Influenza D Virus in Animal Species in Guangdong Province, Southern China

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Molecular tests revealed influenza D viruses of D/OK lineage widely circulating in farmed animal species in Guangdong Province, southern China. In particular, we found high levels of influenza D virus infection in goats and pigs. We also detected viral RNA in serum specimens and feces of animals with certain severe diseases.

Tour types of influenza viruses (A–D) have been confirmed (https://www.cdc.gov/flu/about/viruses/types. htm). The recently discovered influenza D virus is thought to cause respiratory diseases primarily in cattle and to a lesser extent in pigs (1-4). Moreover, serologic evidence for influenza D virus infection in small ruminants and humans has been established (5,6). Since the initial influenza D virus isolation in the United States in 2011 (1), the virus has been reported in China, Mexico, France, Italy, and Japan (7-11). Genetic analysis of the hemagglutininesterase-fusion gene demonstrated that these viruses had 2 distinct lineages, represented by D/OK and D/660 (12). Recently, a novel influenza D virus that emerged in Japan has been proposed as the third lineage (11). D/OK lineage-related viruses were previously identified in native Luxi yellow cattle in Shandong Province, northern China (7). Despite good progress in identifying domestic cattle as the primary reservoir of influenza D virus, we know little about prevalence in other animals. We conducted a study to clarify the origin and transmission dynamics of influenza D virus in goats, buffalo, and pigs as well as farmed cattle.

The Study

In 2016, we collected 607 clinical samples from 4 species of animals with different clinical diseases and 250 nasal swab samples from asymptomatic animals (Table) from 16 farms in 4 cities of Guangdong Province: Guangzhou,

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Qingyuan, Heyuan, and Jiangmen (Figure 1). In addition, we randomly chose 200 archived Holstein dairy cattle serum samples, 40 per year, from 2011–2015 to investigate possible early RNA distribution of influenza D virus in this region. We used the reverse transcription PCR method and subcloning protocol (online Technical Appendix, https://wwwnc.cdc.gov/EID/article/23/8/17-0059-Techapp.pdf). We performed sequence alignment using ClustalW implemented in DNAStar software (DNAStar, Madison, WI, USA), and we conducted phylogenetic analyses based on our obtained sequences and reference truncated sequences (496-bp) of influenza D viruses from GenBank by using MEGA 5.1 software (http://www.megasoftware.net; online Technical Appendix Table).

After testing by reverse transcription PCR with further sequencing confirmation, we found influenza D virus-positive rates in 230 total nasal swab samples of 12.8% (20/156) for dairy cattle, 7.3% (4/55) for native yellow cattle, and 36.8% (7/19) for pigs. Rates in 324 total serum samples were 7.8% (15/193) for dairy cattle, 5.9% (3/51) for buffalo, and 33.8% (27/80) for goats. The influenza D virus-positive rate was also high (28.9%, 13/45) in swine lung samples. In contrast, we found no or low prevalence (<2%) in asymptomatic animals tested (Table). Moreover, all of the archived serum samples were found to be influenza D virus negative. Interestingly, 1 of 8 rectal swabs of goats with severe diarrhea tested positive (Table). Samples from animals with reproductive problems had a positive rate of 4.3% (5/116) (Table).

Sequence alignment analysis showed that the nucleotide sequences of influenza D viruses found in this study shared high similarity (99%–100%) with previously described sequences from China (7) and low similarity (93.8%–98.8%) with sequences originating from the United States, France, Italy, Mexico, and Japan (1,8-12). Similarly, phylogenetic analysis revealed that all influenza D virus sequences in this study clustered together with previous sequences from China and belonged to the D/OK lineage (Figure 2).

Conclusions

When first discovered, influenza D virus was reported in diseased pigs in the United States (1). Later, it was

¹These authors contributed equally to this article.

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Animal species and	Farm type†	Farm location	No.	Age range	Sample type	No. positive/no.	Detection
farm			animals	of animals		samples	rate, %
Holstein dairy cattle							
A	Not all-in-all-out	Guangzhou: Tianhe	2,000	3–5 y	Nasal swab	14/86‡	16.3
A	Not all-in-all-out	Guangzhou: Tianhe	2,000	3–5 y	Serum	10/94‡	10.6
В	Not all-in-all-out	Guangzhou:	800	3–6 y	Nasal swab	6/70‡	8.57
		Luogang					
В	Not all-in-all-out	Guangzhou:	800	3–6 y	Serum	5/99‡	5.05
		Luogang					
C	Not all-in-all-out	Guangzhou: Tianhe	175	2–5 y	Nasal swab	1/50§	2
American Landrace pig							
D	Not all-in-all-out	Guangzhou: Huadu	200	10–15 wks	Lung	4/10‡	40
E	All-in-all-out	Heyuan:	1,000	5–5 wks	Nasal swab	4/10‡	40
		Yuancheng					
E	All-in-all-out	Heyuan:	1,000	3–5 wks	Lung	1/8‡	12.5
		Yuancheng					
F	All-in-all-out	Jiangmen: Kaiping	800	8–20 wks	Nasal swab	3/9‡	30
F	All-in-all-out	Jiangmen: Kaiping	800	8–20 wks	Lung	8/27‡	29.6
G	All-in-all-out	Heyuan: Dongyuan	600	9–15 wks	Nasal swab	1/50§	2
Native hybrid white goat							
Н	Not all-in-all-out	Guangzhou:	200	0.5–5 y	Serum	7/25‡	28
		Zengcheng					
1	Not all-in-all-out	Guangzhou:	300	2–4 y	Serum	20/55¶	36.4
		Luogang					
Native hybrid black goat							
J	Not all-in-all-out	Qingyuan: Jiangkou	150	1–3 y	Rectal swab	1/8#	12.5
K	Not all-in-all-out	Jiangmen: Enping	500	1–4 y	Nasal swab	0/50§	0
Asian buffalo							
L	Not all-in-all-out	Guangzhou: Nansha	150	3–5 у	Serum	2/26¶	7.7
М	Not all-in-all-out	Guandzhou: Panvu	180	3–6 v	Serum	1/25¶	4
N	Not all-in-all-out	Qingvuan: Yingde	400	1–4 v	Nasal swab	0/508	0
Native vellow cattle				: ; ;		2.903	Ŭ
0	Not all-in-all-out	Qingvuan: Qingvin	200	2–5 v	Nasal swah	4/55†	7.3
P	Not all-in-all-out	Qingyuan: Eogang	230	1_3 v	Nasal swab	0/508	0
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Tahlo	Animal species	location sam	nle data and	detection rate	of influenza D	virus Guan	adona Province	China*
i abie.	Animai species,	location, sam	pie uala, and	a delection rate	or innuenza D	virus, Guari	iguong Frovince	, unina

*Feeding type of farms A–G was in captivity (poor biosecurity and high density). Feeding type of farms H–K and N–P was free grazing on the hills in the daytime and in captivity (poor biosecurity and high density) in the nighttime. Feeding type of farms L and M was free grazing in wetland in the daytime and in captivity (poor biosecurity and high density) in the nighttime.

†All-in-all-out is a strategy for the control of infectious disease. The barn is emptied of all animals and the accommodation is cleaned and disinfected and then refilled, all on 1 day.

‡These animals had severe respiratory diseases with a 10%–30% mortality rate, mainly characterized by expiratory dyspnea and abdominal respiration. §These animals were asymptomatic.

These animals had severe reproductive disorders with a 60%–70% abortion rate.

#These animals had severe diarrheal disease, characterized by watery diarrhea, limb weakness, and nearly dying.

identified in cattle and swine herds in several other countries, with or without clinical manifestation (7-11). Moreover, antibodies to influenza D virus were detected in goats, sheep, and humans (5-6). Under experimental conditions, influenza D virus replicated and transmitted among ferrets and guinea pigs (13). We confirmed that influenza D virus is widely present in cattle species (dairy cattle, yellow cattle, and buffalo). We also found influenza D virus at a high prevalence (>30%) in pigs and goats (Table), which is in contrast to the low prevalence found in previous investigations (1,5,10). The high prevalence may be caused by poor biosecurity measures and highdensity feeding mode practices in China's animal industry as well as possible cross-species transmission (13). Taken together, our findings expand the host range of influenza D virus and further emphasize the health concern this virus poses to multiple animal species.

Previous studies have shown that influenza D viruses are mainly found in respiratory tract samples (1-4,7,9-12)and that they have played an etiologic role in bovine respiratory diseases (2-4). In this study, we found that influenza D virus RNA was present in cattle and goat serum samples; it was also present in goat rectal swabs, accompanied by peste des petits ruminants virus and caprine kobuvirus (data not shown). The distribution of influenza D virus in our study is not the same as that described under experimental conditions (3).

Influenza viremia, an indicator of disease severity (14), has been detected in 20.9% of severe cases during the acute phase of infection or before host death. Our detection of influenza D virus genome in serum samples from severely diseased animals (Table) implies that the virus could enter transiently into the animal's circulatory system through capillaries lining the respiratory tract, which

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Figure 1. Farm locations for study of influenza D viruses in cattle, goats, buffalo, and pigs, Guangdong Province, China.

further contributes to the possibility of detecting virus in other organs. Similar to previous studies (2,4), we also found that the reverse transcription PCR positive rate was significantly higher (4%–40%) in diseased animals than the rate (\leq 2%) observed in asymptomatic animals (p<0.05), which suggests a potential correlation between the disease severity and presence of influenza D virus. For influenza D virus found in rectal swabs, it might be that animals have swallowed the virus. Another possibility is that, similar to influenza A and B viruses, influenza D virus can replicate within the intestinal tract (15).

We detected influenza D virus in cattle with reproductive disorders. However, we could not determine whether influenza D virus is associated with reproductive problems. Future studies can be designed to investigate these scientific issues.

To date, 2 lineages of influenza D virus (D/OK and D/660) co-circulate in North America and Europe (8-10,12). However, only the D/OK lineage has been found in China, and a potential third lineage was found in Japan (7,11). Our study confirms and further extends the previous observation that D/OK lineage circulates in East Asia. The viral, host, and ecologic factors that shape the observed contrasting phylodynamics of influenza D viruses among different geographic regions warrant further investigation.

In addition, we found different minor genetic variants circulating on the same farm (Figure 2), indicating the ongoing evolution of influenza D viruses in their hosts (7,8,11). In comparing our sequences to the reference sequences from different animal species, we found 4 frequent nucleotide mutations (at positions 136, 231, 263, and 486) (online Technical Appendix Figure 1), which caused 2 amino acid mutations at positions 77 and 88 (online Technical Appendix Figure 2). Interestingly, among 4 nucleotide mutations, 1 unique nucleotide (T at position 486) was originally from the D/660 lineage. Moreover, we found several consistent sequences co-circulating in multiple animal species (online Technical Appendix Figure 1). Our speculation is that homologous recombination among different influenza D viruses and potential cross-species transmission under field conditions are possible, but further study is needed.

In summary, our study investigating the infection status of influenza D virus in different farmed animal species in Guangdong Province provides novel insights into the epidemiology and evolution of this virus. In particular, we document the molecular evidence for influenza D virus infection in goats and buffalo.

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Technical Appendix

RT-PCR method and subcloning protocol used in this study

A classic reverse transcription PCR (RT-PCR) method was developed using a pair of primers (HEF-F: 5'-AAC CRC ATC TTC TTG TTC TTC A-3' and HEF-R: 5'-TGC TTC TTC WGT GGC ATT ATC T-3') targeting at the partial hemagglutinin-esterase-fusion (HEF) gene (at positions 582–1077) to test the presence of IDVs and further define IDV genetic lineages in those samples. For RT-PCR, the primers were diluted to 10 μ M with ddH₂O.

A 25 µL RT-PCR system was constructed using a one-step RT-PCR kit (Takara Bio Inc.), which contained 0.5 µL of each primer, 0.5 µL of enzyme mixture (including PrimeScript RTase, DNA polymerase, RNase inhibitor), 12.5 µL of 2 × buffer, 8.5 µL of ddH₂O, and 2.5 µL of viral RNA. RT-PCR was performed as follows: reverse transcription (RT) at 50°C for 30 min, 95°C for 5 min, followed by 35 PCR cycles (95°C for 30 sec, 55°C for 30 sec, 72°C for 30 sec), and a final extension at 72°C for 10 min. The positive PCR fragments (\approx 500 bp) were purified and cloned into the pGEM–T vector (TIANGEN, Inc., Beijing). Positive recombinant plasmids were purified according to the manufacturer's instructions (TIANGEN, Inc., Beijing) and sequenced using the Sanger sequencing method (Sangon Biotech Co., Ltd., Shanghai). RT-PCR detection data were analyzed by chi-square test (Ziyue software), and significance was set at p < 0.05. **Technical Appendix Table.** Reference influenza D virus isolates with partial hemagglutinin-esterase-fusion sequences used in this study

				GenBank
Strain or isolate	Specimen origin of isolate	Country	Year	accession no.
D/bovine/Nebraska/9–5/2012	Bovine (nasal swab)	USA	2012	KM392471
D/bovine/Oklahoma/660/2013	Bovine (nasal swab)	USA	2013	KF425662
D/bovine/Mississippi/C00014N/2014	Bovine (nasal swab)	USA	2014	KT581417
D/bovine/Mississippi/C00013N/2014	Bovine (nasal swab)	USA	2014	KT581416
D/bovine/Kansas/11-8/2012	Bovine (nasal swab)	USA	2012	KM392506
D/bovine/Kansas/13-21/2012	Bovine (nasal swab)	USA	2012	KM392492
D/bovine/Texas/3-13/2011	Bovine (nasal swab)	USA	2011	KM392485
D/bovine/Mexico/S56/2015	Bovine (nasal swab)	Mexico	2015	KU171128
D/bovine/Mexico/S8/2015	Bovine (nasal swab)	Mexico	2015	KU171127
D/bovine/Mexico/S7/2015	Bovine (nasal swab)	Mexico	2015	KU171126
D/bovine/France/2986/2012	Bovine (nasal swab)	France	2012	LN559126
D/bovine/Kansas/1-35/2010	Bovine (nasal swab)	USA	2010	KM392478
D/bovine/Mexico/S62/2015	Bovine (nasal swab)	Mexico	2015	KU171129
D/swine/Oklahoma/1334/2011	Swine (nasal swab)	USA	2011	JQ922308
D/bovine/Mississippi/C00030P/2014	Bovine (nasopharyngeal swab)	USA	2014	KT581418
D/bovine/Mississippi/C00046N/2014	Bovine (nasal swab)	USA	2014	KT581412
D/bovine/Minnesota/729/2013	Bovine (nasal swab)	USA	2013	KF425669
D/bovine/Minnesota/628/2013	Bovine (nasal swab)	USA	2013	KF425655
D/bovine/Kansas/14–22/2012	Bovine (nasal swab)	USA	2012	KM392499
D/bovine/Shandong/Y125/2014	Bovine (nasal swab)*	China	2014	KM015494
D/bovine/Shandong/Y127/2014	Bovine (nasal swab)*	China	2014	KM015501
D/bovine/Italy/46484/2015	Bovine (nasal swab)	Italy	2015	KT592526
D/bovine/Italy/1/2014	Bovine (nasal swab)	Italy	2014	KT592522
D/bovine/Shandong/Y217/2014	Bovine (nasal swab)*	China	2014	KM015508
D/swine/Italy/199724-3/2015	Swine (nasal swab)	Italy	2015	KT592533
D/bovine/Ibaraki/7768/2016	Bovine (nasal swab)	Japan	2016	LC128433
D/swine/Guangdong/YS1/2016	Swine (lung)	China	2016	KY441104
D/swine/Guangdong/YS2/2016	Swine (lung)*	China	2016	KY441105
D/swine/Guangdong/P8/2016	Swine (lung)	China	2016	KY441106
D/swine/Guangdong/P14/2016	Swine (nasal swab)	China	2016	KY441107
D/swine/Guangdong/PS1/2016	Swine (nasal swab)*	China	2016	KY441108
D/swine/Guangdong/U1/2016	Swine (serum)	China	2016	KY441109
D/swine/Guangdong/U16/2016	Swine (nasal swab)*	China	2016	KY441110
D/bovine/Guangdong/LG2/2016	Bovine: dairy cow (serum)	China	2016	KY441111
D/bovine/Guangdong/LG5/2016	Bovine: dairy cow (nasal swab)	China	2016	KY441112

				GenBank
Strain or isolate	Specimen origin of isolate	Country	Year	accession no.
D/bovine/Guangdong/LG9/2016	Bovine: dairy cow (nasal swab)*	China	2016	KY441113
D/bovine/Guangdong/QQ1/2016	Bovine: dairy cow (nasal swab)	China	2016	KY441114
D/bovine/Guangdong/QQ4/2016	Bovine: dairy cow (serum)	China	2016	KY441115
D/bovine/Guangdong/QQ7/2016	Bovine: dairy cow (serum)	China	2016	KY441116
D/bovine/Guangdong/QQ12/2016	Bovine: dairy cow (nasal swab)*	China	2016	KY441117
D/bovine/Guangdong/NS1/2016	Bovine: buffalo (serum)	China	2016	KY441118
D/bovine/Guangdong/RS1/2016	Bovine: yellow cattle (nasal swab)	China	2016	KY441119
D/bovine/Guangdong/RS4/2016	Bovine: yellow cattle (nasal swab)*	China	2016	KY441120
D/bovine/Guangdong/PY1/2016	Bovine: buffalo (serum)	China	2016	KY441121
D/caprine/Guangdong/JK1/2016	Caprine: goat (rectal swab)	China	2016	KY441122

*Healthy.

Majority	AACCGCATCTTCTT	GTTCTTCAAGO	TGGATGAAAA	AGCCCGTTGTC	GTATGCAGA	ATCTTCTGTTA	ATCCTGGAGO	CTAAACCTCAA	AGTTTGTGGG.	ACTGAG	
	10	20	30	40	50	60	70	80	90	100	
	I	20	50	10	50		10		50		-
JK1.seq	····		•••••••	· · · · · · · · · · · · · · · ·		· · · · · · C · · · ·	•••••	G	· · · · <mark>·</mark> · · · · ·	100	
LG2.seq	••• <mark>•</mark> •••••	· · · · · · · · · · · · · ·				C	····		· · · · · · · · · · · ·	100	
LG5.seq			••••••			C				100	
LG9.seq		· · · · · · · · · · · · ·	• • • • • • • • • • •			C	• • • • • • • • • • • •			100	
NS1.seq						C	· · · · · · · · · · ·			····· 100	
P14.seq						C				100	
P8.seq						C				100	
PS1.seq	· · · · · · · · · · · · · · · · · · ·					C				100	
PY1.seq						C				100	g
QQ1.seq		<mark>.</mark>				c	•••••	<mark></mark>		100	st
QQ12.seq						C				100	<u>.</u>
QQ4.seq						c.c				100	
QQ7.seq		<mark></mark>				c		<mark></mark> .		100	
RS1.seq						c				100	
RS4.seq						c				100	
Ul.seq						c				100	
Ul6.seq						C				100	D/OK
YS1.seq						C				100	
YS2.seq						c				100	
Y125.seq		<mark>.</mark>				c		<mark></mark>		100	
¥127.seq						c				100	
Y217.seq						C				100	
628.seg										100	
729.seg										100	
1-35. sea											
C00030P.seq			F							100	
14-22.seq	<mark>.</mark>		[<mark>.</mark>			<mark></mark>		100	
C00046N.seq			c					<mark>.</mark>		100	
1334.seg			r							100	
S62.seq			r							100	
46484.sea		.c	r					<mark>.</mark>		A 100	
1.seq			r			c		.c		A 100	
199724-3.sec	1		F				A			A 100	J
S8.seq								G		100	7
S7.seg											
\$56 sea	Д							G	C	100	
13-21 sea	д						G	G	C	100	
11-8 sec	۵						G	G	c	A 100	
9-5 sed	۵.							G	C	100	
660 seg	۵							G	с	100	0,000
COOOLAN sor	λ			••••••••••••	G			G	с	100	
COOOL3N sog	Δ			••••••	G			G		100	
3-13 200	Δ	•••••		7						100	
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7760 sog	с л			• • • • • • • • • • • • •					с	A 100	
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Majority	CAATCGGCAACTTT	TACTTTGCCG	ACAAGCTTCGC	AATTTACA	AATGCAACAAG	CATGTAGTGCA	AGCTTTGTTAC	CTTTGTGTAC	ЗААААСАААА	CAACAT		
	110	120	130	140	150	160	170	180	190	200		
JK1.seq									G	200	ר ו	
LG2.seq										200		
LG5.seq						<mark></mark>		<mark>.</mark>		200		
LG9.seq	· · · · · · · · · · · · · · · · · · ·									200		
NS1.seq					<mark>.</mark>		<mark></mark>	<mark></mark>		200		
P14.seq										200		
P8.sec										200		
PS1.sec												
PY1.sec										200	5	
001.sed											Ĕ	
0012 sea				a						200	ŝ	
001 sod										200	pi	
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Dg1 sog							• • • • • • • • • • • • •			200		
RS1.Seq			• • • • • • • • • • • • •	••••••						200		
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UI.seq		• • • • • • • • • • • •	· · · · <mark>·</mark> · · · · · · · ·	••••••			•••••••••••	· · · · · · · · · · · · · · · ·	· · · · · · · · · · · · ·	200		
U16.seq	•••••		• • • • • • • • • • • •	G .		•••••G•••••				200	1 1	
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YS2.seq	••••••••••••••		• • • • • • • • • • • •	G <mark></mark>						200 -		
Y125.seq			• • • • • • • • • • • • • • • • •	• <mark>••••</mark> •						200		
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Y217.seq	••••••••••••••••••••••••••••••••••••••		<mark>.</mark>		<mark>.</mark>	<mark>.</mark>		• • • • • • • • • • • • •	<mark></mark> .	200		
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729.seq			<mark>.</mark>	C						200		
1-35.seq	••••••			. <mark>.</mark>		<mark>.</mark>				200		
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Majority	TTAACACTTTTGGC	TGTGGAGATTAI	TACCA	AAATTACTATGAI	rgggaatgga	AACCTGATA	GGGGAATGGAT	AACAGAGTGGC	AGCATACAG	AGGAAT		
	210	220	230	240	250	260	270	280	290	300		
JK1.seq	· · · · · · · · · · · · · · · · ·	<mark>.</mark>		.G	C					300	٦	
LG2.seq	· · · · · · · · · · · · · · · · · ·			.G			A <mark></mark>	<mark></mark>		300		
LG5.seq				.G	c				G	300		
LG9.seq				.G	c					300		
NS1.seq				.G	c		. <mark></mark>			300		
P14.seq				.G	c					300		
P8.seq				.G	c					300		
PS1.seq				GG						300		
PY1.seq				.G						300	à	
001.seq				.G			A			300	ţ	
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004.seg		A		.G	c					300	Ē	
007. sea				G	С					300	F	
RS1 sea				G	c					300		
PS4 seg				G	c		D			300		
III soa				с	c					300		
Ul seq					c					300		
VCl.seq				.G						300		DIOR
isi.seq			••••				~			300		
isz.seq	•••••	••••••			·····		······			300 -	82	
1125.seq			• • • • •				• • • • • • • • • • • • • •			300		
Y127.seq			••••		C		••••••			300		
Y21/.seq	•••••				C		••••••		T	300		
628.seq		G	T			•••••	• <mark>•</mark> •••••			GC 300		
729.seq		G	T				• <mark>•</mark> •••••			GC 300		
1-35.seq		<mark></mark>	••••		A		C	<mark></mark>		300		
C00030P.seq			••••	• • • • • • • • • • • • • • •	• • • • • • • • • • •		• <mark>•••••</mark>			300		
14-22.seq			••••	• • • • • • • • • • • • • • •			• <mark>•••••</mark> •••••			300		
C00046N.seq			• • • • •			•••••	. <mark></mark>			300		
1334.seq			• • • • •				• <mark>•••••</mark> •••••			300		
S62.seq			· · · ·				. <mark></mark>			300		
46484.seq			· · · · ·				. <mark></mark>			300		
1.seq							. <mark></mark>			300		
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S7.seq	C				2	G.			.G	300		
S56.seq	T				3	G.				300		
13-21.seq	C				2	G.	c			300		
11-8.seq	A				2	G.				300		
9-5.sea	C				2	G.	R	<mark>.</mark>		300	. ⊢	- D/660
660.sea						G.				300		
C00014N.sec	C					G.				300		
C00013N.seq	C					G.				300		
3-13.sed						G				300		
2986.seg							G			300		
7768 sed	G									300		

ity	AGCAAACGCTGGAG	TTAAAATTGA	ATGTCCTTCCI	AAAATCTTGAA	ACCCTGGGAC	TTACAGCATT	AGATCAACACO	AAGATTCCT	CTAGTACCAR	AAAGG	
	310	320	330	340	350	360	370	380	390	400	
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Peq			• • • • • • • • • • • •							400	
pq			• • • • • • • • • • • •	• • • • • • • • • • • •						400	
eq	· · · · · · · · · · · · · · · · · · ·	•••••	• • • • • • • • • • • •	• • • • • • • • • • • • •	• • • • • • • • • • • •	· · · · · · · · · · · ·	• • • • • • • • • • • • •	••••••	· · · · · · · · · · · · · · ·	400.	
eq	·····		<mark></mark>			<mark></mark>	<mark></mark>	•••••		400	
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eq										400	
q									G.	400	
P									G.	400	
eq	A									400	
P.seq	A									400	
seq									3	400	
N.seq										400	
eq							.A			400	
a			<mark>.</mark>	<mark></mark>			.A			400	
sea	T									400	
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4	GG.T		A		••••••		• • • • • • • • • • • • •	A	G	400	
w.seq	GG.T		A					A	G	400	
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eq	GA				3 			G.A	G	400	

Majority	TCATACTGCTT	CGACACTGATGG	AGGGTACCCTA	FACAAGTAGT'	TCAATCTGAG	TGGTCAGCTT	CACGAAGATCA	AGATAATGCCA	ACAGAAGAAGCA			
	410	420	430	440	450	460	470	480	490			
JK1.seq				G						496	ר ו	
LG2.seq				G					T	496		
LG5.seq				G						496		
LG9.seq				G						496		
NS1.seq				G						496		
P14.seq				G						496		
P8.seq				G			<mark>.</mark>		T	496		
PS1.seq		····		G		<mark>.</mark>				496		
PY1.seq	· · · · · · · · · · · · · · · · · · ·			G		<mark></mark>	<mark>.</mark>	<mark>.</mark>		496	d	
QQ1.seq				G					T	496	stu	
QQ12.seq		.c		G						496	S	
QQ4.seq				G					. <mark></mark>	496	ЧЦ	
QQ7.seq				G	G				T	496		
RS1.seq				G					. <mark></mark>	496		
RS4.seq				G						496		
Ul.seq	<mark>.</mark>			G		<mark></mark>		<mark>.</mark>	. <mark>.</mark>	496		
U16.seq				G					. <mark></mark>	496	}	- D/OK
YS1.seq				G					. <mark>.</mark>	496		
YS2.seq		•••• <mark>••••••</mark> ••		G		• • • • • • • • • • • •			. <mark></mark>	496 -	e:	
Y125.seq				G					. <mark></mark>	496		
Y127.seq		G		G					. <mark></mark>	496		
Y217.seq				G	• • • • • • • • • • • • • • •	• • • • • • • • • • • • •			. <mark>.</mark>	496		
628.seq									. <mark></mark>	496		
729.seq									. <mark></mark>	496		
1-35.seq									. <mark></mark>	496		
C00030P.seq	<mark>.</mark>	····						• • • • • • • • • • • • •	. <mark></mark>	496		
14-22.seq									. <mark>.</mark>	496		
C00046N.seq	· · · <mark>·</mark> · · · · · · · ·				<mark></mark>					496		
1334.seq									· <mark>· ·</mark> · · · · · · · · · ·	496		
S62.seq			<mark>.</mark>			· · · · · · · · · · · ·			. <mark></mark>	496		
46484.seq									. <mark>.</mark>	496		
1.seq				. <mark>.</mark>	<mark></mark>				. <mark></mark>	496		
199724-3.sec	[. .					. <mark>.</mark>	496	4	
S8.seq	T			G	<mark></mark>				T	496	1	
S7.seq	T			G	<mark>.</mark>				T	496		
S56.seq	T	····	••••• <mark>•</mark> •••••	G	<mark>.</mark>			• • • • • • • • • • • •	T	496		
13-21.seq	T	····	<mark>.</mark>	G	<mark></mark>			<mark>.</mark>	T	496		
11-8.seq	T			G					.1 T	496		1000000000000
9-5.seq	T			G					T <mark></mark>	496	ł	– D/660
660.seq	T			G					T	496		
C00014N.seq	T			G					<mark>T</mark>	496		
C00013N.seq	T		••••••	G		• • • • • • • • • • •	• • • • • • • • • • •		^T	496		
3-13.seq	T		••••• <mark>•</mark> •••••	G	• • • • • • • • • • • •		•••••	• • • • • • • • • • • • •	<mark>T</mark>	496		
2986.seq	T		A					<mark>.</mark>	TG	496	_	
7768.seq	T			G				C	🔟	496		

Technical Appendix Figure 1. Multiple sequence alignment results of the current IDV nucleotide

sequences (496 bp) and corresponding published reference sequences.

T	ASSCSSSWMKSPLW	YAESSVNPGA	AKPQVCGTEQ	SATETLETSEC	JIYKCNKHVV(QLCYFVYENKI	TENTEGCGDYY	QNYYDGNGNI	'IG9	MDNRVAAYRGI		
2	10	20	30	40	50	60	70	80	4	90 10	0	
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	Τ		.RA				AL	• • • • • • • • • • •	v.,		100	
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rity	ANAGVKIECPSKIL	NPGTYSIRST	PRFLLVPKRS	ICFDTDGGYP.	IQVVQSEWSA	SRRSDNATEEA
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0	.GSR	R	.K			
c	.GS		К			I
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	.GS	<mark>.</mark>	.к			
.pro	.GS		к		<mark>.</mark>	<mark>.</mark>
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oro	.GSX		К			
ro	GA					
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LO						

Technical Appendix Figure 2. Multiple sequence alignment results of the current IDV amino acid

sequences (165 aa) and corresponding published reference sequences.