Pneumococcal Infection among Children before Introduction of 13-Valent Pneumococcal Conjugate Vaccine, Cambodia

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Vaccination of children with pneumococcal conjugate vaccine (PCV13) was initiated in Cambodia in 2015. To determine baseline data, we collected samples from children in 2013 and 2014. PCV13 serotypes accounted for 62.7% of colonizing organisms in outpatients and 88.4% of invasive pneumococci overall; multidrug resistance was common. Thus, effectiveness of vaccination should be high.

Infection with *Streptococcus pneumoniae* remains a substantial cause of death among children (*1*). In high-income countries, introduction of pneumococcal conjugate vaccine (PCV) has substantially decreased incidence of invasive pneumococcal disease (IPD) (*2*). Data for PCV effect in low-income countries are less robust (*2*). We therefore studied the characteristics of pneumococci responsible for colonization and invasive disease among children in Cambodia before the early 2015 introduction of 13-valent PCV (PCV13).

The Study

The study was conducted at Angkor Hospital for Children, Siem Reap, Cambodia. Before enrollment of a child, written consent was obtained from the parent/guardian. Ethical approval was granted by the hospital institutional review board and the Oxford Tropical Research Ethics Committee. For the colonization study, which was conducted in January (cool/dry season) and August (hot/wet season) 2014, colonization surveys were conducted in the outpatient department. Nasopharyngeal swab samples were collected from children 1 month to 15 years of age who had minor illnesses, excluding nonsevere pneumonia, not requiring hospital admission. Children were eligible for enrollment 1 time per survey. For the invasive disease study, which was conducted during August 1, 2013–July 31, 2014, samples

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were collected from hospitalized children 1 month to 15 years of age who met World Health Organization (WHO) clinical case definitions for pneumonia, meningitis, or sepsis (*3*). Children readmitted within 14 days were excluded from reenrollment. Samples were processed according to the WHO pneumococcal colonization detection protocol (*4*). Pneumococci were confirmed by optochin susceptibility and/or bile solubility and were serotyped by latex agglutination (*5*). Antimicrobial drug susceptibilities were determined according to Clinical and Laboratory Standards Institute guidelines (*6*). Serotype and antimicrobial drug susceptibilities were also determined for all invasive pneumococcal isolates cultured from patients during January 1, 2013–December 1, 2014. Pneumococci were grouped into vaccine serotypes (PCV13: 1, 3, 4, 5, 6A, 6B, 7F, 9V, 14, 18C, 19F, 19A, 23F), nonvaccine serotypes (all others), and nontypeable isolates. Multidrug resistance was defined as resistance to ≥3 agents (online Technical Appendix, http://wwwnc.cdc.gov/EID/article/21/11/15-0914-Techapp1.pdf) (*7*).

The outpatient colonization study included 974 children (Table 1; online Technical Appendix Figure 1). None were known to be HIV infected. Pneumococcal colonization was detected in 601 (61.7%) of children (online Technical Appendix Table 1). Colonization prevalence declined with age: 78.6% (206/262) in those 1–11 months, 61.9% (284/459) in those 12–59 months, and 43.9% (111/253) in those ≥5 years of age. The proportion colonized were 75.2% (342/455) in the cool/dry season and 49.9% (259/519) in the hot/wet season (p<0.001). The adjusted odds ratio for colonization in the hot/wet season was 0.38 (95% CI 0.28–0.51, p<0.001) after controlling for age, household size, cohabitation with other young children, current upper respiratory tract symptoms, and recent antimicrobial use. A total of 667 pneumococci were isolated (Figure 1). Among 601 colonized children, >1 serotype was identified in 11.0% (66/601). PCV13 serotypes accounted for 62.7% (418/667), nonvaccine serotypes for 29.5% (197/667), and nontypeable isolates for 7.8% (52/667) of isolates. The proportion of children colonized by PCV13 serotypes was greater among those <5 years of age (70.2% [344/490]) than among older children (48.6% [54/111]); p<0.001; whereas the opposite was true for colonization with nonvaccine serotypes (27.8% [136/490] vs. 48.6% [54/111]; p<0.001). Colonization with nontypeable isolates did not vary by age (data not

Table 1. Characteristics of 974 children enrolled in outpatient pneumococcal colonization surveys at Angkor Hospital for Children, Siem Reap, Cambodia, January and August 2014*

Characteristic	Overall	January 2014	August 2014	p value†
Total no. enrolled	974	455	519	
Age, median (IQR)	2.5 (0.9–5.1)	1.9 (0.9–4.1)	2.9 (1.1–6.0)	<0.001
Age category, no. (%)				
1–11 mo	262 (26.9)	142 (31.2)	120 (23.1)	0.005
12–59 mo	459 (47.1)	231 (50.8)	228 (43.9)	0.03
5–15 y	253 (26.0)	82 (18.0)	171 (33.0)	<0.001
Male sex, no. (%)	499 (51.2)	241 (53.0)	258 (49.7)	0.3
Reason for outpatient visit, no. (%)				
Upper respiratory tract infection	794 (81.5)	394 (86.6)	400 (77.1)	<0.001
Gastroenteritis	92 (9.5)	47 (10.3)	45 (8.7)	0.4
Other	88 (9.0)	14 (3.1)	74 (14.2)	<0.001
Antimicrobial drug use in preceding month, no. (%)‡	453/967 (46.8)	149/453 (32.9)	175/514 (34.0)	0.7
Household size, median (IQR)	5 (4–6)	5 (4–7)	5 (4–6)	0.01
Other children <5 y of age in household, no. (%)	841/973 (86.4)	422/454 (93.0)	419 (80.7)	<0.001
Attendance at school or daycare, no. (%)	293/973 (30.1)	105/454 (23.1)	188 (36.2)	<0.001

*Weather conditions were cool and dry in January and hot and wet in August. Where data were missing, an alternate denominator is included in the affected cell. IQR, interquartile range.
†For proportions, comparisons were made by using the χ² test. For continuous variables, comparisons were made by using the Wilcoxon rank-sum test.
‡Includes definite and possible (unknown systemic medication) consumption in the community before outpatient visit.

shown). Overall, 68.8% (459/667) of pneumococci were multidrug resistant: 85.4% of PCV13 isolates, 50.0% of nontypeable isolates, and 38.6% of nonvaccine serotypes (p<0.001). Among colonized children, multidrug-resistant pneumococci were more commonly cultured from children <5 years of age (75.1% [368/490]) than from older children (53.2% [59/111]); p<0.001.

From August 1, 2013, through July 1, 2014, a total of 2,613 cases of medical admissions were screened; of these, 1,009 were included in the analysis (online Technical Appendix Figure 1). Median patient age at admission was 1.2 years (interquartile range 0.5–2.4), 56.5% (570/1,009) of patients were male, and 1.4% (14/1,006) were HIV positive. Most cases met the WHO category of severe pneumonia (online Technical Appendix Table 2). Pneumococcal colonization was identified in 29.1% (293/1,008) of children from whom a swab sample was obtained (online Technical Appendix Table 3). Colonization was less frequent in those who had received ≥1 dose of an antimicrobial drug

(most frequently ceftriaxone) in hospital before the swab sample collection (23.8% [187/785]) than among those who had not (48.5% [95/196]); p<0.001. Colonization was identified in 31.3% (175/559) of children during the dry seasons (hot: March–May; cool: November–February) and in 26.3% (118/449) during the wet season (June–October); p = 0.08. A total of 305 pneumococci were isolated, comprising 27 serotypes plus nontypeable isolates (Figure 2). PCV13 serotypes accounted for 71.1% (217/305) of isolates, nonvaccine serotypes for 15.4% (47/305), and nontypeable isolates for 13.4% (41/305). Multidrug resistance was found in 79.3% (242/305) of isolates.

During 2013–2014, a total of 43 cases of IPD were culture proven (online Technical Appendix). Median patient age was 2.5 years (interquartile range 1.4–8.6). Overall, PCV13 serotypes accounted for 38 (88.4%, 95% CI 74.9–96.1) infections (Table 2). Multidrug resistance was identified in 55.8% (24/43): 22/38 (57.9%) of PCV13 serotypes and 2 (40.0%) of 5 nonvaccine serotypes;
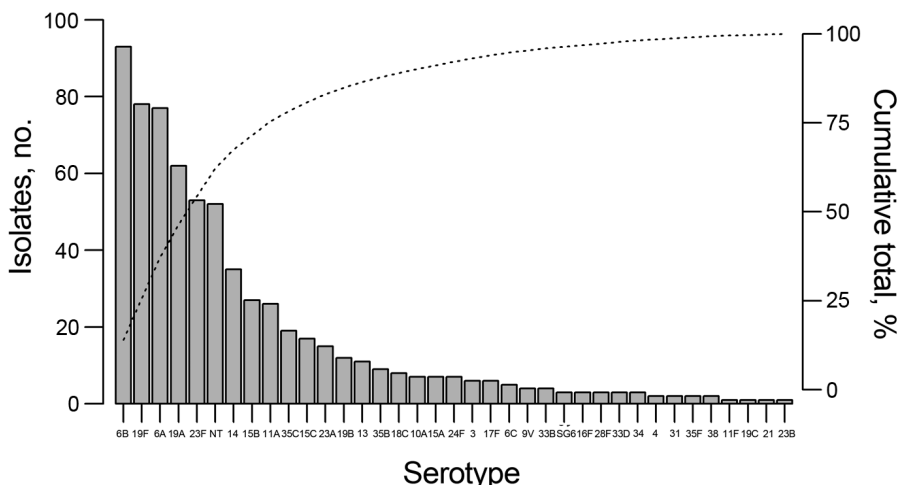

Figure 1. Serotype distribution of 667 pneumococcal isolates cultured from nasopharyngeal swab samples collected from 974 outpatients 1 month–15 years of age, at Angkor Hospital for Children, Cambodia, Siem Reap, January and August 2014. Bars indicate number of isolates; dotted line indicates cumulative total percentage of isolates.
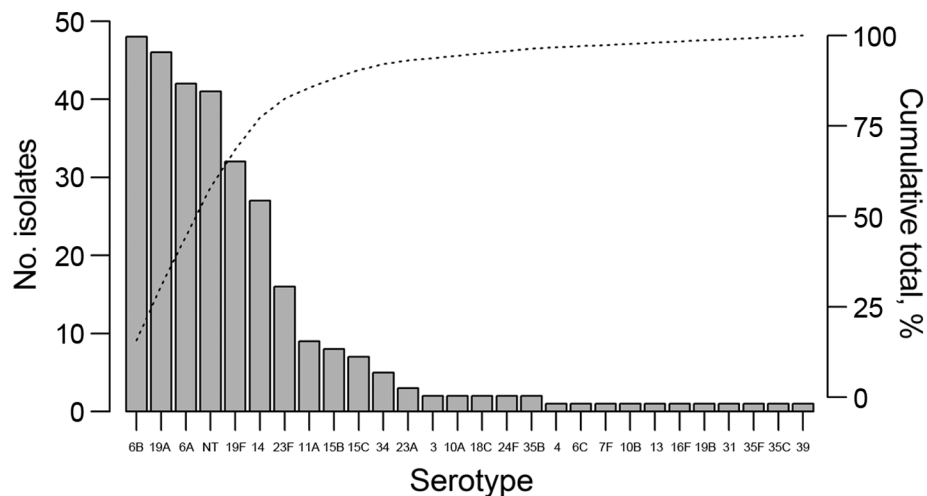
Figure 2. Serotype distribution of 305 pneumococcal isolates cultured from nasopharyngeal swab samples collected from 1,008 hospitalized patients 1 month–15 years of age at Angkor Hospital for Children, Siem Reap, Cambodia, August 2013–July 2014. Bars indicate number of isolates; dotted line indicates cumulative total percentage of isolates.

p = 0.6. Full resistance profiles are provided in online Technical Appendix Table 4.

Conclusions

This study highlights the high potential for reduction of IPD among children after introduction of PCV13 in Cambodia; 88.4% (95% CI 74.9–96.1) of invasive isolates from this 1 surveillance site were serotypes covered by the vaccine. Vaccination should result in decreased drug-resistant pneumococcal infections, although the substantial reservoir of resistance in nonvaccine type and nontypeable pneumococci will probably erode any reduction over time (*8–10*).

Colonization was high among outpatients and similar to that in other Southeast Asia locations (*5,11*). Multidrug resistance was common, probably the result of poor regulation of antimicrobial drug use in Cambodia (*12*); 72.1% of colonizing isolates and 55.8% of invasive isolates were multidrug resistant. For comparison, a recent study of children in Thailand found 31.6% of colonizing pneumococci to be multidrug resistant (*13*).

The range of serotypes detected in the colonization study was broad but slightly more restricted than that detected in other low-income country studies. In a longitudinal colonization study of refugee infants on the Thailand–Myanmar border, 67 serotypes were identified (*5*). This finding may reflect the high prevalence of antimicrobial drug use in the community, which would reduce the colonization prevalence of less resistant nonvaccine serotypes. However, the identification of several serotypes emerging as causes of IPD in South Africa, the United Kingdom, and the United States after introduction of PCV13 (e.g., serotypes 15A, 15B/C, 23B, 24F; which accounted for 7.8% of colonizing pneumococci in our study) is noteworthy, indicating the need for close monitoring for changes in colonization and IPD serotype distribution after PCV13 introduction (*7,14,15*).

The study has several limitations. The absolute number of IPD cases was small, and it was not possible to calculate disease incidence rates. The high prevalence of prehospitalization antimicrobial drug use hampered accurate IPD surveillance. Failure to detect more antimicrobial-drug susceptible nonvaccine type infections as a result of prehospitalization antimicrobial drug use may have falsely elevated the proportion of disease covered by PCV13. The low prevalence of colonization among hospitalized children highlights the need for swab sample collection before in-hospital antimicrobial drug administration for accurate evaluation of colonization in unwell children. Because the study was conducted at 1 site, caution is required when extrapolating the results to the general population of Cambodia. These data provide a baseline against which to monitor effectiveness of vaccinating children with PCV13 in Cambodia.

Table 2. Serotypes of invasive pneumococcal isolates from hospitalized children, Angkor Hospital for Children, Siem Reap, Cambodia, 2013–2014*

Specimen type	No.	PCV13 serotype, no. (%)	Serotypes, (no.)
Blood	35	31 (88.6)	1 (10), 6B (9), 14 (4), 23F (3), 6A (2), 12F† (1), 16F† (1), 19F (1), 19A (1), 18C (1), 28F† (1), nontypeable† (1)
Cerebrospinal fluid‡	3	3 (100)	1 (1), 6B (1), 19A (1)
Pleural fluid‡	4	4 (100)	1 (2), 5 (1), 19A (1)
Vitreous fluid	1	0 (0)	Nontypeable (1)†
Total	43	38 (88.4)	1 (13), 6B (10), 14 (4), 19A (3), 23F (3), 6A (2), NT† (2), 5 (1), 12F† (1), 16F† (1), 19F (1), 18C (1), 28F† (1)

*PCV13, 13-valent pneumococcal conjugate vaccine.
†Non-PCV13 serotypes.
‡Identical pneumococci were isolated from pleural fluid for 3 patients and from cerebrospinal fluid for 2 patients.

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References

1. World Health Organization. Estimated Hib and pneumococcal deaths for children under 5 years of age, 2008 [cited 2014 Mar 9]. http://www.who.int/immunization/monitoring_surveillance/burden/estimates/Pneumo_hib/en/
2. Feikin DR, Kagucia EW, Loo JD, Link-Gelles R, Puhan MA, Cherian T, et al. Serotype-specific changes in invasive pneumococcal disease after pneumococcal conjugate vaccine introduction: a pooled analysis of multiple surveillance sites. PLoS Med. 2013;10:e1001517. http://dx.doi.org/10.1371/journal.pmed.1001517
3. World Health Organization Coordinated Invasive Bacterial Vaccine Preventable Diseases (IB-VPD) Surveillance Network. Surveillance network case definitions [cited 2014 Dec 4]. http://www.who.int/immunization/monitoring_surveillance/resources/IB-VPD_Case_Defs.pdf?ua=1
4. Satzke C, Turner P, Virolainen-Julkunen A, Adrian PV, Antonio M, Hare KM, et al. Standard method for detecting upper respiratory carriage of *Streptococcus pneumoniae*: updated recommendations from the World Health Organization Pneumococcal Carriage Working Group. Vaccine. 2013;32:165–79. http://dx.doi.org/10.1016/j.vaccine.2013.08.062
5. Turner P, Turner C, Jankhot A, Helen N, Lee SJ, Day NP, et al. A longitudinal study of *Streptococcus pneumoniae* carriage in a cohort of infants and their mothers on the Thailand–Myanmar border. PLoS ONE. 2012;7:e38271. http://dx.doi.org/10.1371/journal.pone.0038271
6. Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing. 17th informational supplement. CLSI document M100–S23. Wayne (PA): The Institute; 2013.
7. von Gottberg A, de Gouveia L, Tempia S, Quan V, Meiring S, von Mollendorf C, et al. Effects of vaccination on invasive pneumococcal disease in South Africa. N Engl J Med. 2014;371:1889–99. http://dx.doi.org/10.1056/NEJMoa1401914
8. Dagan R, Klugman KP. Impact of conjugate pneumococcal vaccines on antibiotic resistance. Lancet Infect Dis. 2008;8:785–95. http://dx.doi.org/10.1016/S1473-3099(08)70281-0
9. Mitchell PK, Lipsitch M, Hanage WP. Carriage burden, multiple colonization and antibiotic pressure promote emergence of resistant vaccine escape pneumococci. Philos Trans R Soc Lond B Biol Sci. 2015;370:20140342. http://dx.doi.org/10.1098/rstb.2014.0342
10. Chewapreecha C, Harris SR, Croucher NJ, Turner C, Marttinen P, Cheng L, et al. Dense genomic sampling identifies highways of pneumococcal recombination. Nat Genet. 2014;46:305–9. http://dx.doi.org/10.1038/ng.2895
11. Schultsz C, Vien le M, Campbell JI, Chau NV, Diep TS, Hoang NV, et al. Changes in the nasal carriage of drug-resistant *Streptococcus pneumoniae* in urban and rural Vietnamese schoolchildren. Trans R Soc Trop Med Hyg. 2007;101:484–92. http://dx.doi.org/10.1016/j.trstmh.2006.08.010
12. Emary KR, Carter MJ, Pol S, Sona S, Kumar V, Day NP, et al. Urinary antibiotic activity in paediatric patients attending an outpatient department in north-western Cambodia. Trop Med Int Health. 2015;20:24–8. http://dx.doi.org/10.1111/tmi.12398
13. Thummeepak R, Leerach N, Kunthalert D, Tangchaisuriya U, Thanwisai A, Sitthisak S. High prevalence of multi-drug resistant *Streptococcus pneumoniae* among healthy children in Thailand. J Infect Public Health. 2015;8:274-81. http://dx.doi.org/10.1016/j.jiph.2014.11.002
14. Moore MR, Link-Gelles R, Schaffner W, Lynfield R, Lexau C, Bennett NM, et al. Effect of use of 13-valent pneumococcal conjugate vaccine in children on invasive pneumococcal disease in children and adults in the USA: analysis of multisite, population-based surveillance. Lancet Infect Dis. 2015;15:301–9. http://dx.doi.org/10.1016/S1473-3099(14)71081-3
15. Waight PA, Andrews NJ, Ladhani SN, Sheppard CL, Slack MP, Miller E. Effect of the 13-valent pneumococcal conjugate vaccine on invasive pneumococcal disease in England and Wales 4 years after its introduction: an observational cohort study. Lancet Infect Dis. 2015;15:535–43. http://dx.doi.org/10.1016/S1473-3099(15)70044-7

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Pneumococcal Infection among Children before Introduction of 13-Valent Pneumococcal Conjugate Vaccine, Cambodia

Technical Appendix

Study Site Details

Angkor Hospital for Children (AHC) is a non-governmental pediatric referral hospital in Siem Reap, Cambodia. The hospital serves the population of northern Cambodia: ≈300–500 children attend the out-patient department each day and there are around 4,000 medical admissions per year. In 2013, Cambodia had an estimated under-5 year mortality of 38 per 1000 births (*1*). The country has a tropical climate with monsoon rains between May and October each year.

Methods

Nasopharyngeal swab processing

Flocked nylon nasopharyngeal swabs (NPS; Medical Wire and Equipment, Corsham, UK) were collected in accordance with the updated WHO pneumococcal colonization protocol (*2*). The NPS tips was excised into 1mL STGG (skim milk, tryptone, glucose, glycerol medium; prepared in house) immediately after collection. The NPS-STGG specimens were stored in a cool box for <8 hours before definitive pre-culture storage at −80°C.

Thawed nasopharyngeal swab (NPS-STGG) specimens were vortexed and 100μL of each specimen was plated onto 5% sheep blood agar (prepared in-house) and incubated overnight in a candle jar. All morphologically discrete α-hemolytic colonies were sub-cultured and identified as *Streptococcus pneumoniae* by susceptibility to optochin disc (Oxoid, Basingstoke, UK), with confirmation by 10% bile solubility if optochin zone diameter was 7–14mm. All confirmed pneumococcal isolates were serotyped by latex

agglutination with Quellung confirmation of ambiguous results (*3*). A random selection of 21 phenotypically non-encapsulated non-typeable isolates were confirmed as pneumococci by bile solubility and absence of capsule swelling using Omniserum (SSI Diagnostica, Hillerød, Denmark).

Antimicrobial susceptibilities were determined for all isolates following 2013 CLSI guidelines (*4*). MICs (MIC) to benzylpenicillin and ceftriaxone were determined by the Etest method (bioMérieux, Marcy L'Etoile, France). Susceptibility to chloramphenicol, clindamycin, erythromycin, tetracycline, and trimethoprim-sulphamethoxazole was determined by disc diffusion. Non-susceptibility to penicillin and ceftriaxone was defined as an MIC of >0.06µg/mL and >1 µg/mL, respectively. Multi-drug resistance was defined as resistance (non-susceptibility for the β-lactam drugs) to three or more agents, with clindamycin/erythromycin and benzylpenicillin/ceftriaxone counting as a single agent (*5*).

All culture work was performed at the Angkor Hospital for Children / Cambodia Oxford Medical Research Unit microbiology laboratory which is located within the AHC campus. Internal quality control procedures were in place for all aspects of the laboratory work (media preparation, antimicrobial susceptibility testing, and serotyping).

Invasive pneumococcal disease data

At AHC, blood cultures are routinely taken in hospitalized children with fever or suspected invasive bacterial infection, with lumbar punctures also done in cases of suspected meningitis: between 1st January 2013 and 31st December 2014 11,238 blood cultures and 901 cerebrospinal fluid cultures were processed by the AHC microbiology laboratory. All invasive pneumococcal isolates (defined as *S. pneumoniae* isolated from blood, cerebrospinal fluid (CSF), pleural fluid and other usually sterile sites) cultured between 1st January 2013 and 31st December 2014 were retrieved from −80°C storage and sub-cultured onto 5% sheep blood agar for serotyping as described above. Antimicrobial susceptibilities for these isolates were extracted from the microbiology laboratory database.

Data analysis

Analyses were done using the R statistical package version 3.2 (R Foundation for Statistical Computing, Vienna, Austria). Pneumococcal serotypes were grouped in vaccine serotypes (PCV13: serotypes 1, 3, 4, 5, 6A, 6B, 7F, 9V, 14, 18C, 19F, 19A, 23F), non-vaccine serotypes (NVT: all other typeable pneumococci), and non-typeable pneumococci (NT). Colonization was a binary variable but children colonized by multiple pneumococci

could be included in more than serotype group (PCV13, NVT, or NT). Continuous variables were described using median and interquartile ranges; groups were compared using the Wilcoxon Rank Sum test. Proportions were compared using the Chi-squared or Fisher exact test, as appropriate. For these tests, two-tailed p-values of <0.05, were indicative of statistical significance. Multivariable logistic regression was used to assess factors potentially influencing pneumococcal colonization. Adjusted odds ratios (AOR) with 95% confidence intervals (CI) not including one were indicative of statistical significance.

Technical Appendix Table 1. Pneumococcal nasopharyngeal colonization and antibiotic exposure in 974 out-patient children, Angkor Hospital for Children, Siem Reap, Cambodia, 2014

Variable	Colonization, n (%)*			
All children (N)	Abx (48)	Possible Abx (283)	No Abx (643)	Total (974)
Colonized	25 (52.1)	168 (59.4)	408 (63.5)	601 (61.7)
Colonized children (N)	Abx (25)	Possible Abx (168)	No Abx (408)	Total (601)
PCV13 serotype(s)	20 (80.0)	111 (66.1)	267 (65.4)	398 (66.2)
NVT serotype(s)	4 (16.0)	56 (33.3)	130 (31.9)	190 (31.6)
NT isolate	2 (8.0)	11 (6.5)	38 (9.3)	51 (8.5)
MDR isolate(s)	20 (80.0)	127 (75.6)	280 (68.6)	427 (71.0)

*Colonization data are stratified by antimicrobial exposure in the month before collection of the nasopharyngeal swab sample. (Abx, definite antimicrobial exposure; Possible Abx, exposure to an unknown systemic medication; No Abx, no antimicrobial exposure. Children colonized by >1 pneumococcal serotype may be represented more than once within the colonized group (total number of colonized children = 601; total number of pneumococci isolated = 667).

Technical Appendix Table 2. Characteristics and WHO surveillance definition categorization of 1,009 hospitalized children enrolled into the colonization study at Angkor Hospital for Children, Siem Reap, Cambodia, August 2013–July 2014

Characteristic	Age category, n (%)				Total, n (col %)
	<2 mo	2–11 mo	12–59 mo	5–15 y	
Total enrolled*	77	358	444	130	1009
Male Sex (%)	49 (63.6)	212 (59.2)	238 (53.6)	71 (54.6)	570 (56.5)
Report antimicrobial use in preceding month, n (%)†	42 (54.5)	227/354 (64.1)	280/438 (63.9)	87/127 (68.5)	636/996 (63.9)
Household size, median (IQR)	6 (4 – 7)	5 (4 – 7)	5 (4 – 7)	5 (4 – 7)	5 (4 – 7)
Other children <5 years of age in household, n (%)	77 (100.0)	355/355 (100.0)	432/442 (97.7)	53/129 (41.1)	917/1003 (91.4)
Attendance at school or day care, n (%)	0/75 (0.0)	0/354 (100.0)	17/441 (3.9)	81/129 (62.8)	98/999 (9.8)
Surveillance category					
Severe pneumonia	61 (8.5)	290 (40.3)	304 (42.3)	64 (8.9)	719 (71.2)
Suspected meningitis	13 (8.6)	41 (27.0)	71 (46.6)	27 (17.8)	152 (15.1)
Pneumonia	0 (0.0)	20 (26.0)	46 (59.7)	11 (14.3)	77 (7.6)
Probable bacterial meningitis	1 (2.9)	5 (14.7)	16 (47.1)	12 (35.3)	34 (3.4)
Very severe disease	2 (8.7)	1 (4.3)	6 (26.1)	14 (60.9)	23 (2.3)
Confirmed bacterial meningitis	0 (0.0)	1 (25.0)	1 (25.0)	2 (50.0)	4 (0.4)

*Where there was missing data an alternate denominator is included in the affected cell.
†Includes definite and possible (unknown systemic medication) consumption in the community before hospitalization.

Technical Appendix Table 3. Pneumococcal nasopharyngeal colonization and antibiotic exposure in 1,008 hospitalized children, Angkor Hospital for Children, Siem Reap, Cambodia, 2014

Variable	Colonization, n (%)*			
All children (N)	Abx (815)	Possible Abx (114)	No Abx (79)	Total (1008)
Colonized	203 (24.9)	54 (47.7)	36 (45.6)	293 (29.1)
Colonized children (N)	Abx (203)	Possible Abx (54)	No Abx (36)	Total (293)
PCV13 serotype	155 (76.4)	34 (63.0)	26 (72.2)	215 (73.4)
NVT serotype	27 (13.3)	11 (20.4)	9 (25.0)	47 (16.0)
NT isolate	26 (12.8)	11 (20.4)	3 (8.3)	40 (13.7)
MDR isolate	165 (81.3)	39 (72.2)	30 (83.3)	234 (79.9)

*Colonization data are stratified by antimicrobial exposure in the month before the nasopharyngeal swab, including drugs administered in hospital before swabbing (Abx: definite antimicrobial exposure; Possible Abx: exposure to an unknown systemic medication; No Abx: no antimicrobial exposure). Children colonized by >1 pneumococcal serotype may be represented more than once within the colonized group (total number of colonized children = 293; total number of pneumococci isolated = 305).

Technical Appendix Table 4. Antimicrobial susceptibilities of colonizing and invasive pneumococci cultured from children attending Angkor Hospital for Children, Siem Reap, Cambodia, 2013–14

	PCV13 serotypes	NVT serotypes	NT isolates	All isolates
Colonization isolates		Resistant, n (%)*		
Out-patient				
Total isolates	418	197	52	667
Penicillin	383 (91.6)	96 (48.7)	44 (84.6)	523 (78.4)
Ceftriaxone	98 (23.4)	8 (4.1)	8 (15.4)	114 (17.1)
Clindamycin	199 (47.6)	38 (19.3)	10 (19.2)	247 (37.0)
Chloramphenicol	71 (17.0)	8 (4.1)	2 (3.8)	81 (12.1)
Co-trimoxazole	338 (80.9)	82 (41.6)	32 (61.5)	452 (67.8)
Erythromycin	265 (63.4)	42 (21.3)	12 (23.1)	319 (47.8)
Tetracycline	394 (94.3)	148 (75.1)	38 (73.1)	580 (87.0)
MDR	357 (85.4)	76 (38.6)	26 (50.0)	459 (68.8)
In-patient				
Total isolates	217	47	41	305
Penicillin	200 (92.2)	26 (55.3)	30 (73.2)	256 (83.9)
Ceftriaxone	42 (19.4)	4 (8.5)	3 (7.3)	49 (16.1)
Clindamycin	114 (52.5)	7 (14.9)	2 (4.9)	123 (40.3)
Chloramphenicol	46 (21.2)	4 (8.5)	4 (9.8)	54 (17.7)
Co-trimoxazole	204 (94.0)	29 (61.7)	26 (63.4)	259 (84.9)
Erythromycin	135 (62.2)	8 (17.0)	6 (14.6)	149 (48.9)
Tetracycline	207 (95.4)	39 (83.0)	39 (95.1)	285 (93.4)
MDR	197 (90.8)	21 (44.7)	24 (58.5)	242 (79.3)
Invasive pneumococcal isolates				
Total isolates	38	3	2	43
Penicillin	20 (52.6)	1 (33.3)	1 (50.0)	22 (51.2)
Ceftriaxone	0 (0)	0 (0)	0 (0)	0 (0)
Clindamycin	11 (28.9)	0 (0)	0 (0)	11 (25.6)
Chloramphenicol	16 (42.1)	1 (33.3)	0 (0)	17 (39.5)
Co-trimoxazole	32 (84.2)	0 (0)	2 (100)	34 (79.1)
Erythromycin	12 (31.6)	0 (0)	0 (0)	12 (27.9)
Tetracycline	36 (94.7)	3 (100)	2 (100)	41 (95.3)
MDR	22 (57.9)	1 (33.3)	1 (50.0)	24 (55.8)

*Antimicrobial susceptibilities were determined following 2013 CLSI guidelines (*4*). Resistance was defined as disk test "resistant" except for the β-lactam drugs: benzylpenicillin MICs of >0.06 µg/mL were classified as "resistant"; ceftriaxone MICs of $\geq$1 µg/mL were classified as "resistant." Multidrug resistance (MDR) was defined as resistance to three or more agents, with clindamycin/erythromycin and benzylpenicillin/ceftriaxone counting as a single agent (*5*).
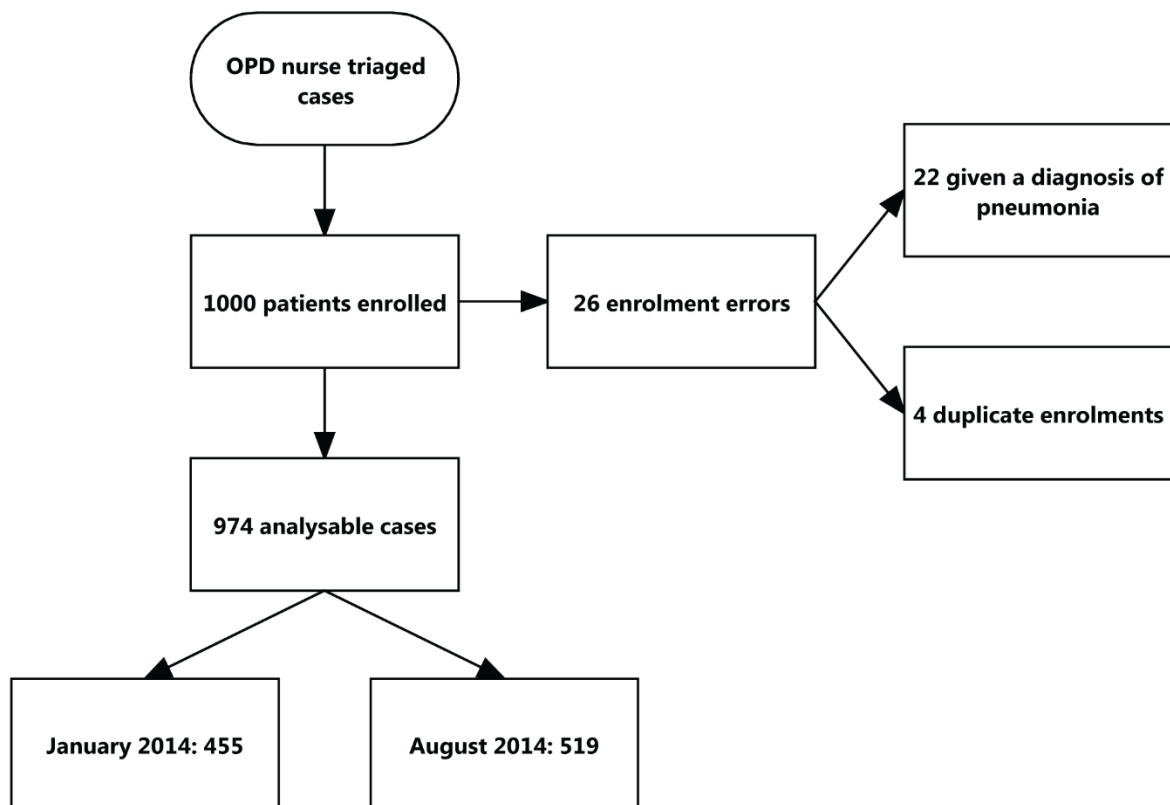
Technical Appendix Figure 1. Study flowchart for the out-patient pneumococcal colonization surveys at Angkor Hospital for Children, Siem Reap, Cambodia, January and August 2014.
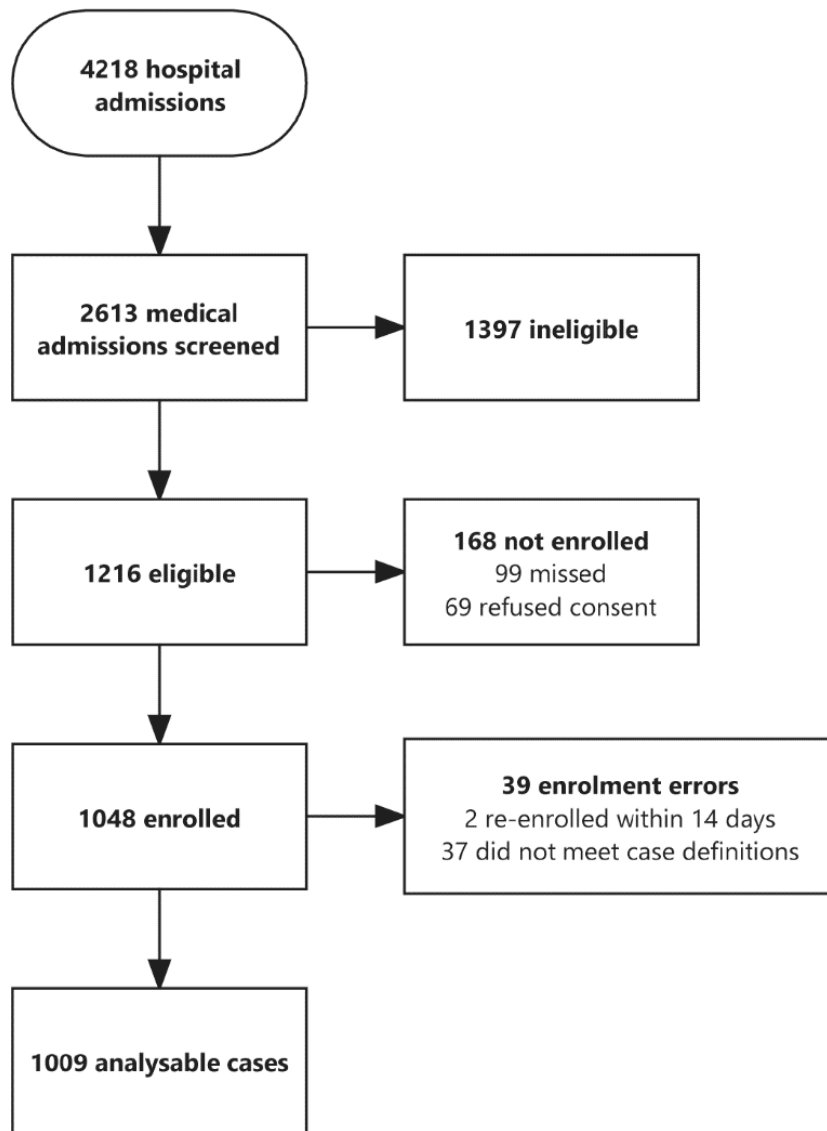
Technical Appendix Figure 2. Study flowchart for the hospitalized patient pneumococcal colonization study at Angkor Hospital for Children, Siem Reap, Cambodia, August 2013–July 2014.

References

1. Alliance GAVI. Cambodia [cited 2015 Apr 25]. http://www.gavi.org/country/cambodia/

2. Satzke C, Turner P, Virolainen-Julkunen A, Adrian PV, Antonio M, Hare KM, et al. Standard method for detecting upper respiratory carriage of *Streptococcus pneumoniae*: updated recommendations from the World Health Organization Pneumococcal Carriage Working Group. Vaccine. 2013;32:165–79. PubMed http://dx.doi.org/10.1016/j.vaccine.2013.08.062

3. Turner P, Turner C, Jankhot A, Helen N, Lee SJ, Day NP, et al. A longitudinal study of *Streptococcus pneumoniae* carriage in a cohort of infants and their mothers on the Thailand–Myanmar border. PLoS ONE. 2012;7:e38271. PubMed http://dx.doi.org/10.1371/journal.pone.0038271

4. Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing. 17th informational supplement. CLSI document M100–S23. Wayne (PA): The Institute; 2013.

5. von Gottberg A, de Gouveia L, Tempia S, Quan V, Meiring S, von Mollendorf C, et al. Effects of vaccination on invasive pneumococcal disease in South Africa. N Engl J Med. 2014;371:1889–99. PubMed http://dx.doi.org/10.1056/NEJMoa1401914