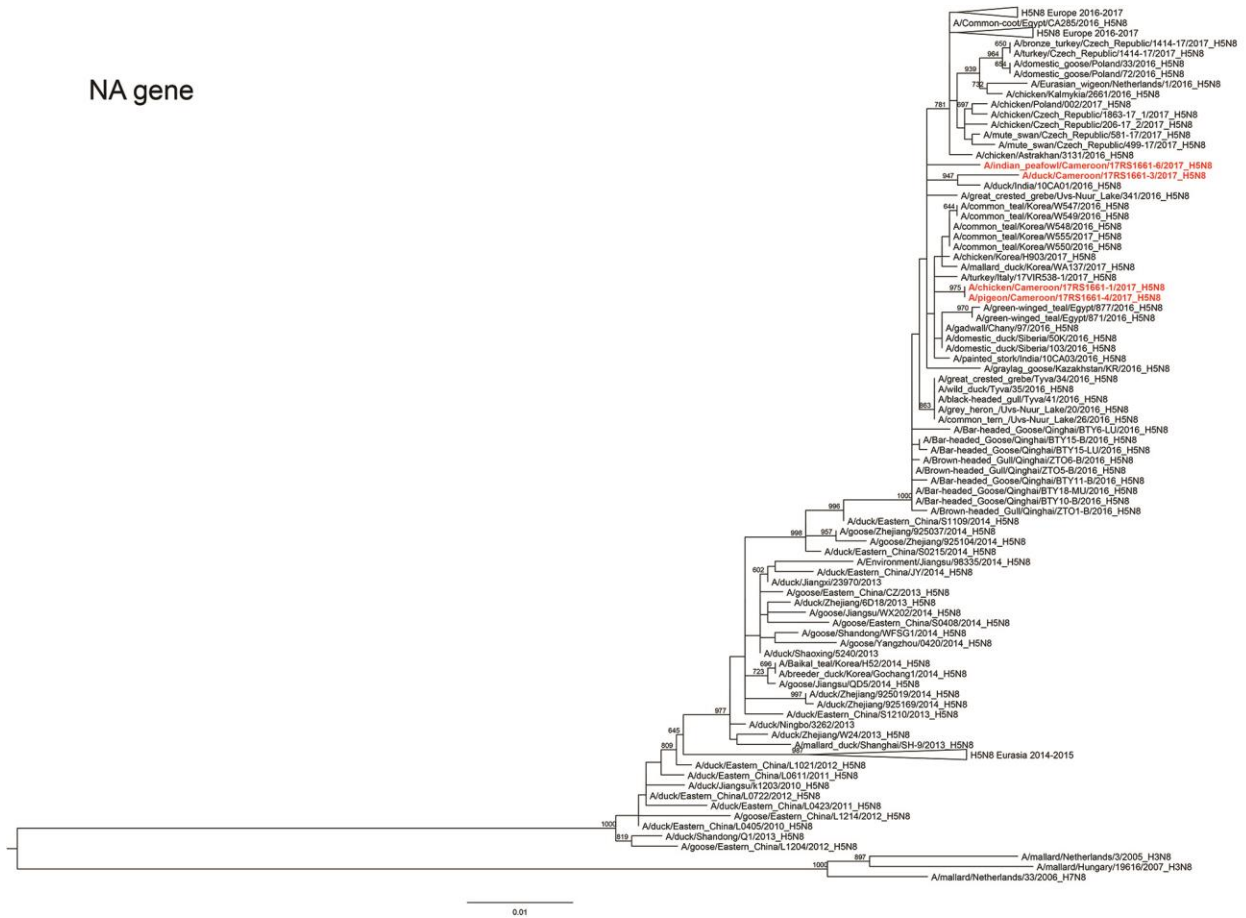
Technical Appendix 1 Table. Number of positive samples in the total number of sampled animals (no. pos/no. samples) from each market

Animal	Maroua market no. pos/no. samples	Yagoua market no. pos/no. samples	Guidiguis market no. pos/no. samples	Total no. pos/no. samples
Chicken	1/13	0/26	0/11	1/50
Pigeon	1/4	0/1	0/1	1/6
Guinea fowl	0/4	2/4	0/3	2/11
Duck	0/16	1/24	0/15	1/55
Total	2/37	3/55	0/30	5/122



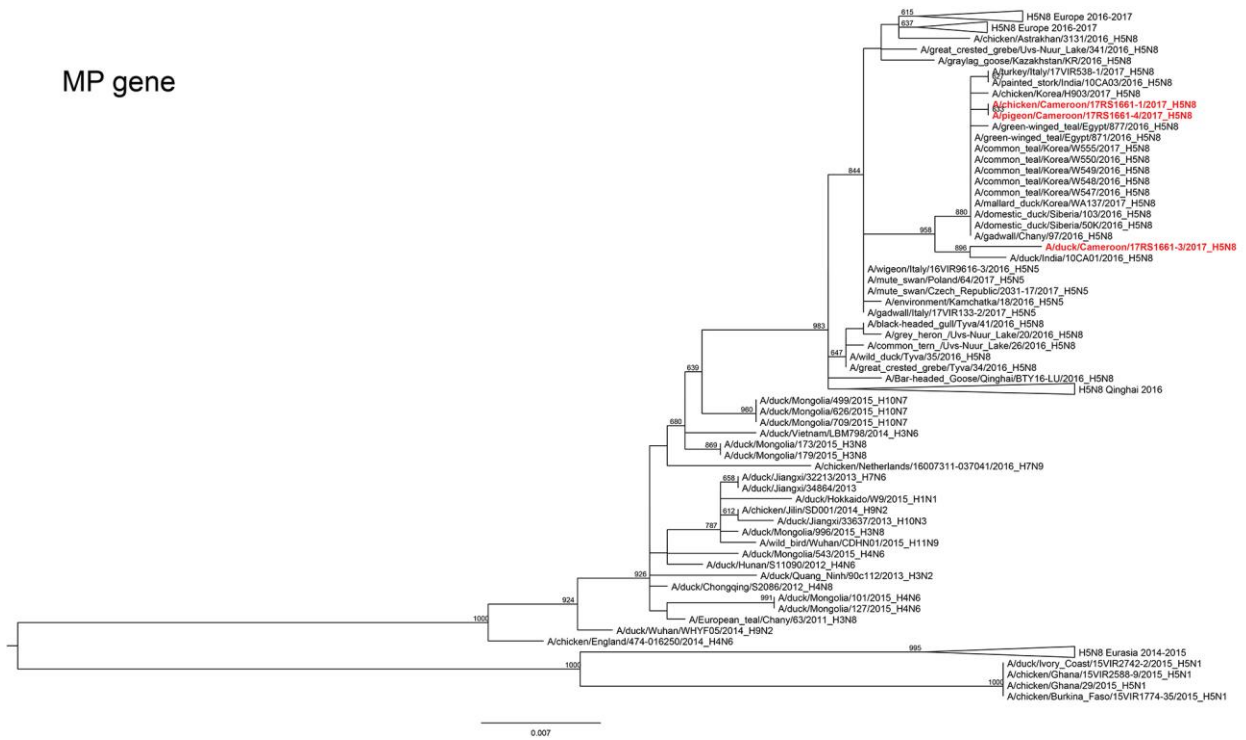
Technical Appendix 1 Figure 1. Maximum likelihood phylogenetic tree of the hemagglutinin (HA) gene. The H5N8 viruses from Cameroon are marked in red. Bootstrap supports higher than 600/1000 are indicated next to the nodes; scale bar indicates numbers of nucleotide substitutions per site.

NA gene



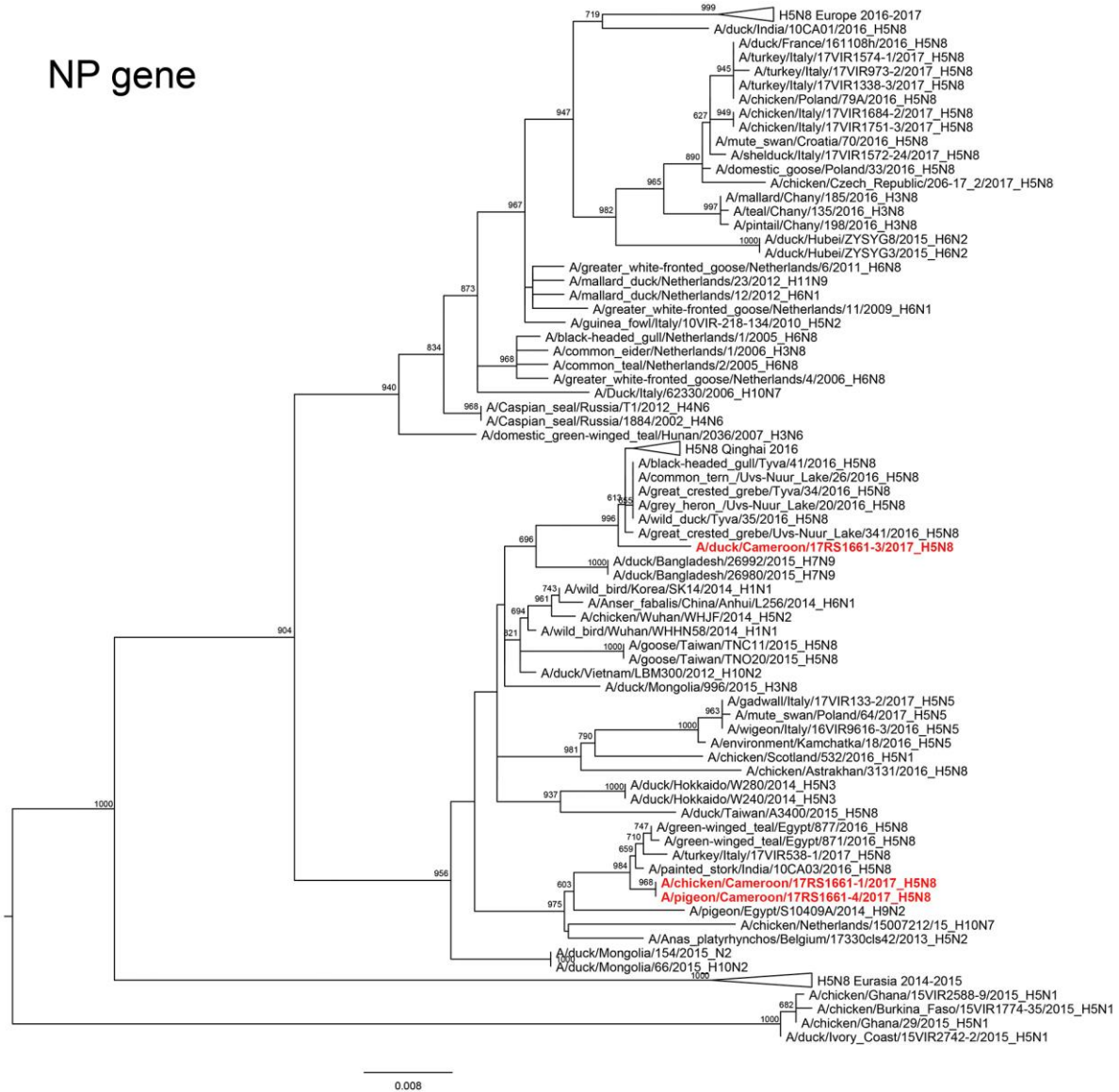
Technical Appendix 1 Figure 2. Maximum likelihood phylogenetic tree of the neuraminidase (NA) gene. The H5N8 viruses from Cameroon are marked in red. Bootstrap supports higher than 600/1000 are indicated next to the nodes; scale bar indicates numbers of nucleotide substitutions per site.

MP gene



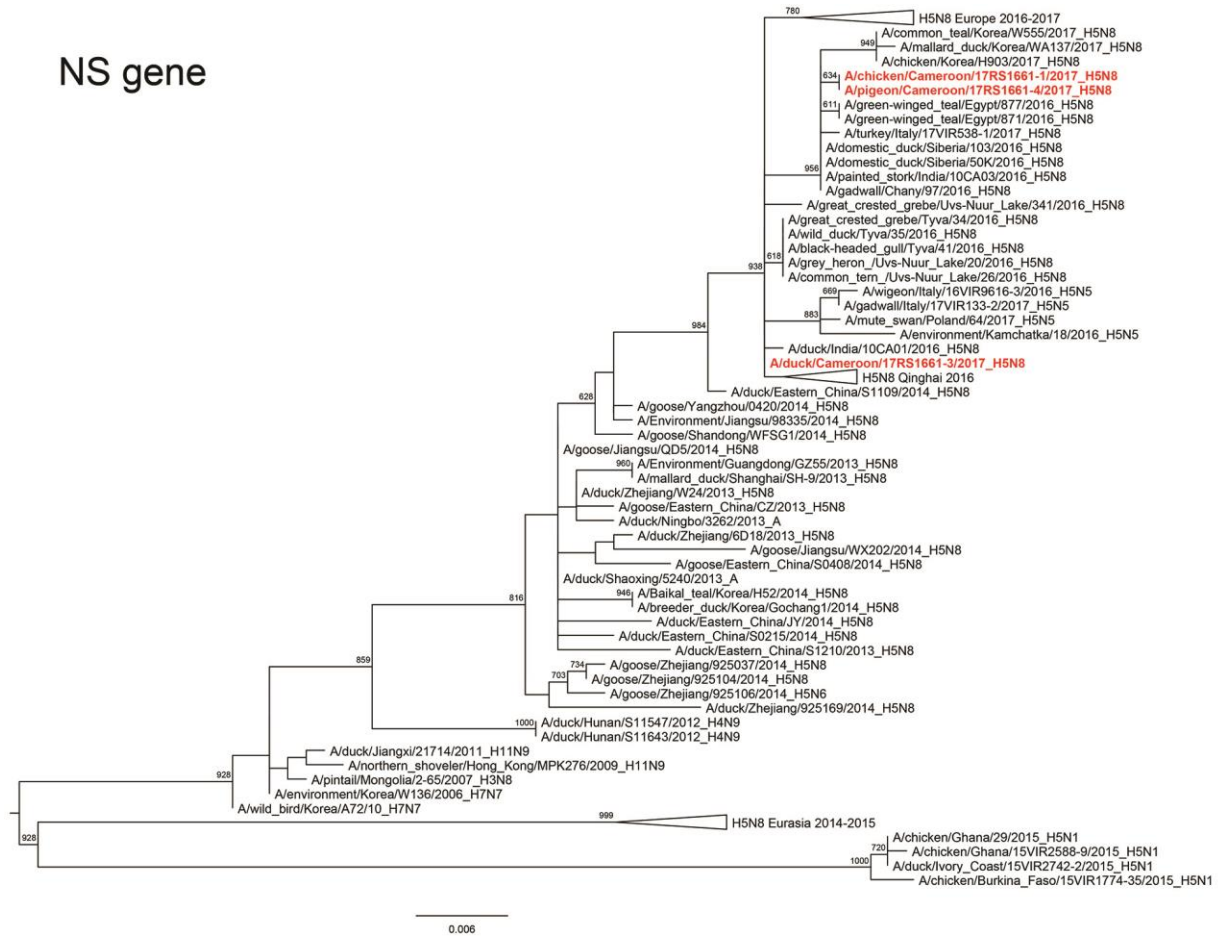
Technical Appendix 1 Figure 3. Maximum likelihood phylogenetic tree of the matrix protein (MP) gene. The H5N8 viruses from Cameroon are marked in red. Bootstrap supports higher than 600/1000 are indicated next to the nodes; scale bar indicates numbers of nucleotide substitutions per site.

NP gene



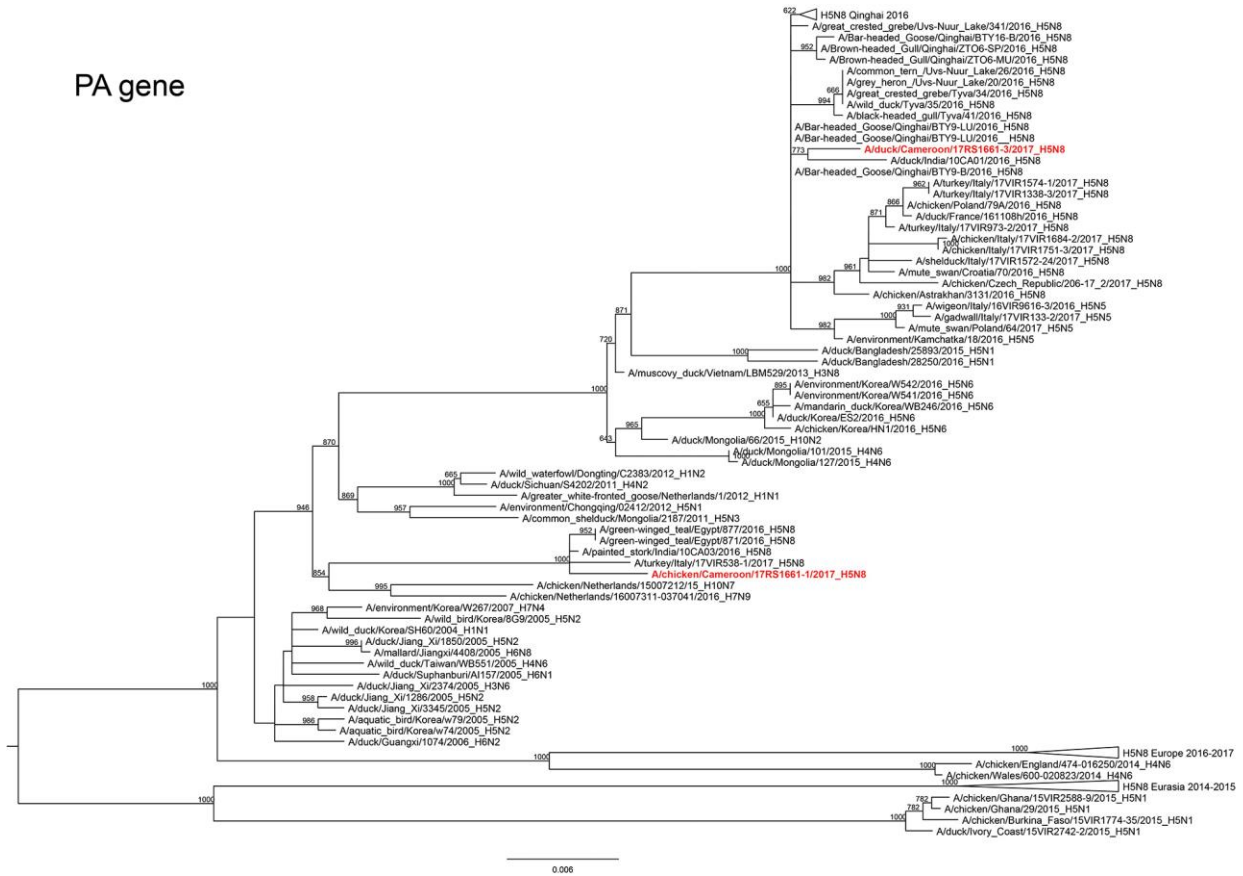
Technical Appendix 1 Figure 4. Maximum likelihood phylogenetic tree of the nucleoprotein (NP) gene. The H5N8 viruses from Cameroon are marked in red. Bootstrap supports higher than 600/1000 are indicated next to the nodes; scale bar indicates numbers of nucleotide substitutions per site.

NS gene



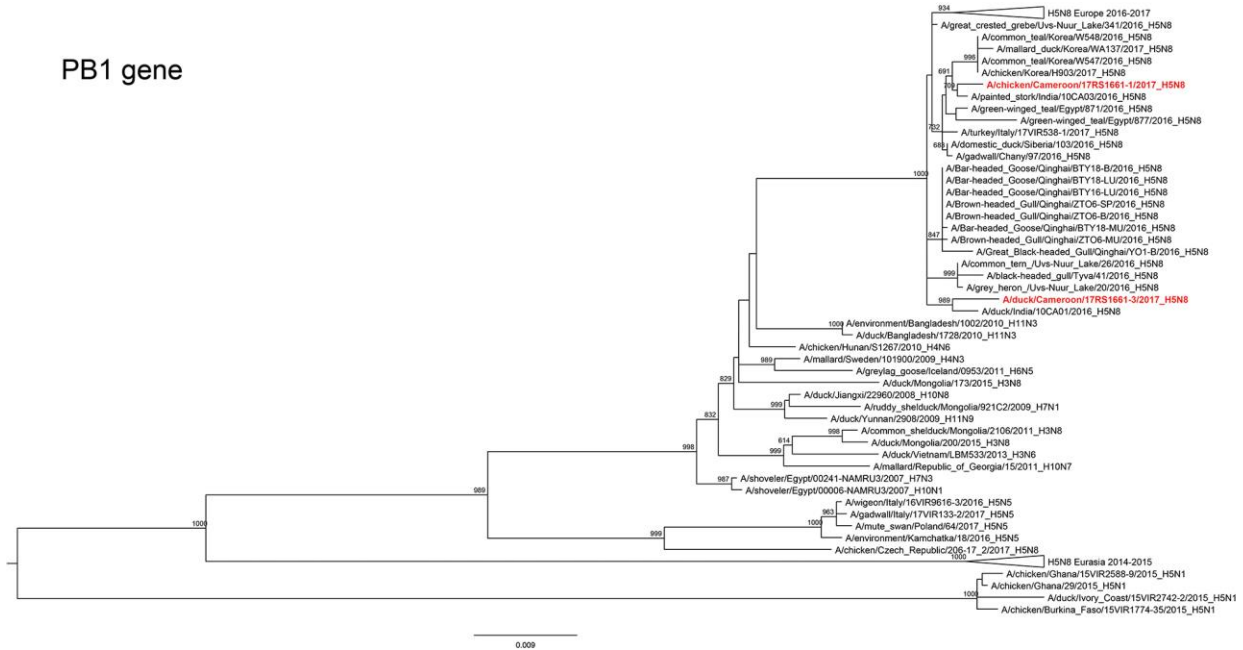
Technical Appendix 1 Figure 5. Maximum likelihood phylogenetic tree of the nonstructural protein (NS) gene. The H5N8 viruses from Cameroon are marked in red. Bootstrap supports higher than 600/1000 are indicated next to the nodes; scale bar indicates numbers of nucleotide substitutions per site.

PA gene



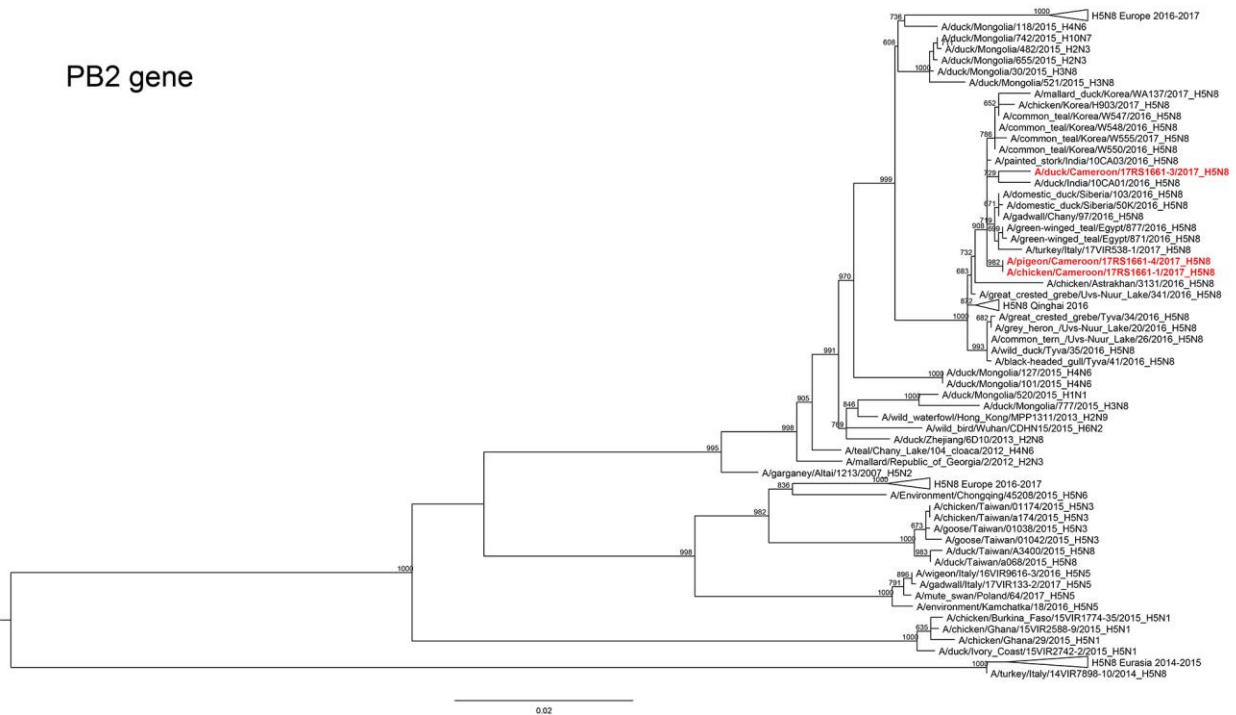
Technical Appendix 1 Figure 6. Maximum likelihood phylogenetic tree of the polymerase acidic protein (PA) gene. The H5N8 viruses from Cameroon are marked in red. Bootstrap supports higher than 600/1000 are indicated next to the nodes; scale bar indicates numbers of nucleotide substitutions per site.

PB1 gene

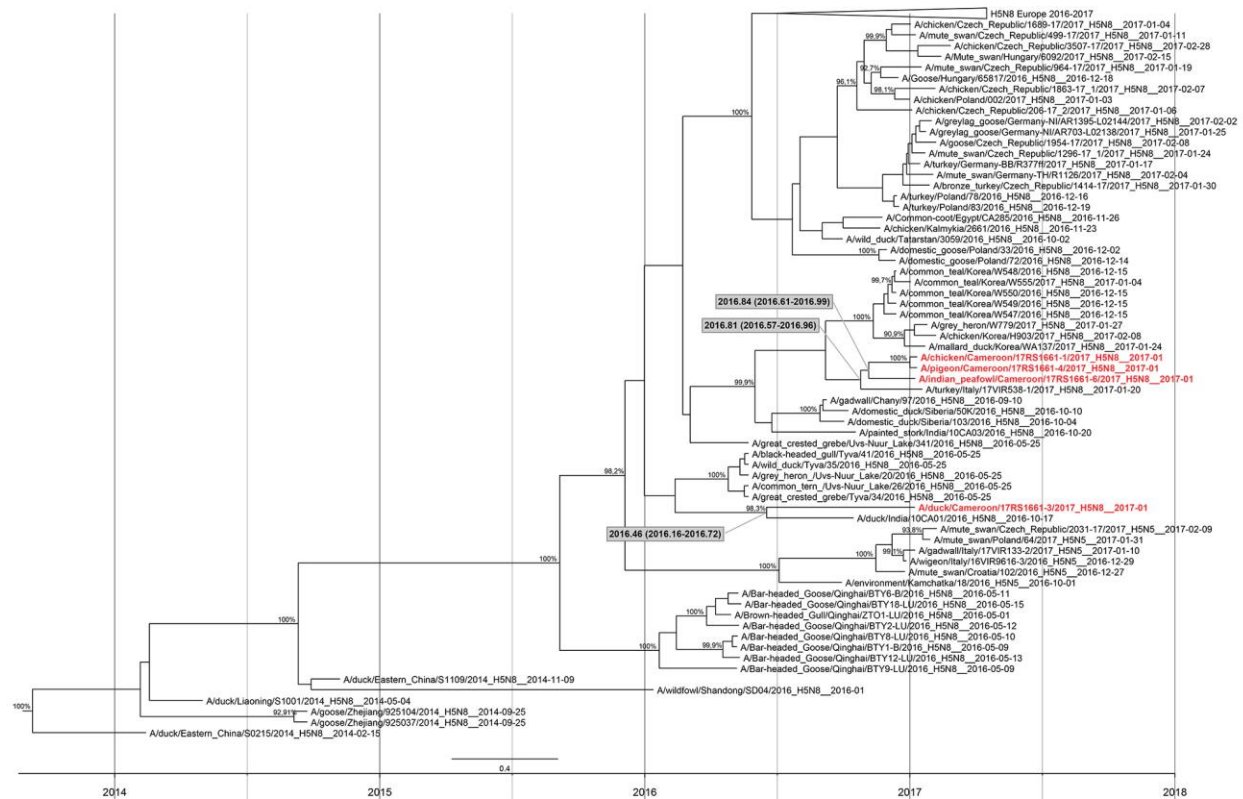


Technical Appendix 1 Figure 7. Maximum likelihood phylogenetic tree of the polymerase basic protein 1 (PB1) gene. The H5N8 viruses from Cameroon are marked in red. Bootstrap supports higher than 600/1000 are indicated next to the nodes; scale bar indicates numbers of nucleotide substitutions per site.

PB2 gene



Technical Appendix 1 Figure 8. Maximum likelihood phylogenetic tree of the polymerase basic protein 2 (PB2) gene. The H5N8 viruses from Cameroon are marked in red. Bootstrap supports higher than 600/1000 are indicated next to the nodes; scale bar indicates numbers of nucleotide substitutions per site.



Technical Appendix 1 Figure 9. Maximum clade credibility phylogenetic tree of the hemagglutinin (HA) gene. The H5N8 viruses from Cameroon are marked in red. Posterior probability values higher than 90% are indicated next to the nodes. The mean time to the most recent common ancestor (tMRCA) and the 95% highest posterior density intervals of the relevant nodes are indicated in the gray boxes.