

Acute Encephalopathy Associated with Influenza A Infection in Adults

Nelson Lee, Chun Kwok Wong, Paul K.S. Chan, Niklas Lindegardh, Nicholas J. White, Frederick G. Hayden, Edward H.C. Wong, Ka Shing Wong, Clive S. Cockram, Joseph J.Y. Sung, and David S.C. Hui

We report acute encephalopathy associated with influenza A infection in 3 adults. We detected high cerebrospinal fluid (CSF) and plasma concentrations of CXCL8/IL-8 and CCL2/MCP-1 (CSF/plasma ratios >3), and interleukin-6, CXCL10/IP-10, but no evidence of viral neuroinvasion. Patients recovered without sequelae. Hyperactivated cytokine response may play a role in pathogenesis.

Influenza-associated acute encephalopathy has been described in children, and results in a high frequency of neurologic sequelae and death. Altered consciousness, disorientation, and seizures occur within a few days after the onset of fever and respiratory symptoms (1–3). In some patients, symptoms are transient but in others rapid progression to necrotizing encephalitis, deep coma, and death may occur (1–3). Cases in adults are infrequently reported and remain poorly characterized, although the more complex clinical scenarios in adults may have hindered case recognition (1,4–6). The pathogenesis is unclear, but a hyperactivated cytokine response, rather than viral invasion, is believed responsible in most childhood cases (1–5). We describe 3 cases of acute encephalopathy associated with influenza A infection in adults. The clinical, virologic, immunologic findings (cytokines in plasma and cerebrospinal fluid [CSF]), and CSF penetration of oseltamivir for these cases are reported.

Author affiliations: The Chinese University of Hong Kong, Hong Kong Special Administrative Region, People's Republic of China (N. Lee, C.K. Wong, P.K.S. Chan, E. Wong, K.S. Wong, C.S. Cockram, J.J.Y. Sung, D.S.C. Hui); Mahidol University, Bangkok, Thailand (N. Lindegardh, N.J. White); University of Oxford, Oxford, UK (N. Lindegardh, N.J. White); and University of Virginia School of Medicine, Charlottesville, Virginia, USA (F.G. Hayden)

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The Study

At Prince of Wales Hospital, Hong Kong (7), from January 2007 through August 2008, influenza infection was diagnosed for >460 hospitalized adult patients for whom acute febrile respiratory illnesses had been diagnosed. Nasopharyngeal aspiration and immunofluorescence assays (IFA) were used for rapid diagnosis of influenza A and B infection, confirmed by virus isolation. Thirteen (2.8%) patients had signs of confusion or altered consciousness, together with fever and respiratory symptoms (mean \pm SD age 77.7 ± 8.8 years). We studied 3 patients from whom CSF was obtained for analysis, and who fulfilled the definition of influenza-associated acute encephalopathy (altered mental status >24 hours within 5 days of influenza onset and without alternative explanation) (1,2,4–6).

Nasopharyngeal aspirates were subjected to IFA, virus isolation, and subsequent subtyping (7). CSF specimens were subjected to virus isolation using MDCK cells, and reverse transcription–PCR to detect influenza virus RNA by using H1/H3 subtype-specific primers. Herpes simplex virus, herpes zoster virus, and enterovirus DNA/RNA was detected using PCRs (online Technical Appendix, available from www.cdc.gov/EID/content/16/1/139-Techapp.pdf).

CSF and plasma samples collected on the same day were analyzed simultaneously for the concentrations of 11 cytokines/chemokines by bead-based multiplex flow cytometry. Their assay methods and plasma reference ranges (established from >100 healthy persons) have been described (online Technical Appendix) (7). In CSF, in patients without central nervous system (CNS) disease/infection, cytokines/chemokines are either undetectable (e.g., interleukin-6 [IL-6], CXCL8/IL-8, CXCL10/IP-10, CXCL9/MIG) or present at low levels (e.g., CCL2/MCP-1) (8–10).

Concentrations of oseltamivir phosphate (OP) and its biologically active metabolite oseltamivir carboxylate (OC) were measured in CSF and plasma taken simultaneously from 1 patient who received concurrent treatment, using tandem mass spectrometry (11). The assay methods have been described (online Technical Appendix).

The clinical and virologic findings are summarized in Table 1. All case-patients were elderly (72–86 years of age), but none were known to have neuropsychiatric illness, dementia, or to be taking psychotropic medication. None had received updated influenza vaccination (6). Confusion and altered consciousness developed in patients 1 and 2 one to 2 days after the onset of fever and cough. These patients had no meningismus, focal neurologic deficit, hypotension, respiratory distress, or metabolic disturbances. Brain computed tomography (CT) scans showed no acute cerebral lesion. CSF analyses showed no bacterial or viral pathogen or pleocytosis. Oseltamivir was given to patient 2 only when influenza A was later confirmed

Table 1. Clinical and laboratory findings in 3 patients with acute encephalopathy associated with influenza infection, Prince of Wales Hospital, Hong Kong*

Clinical and laboratory findings	Patient 1	Patient 2	Patient 3
Age, y/sex	76/M	86/F	72/M
Concurrent illnesses	Ischemic heart disease	Diabetes mellitus, hypertension	COPD
Influenza vaccination within 6 mo	None	None	None
Symptoms on examination	Fever >38°C, cough, disorientation, incoherent speech, mental dullness	Fever >38°C, cough, delirious, impaired consciousness, did not follow verbal command	Fever >38°C, cough, disorientation, agitation, incoherent speech, involuntary 4-limb tremor
Focal neurologic sign or meningism	Absent	Absent	Absent
Chest radiograph, consolidation	Absent	Absent	Absent
Antiviral (oseltamivir)	None	Given	Given
Outcome (duration of encephalopathy)	Recovered (2–3 d)	Recovered (3–4 d)	Recovered (6–7 d)
Brain CT scan (noncontrast)	Normal	Old ischemic changes; known small, calcified meningioma	Normal
Virus isolated from NPA	Seasonal (H1N1) 2008	Subtype H3N2	Subtype H3N2
CSF testing results			
Opening pressure, cm H ₂ O	11	9	14
Cell count (x 10 ⁶ /L)	1	–	0
Glucose, mmol/L	4.2	7.4	3.7
Protein, g/L	0.46	0.47	0.16
Virus isolated	None	None	None
RT-PCR for H3 and H1 influenza virus	Negative	Negative	Negative
Bacterial culture	Negative	Negative	Negative
Others	HSV, HZV, and enterovirus PCR negative	HSV, HZV, and enterovirus PCR negative	HSV PCR negative

*COPD, chronic obstructive pulmonary disease; CT, computed tomographic scan; NPA, nasopharyngeal aspirate; CSF, cerebrospinal fluid; RT-PCR, reverse transcription-PCR; HSV, herpes simplex virus; HZV, herpes zoster virus. In all cases, there was no hypoglycemia, and liver and renal function test results were normal. C-reactive protein level was elevated in all cases. For patient 3, an electroencephalogram was performed and showed generalized slowing of background consistent with moderate encephalopathic change (similar to that observed in septic encephalopathy) (1,6). Findings are consistent with previous reports on adult cases of influenza-associated encephalopathy: patients are all unvaccinated, pleocytosis and cerebral imaging abnormalities (even with magnetic resonance imaging) are usually absent, and symptoms are generally self-limiting (1,6). Most reports have mentioned influenza A as a cause of encephalopathy, and more commonly subtype H3N2 (1–6).

by nasopharyngeal aspirate/IFA; patient 1 did not receive antiviral treatment. Both patients recovered in the next 2 days. Patient 3 had fever, severe chronic obstructive pulmonary disease exacerbation requiring noninvasive ventilatory support, complicated by acute coronary syndrome. He was given oseltamivir, 75 mg 2×/day, after influenza A infection was confirmed. Agitation and confusion developed in the patient on day 3–4 of illness (onset after the third dose of oseltamivir), despite resolution of the patient's respiratory failure. These symptoms were followed by involuntary, tremulous movements involving all 4 limbs, while at rest and during movement. Brain CT scan was normal. Electroencephalogram showed generalized slowing. Oseltamivir was stopped after the ninth dose, but tremor persisted. CSF analyses showed no pathogen or pleocytosis. The patient's symptoms resolved in the next 3–4 days without sequelae.

Despite apparently normal CSF findings, high concentrations of cytokines/chemokines were detected in the CSF and plasma specimens of all patients (Table 2). Plasma concentrations of IL-6, CXCL8/IL-8, CXCL10/IP-10, CCL2/MCP-1, and CXCL9/MIG were elevated at median values of 2.0, 2.8, 11.9, 3.7, and 2.1× the upper limits of their re-

spective reference ranges (comparable to or higher than that observed in other hospitalized influenza patients) (Table 2) (7). Other cytokines were not elevated (4,7). In their CSF, IL-6, CXCL8/IL-8, CXCL10/IP-10, and CCL2/MCP-1 were consistently detected, and were elevated at median values of 2.6, 15.0, 3.4, and 20.0 × the upper limits of their respective plasma reference ranges. The CSF/plasma concentration ratios of CXCL8/IL-8 and CCL2/MCP-1 were >3 (median CSF/plasma ratio 5.4 and 8.0, respectively).

Simultaneous CSF and plasma OC and OP concentrations were determined for patient 3, as symptoms progressed at 18 h after oseltamivir. The concentrations (mean ± SD) of OC in duplicate CSF and plasma samples were 18.3 ± 0.9 ng/mL and 143.8 ± 3.3 ng/mL, respectively; the CSF/plasma concentration ratio was 12%–13%. The OP plasma concentration was 1.05 ± 0.03 ng/mL; it was not detectable in the CSF.

Conclusions

We report 3 adults with acute encephalopathy (altered consciousness, confusion) associated with influenza. High CSF and blood cytokine/chemokine (CXCL8/IL-8, CCL2/MCP-1, IL-6, CXCL10/IP-10) levels were detected. No ev-

Table 2. Cytokine and chemokine concentrations in CSF and plasma samples from 3 patients with acute encephalopathy associated with influenza A infection, Prince of Wales Hospital, Hong Kong*

Cytokine or chemokine	Reference range, pg/mL	CSF/plasma cytokine concentration, pg/mL (ratio)		
		Patient 1	Patient 2	Patient 3
IL-6†	<3.1	8.0/6.3 (1.3)	11.6/35.1 (0.3)	2.2/5.9 (0.4)
CXCL8/IL-8‡	<5.0	84.0/15.5 (5.4)	74.8/13.8 (5.4)	21.9/6.3 (3.5)
CXCL10/IP-10†	202–1,480	15,374/102,019 (0.2)	5,101/17,594 (0.3)	1,371/1,550 (0.9)
CCL2/MCP-1‡	< 10-57	996/82 (12.1)	1,287/336 (3.8)	–
CXCL9/MIG	48–482	11,58/14,001 (0.1)	70/333 (0.2)	145/1,019 (0.1)
IFN-γ	<15.6	UD/14.4	4.7/10.1	0.4/2.0
IL-12p70	<7.8	1.5/UD	1.3/UD	UD/UD
TNF-α	<10.0	1.7/1.4	UD/1.2	UD/UD
IL-10	<7.8	2.5/2.2	UD/7.3	UD/1.7
IL-1β	<3.9	UD/UD	UD/3.7	UD/UD
CCL5/RANTES	4,382–18,783	4/2,507	14/1,609	1.3/814

*CSF, cerebrospinal fluid; –, test not done due to inadequate sample; UD, undetectable (i.e., below the detection limit of the cytokine/chemokine assay). Cytokines: Interleukin (IL)–1β, IL-6, IL-10, IL-12p70, tumor necrosis factor α (TNF-α). Chemokines: CXCL8/IL-8, monokine induced by interferon-γ (IFN-γ) (CXCL9/MIG), IFN-γ-inducible protein-10 (CXCL10/IP-10), monocyte chemoattractant protein-1 (CCL2/MCP-1), and regulated upon activation normal T cell-expressed and secreted (CCL5/RANTES). The plasma reference ranges are established from >100 healthy adults. The assay sensitivities of IL-1β, IL-6, IL-10, IL-12p70, TNF-α, IL8, MIG, IP-10, MCP-1, RANTES, and IFN-γ are 2.5, 3.3, 3.7, 1.9, 7.2, 0.2, 2.5, 2.8, 2.7, 1.0, and 7.1 pg/mL, respectively. Coefficients of variation are all <10%. In an earlier study involving 39 adult influenza patients hospitalized with cardio-respiratory complications (7), the median (interquartile range) plasma concentrations of IL-6, IL-8, IP-10, MCP-1, and MIG were 10.6 (4.2–18.4), 5.4 (2.5–8.7), 7,043.0 (4,025.1–1,2381.1), 76.5 (49.5–97.0), and 992.1 (499.1–1,992.3) pg/mL, respectively. In CSF, in subjects without neurologic disease/infection, these cytokines/chemokines are either undetectable or present at low levels (8–10). In a pediatric influenza cohort, CSF cytokine levels were substantially higher in encephalopathy cases when compared to those with febrile seizure; CSF/plasma concentration was <1 (8).

†CSF cytokine concentrations above plasma reference ranges.

‡CSF/plasma cytokine concentration ratio consistently >3 (3.5–12.1), in addition to CSF cytokine concentrations being above the plasma reference ranges. For IFN-γ, IL-12p70, TNF-α, IL-10, IL-1β and RANTES, because of their low/undetectable levels, the CSF/plasma ratios were not calculated. CSF specimens from patients 1 and 2 were collected at the peak of symptoms, and before antiviral treatment (if given); CSF from patient 3 was collected when persistent tremor developed 18 hours after the ninth dose of oseltamivir; the drug was stopped afterward.

idence of direct viral neuroinvasion was found. All patients recovered rapidly without sequelae (1,6).

Our findings agree with studies of influenza-associated encephalopathy in children. Influenza virus is rarely detected in the CSF, and pleocytosis is often absent (1,2,4–6). High levels of cytokines (e.g., IL-6, soluble tumor necrosis factor receptor 1) can be consistently found in CSF/blood specimens, correlating with disease severity and outcomes (hyperactivated cytokine response is absent in febrile seizure associated with influenza) (2–4,8). We found a broader range of cytokines/chemokines being activated (7); for certain cytokines (CXCL8/IL-8, CCL2/MCP-1), the CSF concentrations were 3× those in plasma. IL-6, CXCL8/IL-8, CCL2/MCP-1 and CXCL10/IP-10 have been shown to play pathogenic roles in CNS viral infections, cerebral injury, and acute brain syndrome in susceptible patients (7,9,10,12). The high CSF/plasma ratios suggest that for some cytokines, activation within the CNS might have occurred along with respiratory-tract and systemic productions (cytokines are not detected in CSF normally; (Table 2) (4,7–10,12). Resident macrophages/monocytes, astrocytes, microglial and endothelial cells in the CNS are shown to release cytokines/chemokines when stimulated by viral/influenza infection; activation mechanisms without involving overt CNS invasion have been suggested (1,4,9,12–14). Cytokines may cause direct neurotoxic effects, cerebral metabolism changes, or breakdown of the blood-brain-barrier (endothelial injury) to produce symptoms (1–4,8,12–14).

Whether early viral suppression by antivirals can lead to attenuation of these cytokine responses and better outcomes warrants further study (7).

We measured oseltamivir concentrations because of the concerns over its neuropsychiatric side-effects in children and adolescents. However, only the active metabolite (OC) was detected in the CSF of patient 3; the CSF/plasma concentration ratio was 12%–13% (18.3/143.8 ng/mL) at 18-hours postdose. This degree of CSF penetration is similar to that observed among healthy patients, with a Cmax CSF/plasma concentration ratio of 3.5% (at ≈8 hours), and a ratio of ≈10% at 18 hours (concentration-time profiles for plasma/CSF differ). Assuming a similar ratio, the CSF OP concentration would have fallen below the assay's detection limit (0.25 ng/mL) by 18 hours (11,15). The low CSF drug-penetration, together with high cytokines in CSF and symptom progression despite drug withdrawal suggest that the manifestations of patient 3 may have been disease-related. Symptoms developed in patients 1 and 2 without antiviral exposure. Further investigations on the CNS effects of oseltamivir in the clinical setting are needed.

Our study is limited by the small patient number and the lack of feasibility in obtaining CSF for study/comparison in influenza patients without neurologic symptoms. Further studies on the clinical spectrum of influenza encephalopathy and encephalitis in adults (1,6) and their pathogenesis are indicated. In conclusion, acute encephalopathy may occur in adults with influenza. Exuberant

cytokine/chemokine response may play an important role in its pathogenesis.

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Dr Lee is an associate professor in infectious diseases at The Chinese University of Hong Kong. His primary research interest is severe emerging infections, including severe acute respiratory syndrome and influenza.

References

1. Studahl M. Influenza virus and CNS manifestations. *J Clin Virol*. 2003;28:225–32. DOI: 10.1016/S1386-6532(03)00119-7
2. Morishima T, Togashi T, Yokota S, Okuno Y, Miyazaki C, Tashiro M, et al.; Collaborative Study Group on Influenza-Associated Encephalopathy in Japan. Encephalitis and encephalopathy associated with an influenza epidemic in Japan. *Clin Infect Dis*. 2002;35:512–7. DOI: 10.1086/341407
3. Togashi T, Matsuzono Y, Narita M, Morishima T. Influenza-associated acute encephalopathy in Japanese children in 1994–2002. *Virus Res*. 2004;103:75–8. DOI: 10.1016/j.virusres.2004.02.016
4. Kawada J, Kimura H, Ito Y, Hara S, Iriyama M, Yoshikawa T, et al. Systemic cytokine responses in patients with influenza-associated encephalopathy. *J Infect Dis*. 2003;188:690–8. DOI: 10.1086/377101
5. van Zeijl JH, Bakkens J, Wilbrink B, Melchers WJ, Mullaart RA, Galama JM. Influenza-associated encephalopathy: no evidence for neuroinvasion by influenza virus nor for reactivation of human herpesvirus 6 or 7. *Clin Infect Dis*. 2005;40:483–5. DOI: 10.1086/427027
6. Steininger C, Popow-Kraupp T, Laferl H, Seiser A, Gödl I, Djamshidian S, et al. Acute encephalopathy associated with influenza A virus infection. *Clin Infect Dis*. 2003;36:567–74. DOI: 10.1086/367623
7. Lee N, Wong CK, Chan PKS, Lun SWM, Lui G, Wong B, et al. Hypercytokinemia and hyperactivation of phospho-p38 mitogen-activated protein kinase in severe human influenza A infections. *Clin Infect Dis*. 2007;45:723–31. DOI: 10.1086/520981
8. Ichiyama T, Morishima T, Isumi H, Matsufuji H, Matsubara T, Furukawa S. Analysis of cytokine levels and NF-kappaB activation in peripheral blood mononuclear cells in influenza virus-associated encephalopathy. *Cytokine*. 2004;27:31–7. DOI: 10.1016/j.cyto.2004.03.012
9. Lahrtz F, Piali L, Spanaus KS, Seebach J, Fontana A. Chemokines and chemotaxis of leukocytes in infectious meningitis. *J Neuroimmunol*. 1998;85:33–43. DOI: 10.1016/S0165-5728(97)00267-1
10. Sprenger H, Rösler A, Tonn P, Braune HJ, Huffmann G, Gemsa D. Chemokines in the cerebrospinal fluid of patients with meningitis. *Clin Immunol Immunopathol*. 1996;80:155–61. DOI: 10.1006/clin.1996.0109
11. Lindegårdh N, Hanpithakpong W, Wattanagoon Y, Singhasivanon P, White NJ, Day NP. Development and validation of a liquid chromatographic-tandem mass spectrometric method for determination of oseltamivir and its metabolite oseltamivir carboxylate in plasma, saliva and urine. *J Chromatogr B Analyt Technol Biomed Life Sci*. 2007;859:74–83. DOI: 10.1016/j.jchromb.2007.09.018
12. Semmler A, Hermann S, Mormann F, Weberpals M, Paxian SA, Okulla T, et al. Sepsis causes neuroinflammation and concomitant decrease of cerebral metabolism. *J Neuroinflammation*. 2008;5:38.
13. Wang G, Zhang J, Li W, Xin G, Su Y, Gao Y, et al. Apoptosis and proinflammatory cytokine responses of primary mouse microglia and astrocytes induced by human H1N1 and avian H5N1 influenza viruses. *Cell Mol Immunol*. 2008;5:113–20.
14. Yokota S, Imagawa T, Miyamae T, Ito S, Nakajima S, Nezu A, et al. Hypothetical pathophysiology of acute encephalopathy and encephalitis related to influenza virus infection and hypothermia therapy. *Pediatr Int*. 2000;42:197–203. DOI: 10.1046/j.1442-200-x.2000.01204.x
15. Jhee SS, Yen M, Ereshefsky L, Leibowitz M, Schulte M, Kaeser B, et al. Low penetration of oseltamivir and its carboxylate into cerebrospinal fluid in healthy Japanese and caucasian volunteers. *Antimicrob Agents Chemother*. 2008;52:3687–93. DOI: 10.1128/AAC.00327-08

Address for correspondence: Nelson Lee, Division of Infectious Diseases, Department of Medicine and Therapeutics, Prince of Wales Hospital, The Chinese University of Hong Kong, 9/F Clinical Sciences Building, Department of Medicine and Therapeutics, Shatin, Hong Kong, People's Republic of China; email: leelsn@cuhk.edu.hk

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Technical Appendix

Methods on Virus Detection and the Measurements of Cytokine and Oseltamivir Concentrations

Section I. Methods of Virus Detection

Influenza viral RNA was detected using H1/H3 subtype-specific primers (1,2) (H1: 5'-TGA GGG AGC AAT TGA GTT CA-3' and 5'-TGC CTC AAA TAT TAT TGT GT-3'; H3: 5'-GCA ACT GTT ACC CTT ATG AT-3' and 5'-TCA TTG TTT GGC ATA GTC AC-3'). After RNA extraction, a 10- μ L aliquot of the extracted preparation was mixed with 15 μ L of reverse transcription-PCR (RT-PCR) master mix, which containing 12.5 μ L of 2 \times reaction mix, 0.5 μ L of SuperScript III RT/ Platinum Taq Mix (Invitrogen, Carlsbad, CA, USA), 5 pmole of each forward and reverse primer, and adjusted to a volume of 15 μ L with nuclease-free distilled water. For influenza A H1, the reaction condition was 30 min at 45°C for reverse transcription followed by 2 min at 94°C, and 30 cycles of 1 min at 94°C, 1 min at 52°C and 1 min at 68°C; and a final extension of 10 min at 68°C. The reaction condition for influenza A H3 was similar, except that reverse transcription was carried out at 50°C and the annealing temperature was 55°C. The PCR products were separated by gel electrophoresis and visualized by SYBR safe DNA stain (Invitrogen) under gel documentation system. The product size of influenza A H1 was 431 bp; the influenza A H3 product was 232 bp.

Enterovirus RNA was detected by one-step real-time RT-PCR (3,4). Amplification is specific for the 5' untranslated region. Enterovirus RNA extracted from cell-culture fluid of clinical isolate confirmed by immunofluorescent was used as the source of positive control. RNA from the plasmid pAW109 was used as internal control for nucleic acid extraction as well as for the presence of inhibitors in the clinical samples. Primers and probes used were as follows: primer for enterovirus (EV), 5'-ACATGGTGTGAAGAGTCTATTGAGCT-3', 5'-CCAAAGTAGTCGGTCCGC-3'; primer for pAW, 5'-GCC TGG GTT CCC TGT TCC-3', 5'-CGA CGT ACC CCT GAC ATG G-3'; probe for EV, 5'-(FAM)TCCGGCCCCTGAATGCGGCTAAT-3' (TAMARA); and probe for pAW, 5'-(TET)CAGGCCAATGTCTCACCAAGC TCT-3' (TAMARA).

Herpes Simplex Virus I (HSV1) was detected by real-time PCR (5). Amplification is specific for glycoprotein D. DNA extracted from cell-culture fluid confirmed positive for HSV-1 by immunofluorescent staining was used as the source of positive control. TaqMan Exogenous Internal Positive Control (EXO-IPC) DNA (Applied Biosystems, Foster City, CA, USA) was used as internal control for nucleic acid extraction as well as for the presence of inhibitors in the clinical samples. Primers used were 5'-CGGCCGTGTGACACTATCG-3', 5'-CTCGTAAAATGGCCCCTCC-3'; TaqMan Probe: 5'FAM-CCATACCGACCACACCGACGAACC-TAMRA-3'.

Herpes simplex virus 2 (HSV2) was detected by real-time PCR (5). Amplification is specific for glycoprotein G. DNA extracted from cell-culture fluid confirmed positive for HSV2 by immunofluorescent staining was used as the source of positive control. TaqMan Exogenous Internal Positive Control (EXO-IPC) DNA (Applied Biosystems) was used as internal control for nucleic acid extraction as well as for the presence of inhibitors in the clinical samples. Primers

used were 5'-CGCTCTCGTAAATGCTTCCCT-3' and 5'-TCTACCCACAACAGACCCACG-3'
(4)

Varicella zoster virus (VZV) was detected by real-time PCR (5,6). Amplification is specific for the diploid gene open reading frame 62 encoding for the major immediate transactivator. DNA extracted from cell-culture fluid of the VZV vaccine strain VarilRix (GlaxoSmithKline, Rixelsart, Belgium) or from clinical specimens confirmed positive for VZV by immunofluorescent staining was used as the source of positive control. TaqMan Exogenous Internal Positive Control (EXO-IPC) DNA (Applied Biosystems) was used as internal control for nucleic acid extraction as well as for the presence of inhibitors in the clinical samples. Primers used were 5'-TCTTGTCGAGGAGGCTTCTG-3', 5'-TGTGTGTCCACCGGATGAT-3'; TaqMan Probe: 5'-TET-TCTCGACTGGCTGGGACTTGCG-TAMARA-3'.

Section 2. Measurement of Cytokine Concentration

Plasma concentrations of 10 cytokines and chemokines: interleukin(IL)-1 β , IL-6, IL-10, IL-12p70, tumor-necrosis-factor (TNF)- α , CXCL8/IL-8, monokine induced by interferon- γ (IFN- γ) (CXCL9/MIG), IFN- γ -inducible protein-10 (CXCL10/IP-10), monocyte chemoattractant protein-1 (CCL2/MCP-1), and regulated upon activation normal T cell-expressed and secreted (CCL5/RANTES), were assayed using cytometric bead array (CBA) reagents (BD Pharmingen, San Diego, CA, USA) with 4-color FACSCalibur flow cytometer (BD Biosciences Corp, San Jose, CA, USA) (2). In CBA, different bead populations with distinct fluorescence intensities had been coated with capturing antibodies specific for different cytokines or chemokines. After incubation with 50 μ L of plasma/cerebrospinal fluid (CSF), the cytokine/chemokine captured beads were mixed with phycoerythrin-conjugated detection antibodies to form sandwich complexes. Fluorescence flow cytometry of the beads provided simultaneous quantification of a panel of

cytokines and chemokines. IFN- γ was quantified by an ELISA (R & D Systems Inc., Minneapolis, MN, USA).

Section 3. Measurement of Oseltamivir Concentration

Oseltamivir concentration assays were performed by the Clinical Pharmacology Laboratory in the Faculty of Tropical Medicine, Bangkok, by using tandem mass spectrometry (7). Plasma and CSF samples were analyzed by using mixed mode (MPC-SD, 3M Empore Bracknell, UK) solid phase extraction and HILIC-LC-MS/MS. Stable isotope labeled (i.e., deuterated oseltamivir phosphate [OP] and oseltamivir carboxylate [OC]) internal standards were added to the samples before solid phase extraction. The drugs were quantified using an API 5000 triple quadrupole mass spectrometer (Applied Biosystems/MDS SCIEX, Foster City, CA, USA), with a TurboVTM ionisation source interface operated in the positive ion mode. Quantification was performed using selected reaction monitoring for the transitions m/z 313 – 225 and 316 – 228 for OP and deuterated OP, respectively, and 285 – 197 and 288 – 200 for OC and deuterated OC, respectively. The performance of the assay was demonstrated by analysis of 3 replicates of quality control samples at 3 levels. The coefficients of variation (CV%) during the analysis of OP were 4.1%, 1.9% and 2.1% at 3 ng/mL, 20 ng/mL, and 150 ng/mL, respectively (limit of detection of OP, 0.25 ng/mL). The coefficients of variation (CV%) during the analysis of OC were 2.3%, 0.5%, and 3.1% at 30 ng/mL, 400 ng/mL, and 4,000 ng/mL, respectively.

References

1. Lee N, Chan PKS, Choi KW, Lui G, Wong B, Cockram CS, et al. Factors associated with early hospital discharge of adult influenza patients. *Antivir Ther.* 2007;12:501–8. [PubMed](#)

2. Lee N, Wong CK, Chan PKS, Lun SWM, Lui G, Wong B, et al. Hypercytokinemia and hyperactivation of phospho-p38 mitogen-activated protein kinase in severe human influenza A infections. *Clin Infect Dis.* 2007;45:723–31. [PubMed DOI: 10.1086/520981](#)
3. Dierssen U, Rehren F, Henke-Gendo C, Harste G, Heim A. Rapid routine detection of enterovirus RNA in cerebrospinal fluid by a one-step real-time RT-PCR assay. *J Clin Virol.* 2008;42:58–64. [PubMed DOI: 10.1016/j.jcv.2007.11.016](#)
4. Monpoeho S, Coste-Burel M, Costa-Mattioli M, Besse B, Chomel JJ, Billaudel S, et al. Application of a real-time polymerase chain reaction with internal positive control for detection and quantification of enterovirus in cerebrospinal fluid. *Eur J Clin Microbiol Infect Dis.* 2002;21:532–6. [PubMed DOI: 10.1007/s10096-002-0766-5](#)
5. Weidmann M, Meyer-König U, Hufert FT. Rapid detection of herpes simplex virus and varicella-zoster virus infections by real-time PCR. *J Clin Microbiol.* 2003;41:1565–8. [PubMed DOI: 10.1128/JCM.41.4.1565-1568.2003](#)
6. Wang K, Lau TY, Morales M, Mont EK, Straus SE. Laser-capture microdissection: refining estimates of the quantity and distribution of latent herpes simplex virus 1 and varicella-zoster virus DNA in human trigeminal ganglia at the single-cell level. *J Virol.* 2005;79:14079–87. [PubMed DOI: 10.1128/JVI.79.22.14079-14087.2005](#)
7. Lindegårdh N, Hanpithakpong W, Wattanagoon Y, Singhasivanon P, White NJ, Day NP. Development and validation of a liquid chromatographic-tandem mass spectrometric method for determination of oseltamivir and its metabolite oseltamivir carboxylate in plasma, saliva and urine. *J Chromatogr B Analyt Technol Biomed Life Sci.* 2007;859:74–83. [PubMed DOI: 10.1016/j.jchromb.2007.09.018](#)