

Novel Respiratory Virus Infections in Children, Brazil

Maria Carolina M. Albuquerque, Gisele P.A. Pena, Rafael B. Varella, George Gallucci, Dean Erdman, and Norma Santos

Recently discovered respiratory viruses were detected in 19 (9.2%) of 205 nasal swab specimens from children in Brazil with respiratory illnesses. Five each were positive for human metapneumovirus (HMPV) alone and human bocavirus (HBoV) alone, 3 for human coronaviruses (HCoV-HKU1 or -NL63) alone, and 6 for more than 1 recently discovered virus.

Viral infections are among the leading causes of respiratory disease in children. Most of these infections are caused by respiratory syncytial virus (RSV), influenza virus A or B (FluV), parainfluenza virus (PIV), rhinovirus (RV), or adenovirus (AdV). Several recently discovered viruses, such as human metapneumovirus (HMPV), human bocavirus (HBoV), and the human coronaviruses (HCoVs) NL63 and HKU1, have been identified as potential respiratory pathogens (1). In addition, 2 new human polyomaviruses (HPyVs), KIPyV and WUPyV, have been detected in patients with respiratory infections (1). In Brazil, epidemiologic studies have demonstrated the extent to which viruses cause respiratory illness in children (2–4). However, because such studies have focused on the most common viral pathogens, the extent to which the novel respiratory viruses are etiologic agents of respiratory disease in Brazilian children remains unknown. In this study, we sought to investigate the occurrence of respiratory infections associated with HMPV, HBoV, HCoV-HKU1, HCoV-NL63, KIPyV, and WUPyV among children in Brazil.

The Study

The study protocol was reviewed and approved by the research ethics committees of the Institute of Puericulture and Pediatrics Martagão Gesteira of the Federal University of Rio de Janeiro and the Educational Foundation of Serra dos Órgãos of Teresópolis. The parents of all children involved in the study gave informed consent for their chil-

dren's participation in accordance with Resolution 196/96 of the Brazilian Ministry of Health.

Nasal swabs from 205 children (median age 3.3 years; range 1 month to 15 years) with acute upper or lower respiratory illnesses were collected from March 2006 through October 2007 and tested for viral pathogens. Acute respiratory illness was defined by the presence of rhinorrhea, cough, respiratory distress, or sore throat, associated or not with fever, for a maximum duration of 7 days. The specimens were collected from hospitalized patients, emergency departments, and walk-in clinics at 2 university hospitals in the cities of Rio de Janeiro and Teresópolis. Relevant clinical information, including patients' hospitalization status, age, sex, and clinical symptoms, was collected during the first medical visit by means of a standard questionnaire.

The nasal swabs were immersed in 1 mL of virus transport media and kept at -70°C until processing. Nucleic acid was extracted from 200 μL of the sample by using the Wizard Genomic DNA Purification kit (Promega, Madison, WI, USA) or RNAagents kit (Promega) according to the manufacturer's instructions. Specimens were tested for presence of FluV A and B (5), PIV 1–4 (6), AdV (7), RSV (8), RV (8), HMPV (9), HBoV (10), WUPyV (11), and KIPyV (12) by conventional PCR assays as previously described. A real-time PCR protocol was used for detection of HCoVs (229E, OC43, NL63, and HKU1) (13).

Of the 205 samples tested, 63 (30.7%) were positive for at least 1 of the viral pathogens specified above. Nineteen (9.2%) were positive for at least 1 of the newly described viruses: 5 for HMPV only, 5 for HBoV only, 3 for HCoV-HKU1 or HCoV-NL63 only, and 6 for co-infections with these viruses, including 2 samples positive for KIPyV or WUPyV. Of the samples positive for common respiratory viruses, 33 were positive for rhinovirus only, 5 for FluV A only, 3 for RSV only, and 1 each for HCoV-OC43 and AdV only. Two samples were positive for >1 common respiratory viruses, and PIV was not detected (online Appendix Table, available from www.cdc.gov/EID/content/15/5/806-appT.htm). The age of the patients infected with the newly described viruses ranged from 4 months to 11 years (median 2.7 years). The most frequent clinical symptoms were fever, rhinorrhea, cough, sore throat, wheezing, bronchiolitis, and pneumonia (Table). Although the specimens were collected from symptomatic children and a wide range of viruses were screened for by highly sensitive methods, less than one third of the samples were positive. Perhaps if we had collected nasopharyngeal swabs or aspirates, the percentage of samples that tested positive would have been higher.

The diagnosis for most patients infected with HCoV-NL63 or HCoV-HKU1 was pneumonia. Of the 3 patients with single infections, 2 were hospitalized and 1 was treated at the emergency department; the 2 patients with mixed

Author affiliations: Universidade Federal do Rio de Janeiro, Rio de Janeiro, Brazil (M.C.M. Albuquerque, G.P.A. Pena, N. Santos); Centro Universitário Serra dos Órgãos, Teresópolis, Rio de Janeiro (R. B. Varella); and Centers for Disease Control and Prevention, Atlanta, Georgia, USA (G. Gallucci, D. Erdman)

DOI: 10.3201/eid1505.081603

Table. Clinical symptoms observed among patients with a single infection, by virus, Brazil, 2006–2007*

Clinical symptom	% Patients (no. positive/no. tested)							
	HCoV-HKU1 or NL63	HMPV	HBoV	RV	FluV A	RSV	HCoV-OC43	AdV
Fever	100 (3/3)	80 (4/5)	80 (4/5)	52 (17/33)	40 (2/5)	100 (3/3)	0 (0/1)	100 (1/1)
Rhinorrhea	33 (1/3)	60 (3/5)	60 (3/5)	67 (22/33)	80 (4/5)	100 (3/3)	0 (0/1)	0 (0/1)
Cough	33 (1/3)	100 (5/5)	80 (4/5)	67 (22/33)	40 (2/5)	33 (1/3)	0 (0/1)	100 (1/1)
Sore throat	0 (0/3)	20 (1/5)	20 (1/5)	6 (2/33)	60 (3/5)	0 (0/3)	0 (0/1)	0 (0/1)
Wheezing	33 (1/3)	40 (2/5)	80 (4/5)	27 (9/33)	20 (1/5)	0 (0/3)	0 (0/1)	100 (1/1)
Bronchiolitis	33 (1/3)	20 (1/5)	0 (0/5)	12 (4/33)	0 (0/5)	0 (0/3)	100 (1/1)	0 (0/1)
Pneumonia	100 (3/3)	0 (0/5)	40 (2/5)	27 (9/33)	20 (1/5)	0 (0/3)	0 (0/1)	0 (0/1)

*HCoV, human coronavirus; HMPV, human metapneumovirus; HBoV, human bocavirus; RV, rhinovirus; FluV A, influenza virus A; RSV, respiratory syncytial virus; AdV, adenovirus.

infection of HCoV-NL63 and HMPV or RV were treated at walk-in clinics.

Patients with HMPV infections had a myriad of symptoms, including fever, cough, rhinorrhea, wheezing, and sore throat. Of the 5 patients with single infections, 4 were treated at walk-in clinics and 1 was treated at an emergency department. A patient co-infected with HMPV and HCoV-NL63 was treated at a walk-in clinic, and a patient co-infected with HMPV and KIPyV and 1 co-infected with HMPV, HCoV-OC43, AdV, and RV were treated at an emergency department. The patient co-infected with KIPyV was a 4-year-old boy with cough, fever, rhinorrhea, and wheezing.

HBoV was detected in samples from 5 patients as a single infection and in samples from 2 patients as a co-infection with RV or WUPyV. Three patients had pneumonia (2 single infections and 1 co-infection with RV). Two of the 3 were treated at walk-in clinics; 1 of the patients with only HBoV infection was hospitalized. The patient co-infected with WUPyV was a 10-month-old boy who had been treated at an emergency department after exhibiting cough, rhinorrhea, and laryngomalacia.

Conclusions

Previous studies have documented the importance of respiratory virus infections among pediatric patients in Brazil (2–4). However, the effect of the so-called emerging respiratory viruses on the children of Brazil is yet to be clarified. Few studies have demonstrated the circulation of HMPV among Brazilian children (4,14), and to our knowledge, none have described the circulation of HBoV, HCoV, or HPyVs as respiratory pathogens in Brazil, although 1 study did report the presence of HBoV in the stools of Brazilian children with gastroenteritis (15). Our finding that HMPV, HBoV, HCoV-HKU1 or HCoV-NL63, or the newly described KIPyV or WUPyV was present in 9.2% of the tested samples suggests that these viruses could be important respiratory pathogens in the country. However, further investigative studies that include appropriately matched control groups will be necessary to demonstrate that these novel viruses act as etiologic agents of respiratory disease in Brazil.

Acknowledgments

We thank Shifaq Kamili and Soluza dos Santos Gonçalves for their technical assistance.

This manuscript represents a portion of a thesis submitted by M.C.M.A. to the Universidade Federal do Rio de Janeiro, Brazil, as a partial fulfillment of the requirements for a Doctor of Science degree.

This study was supported in part by Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq), Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES) and by Fundação de Amparo à Pesquisa do Estado do Rio de Janeiro (FAPERJ), Brazil.

Ms Albuquerque is a PhD candidate at Universidade Federal do Rio de Janeiro, Brazil. Her research interests include the diagnosis of enteric and respiratory infections and the epidemiology of enteric and respiratory viruses.

References

1. Sloots TP, Whitley DM, Lambert SB, Nissen MD. Emerging respiratory agents: new viruses for old diseases? *J Clin Virol*. 2008;42:233–43. DOI: 10.1016/j.jcv.2008.03.002
2. Tsuchiya LR, Costa LMD, Raboni SM, Nogueira MB, Pereira LA, Rotta I, et al. Viral respiratory infection in Curitiba, Southern Brazil. *J Infect*. 2005;51:401–7. DOI: 10.1016/j.jinf.2004.12.002
3. Costa LF, Yokosawa J, Mantese OC, Oliveira TF, Silveira HL, Nepomuceno LL, et al. Respiratory viruses in children younger than five years old with acute respiratory disease from 2001 to 2004 in Uberlândia, MG, Brasil. *Mem Inst Oswaldo Cruz*. 2006;101:301–6. DOI: 10.1590/S0074-02762006000300014
4. Thomazelli LM, Vieira S, Leal AL, Sousa TS, Oliveira DB, Golono MA, et al. Surveillance of eight respiratory viruses in clinical samples of pediatric patients in southeast Brazil. *J Pediatr (Rio J)*. 2007;83:422–8.
5. Stockton J, Ellis JS, Saville M, Clewley JP, Zambon MC. Multiplex PCR for typing and subtyping influenza and respiratory syncytial viruses. *J Clin Microbiol*. 1998;36:2990–5.
6. Aguilar JC, Perez-Brena MP, Garcia ML, Cruz N, Erdman DD, Echevarria JE. Detection and identification of human parainfluenza viruses 1, 2, 3, and 4 in clinical samples of pediatric patients by multiplex reverse transcriptase-PCR. *J Clin Microbiol*. 2000;38:1191–5.
7. Allard A, Albinsson B, Wandell G. Rapid typing of adenoviruses by a general PCR combined with restriction endonuclease analysis. *J Clin Microbiol*. 2001;39:498–505. DOI: 10.1128/JCM.39.2.498-505.2001

8. Myatt TA, Johnston SJ, Rudnick S, Milton DK. Airborne rhinovirus detection and effect of ultraviolet irradiation on detection by a semi-nested RT-PCR assay. *BMC Public Health*. 2003;3:5. DOI: 10.1186/1471-2458-3-5
9. Mackay IM, Jacob KC, Woolhouse D, Waller K, Sirmis MW, Whiley DM, et al. Molecular assay for detection of human metapneumovirus. *J Clin Microbiol*. 2003;41:100–5. DOI: 10.1128/JCM.41.1.100-105.2003
10. Sloots TP, McErlean P, Speicher DJ, Arden K, Nissen MD, Mackay IA. Evidence of human coronavirus HKU1 and human bocavirus in Australian children. *J Clin Virol*. 2006;35:99–102. DOI: 10.1016/j.jcv.2005.09.008
11. Gaynor AM, Nissen MD, Whiley DM, Mackay IM, Lambert SB, Wu G, et al. Identification of a novel polyomavirus from patients with acute respiratory tract infections. *PLoS Pathog*. 2007;3:e64. DOI: 10.1371/journal.ppat.0030064
12. Allander T, Andreasson K, Gupta S, Bjerkner A, Bogdanovic G, Persson MA, et al. A newly reported human polyomavirus, KI virus, is present in the respiratory tract of Australian children. *J Virol*. 2007;81:4130–6. DOI: 10.1128/JVI.00028-07
13. Dare RK, Fry AM, Chittaganpitch M, Sawanpanyalert P, Olsen SJ, Erdman DD. Human coronavirus infections in rural Thailand: a comprehensive study using real-time reverse-transcription polymerase chain reaction assays. *J Infect Dis*. 2007;196:1321–8. DOI: 10.1086/521308
14. Cuevas LE, Ben Nsser AM, Dove W, Gurgel RQ, Greensill J, Hart CA. Human metapneumovirus and respiratory syncytial virus, Brazil. *Emerg Infect Dis*. 2003;9:1626–8.
15. Albuquerque MC, Rocha LN, Benati FJ, Soares CC, Maranhão AG, Ramírez ML, et al. Human bocavirus infection in children with gastroenteritis, Brazil. *Emerg Infect Dis*. 2007;13:1756–8.

Address for correspondence: Norma Santos, Departamento de Virologia, Instituto de Microbiologia, Universidade Federal do Rio de Janeiro, Cidade Universitária, CCS-BI. I, Ilha do Fundão, Rio de Janeiro-RJ, 21.941-590, Brazil; email: nsantos@micro.ufrj.br

EMERGING INFECTIOUS DISEASES

A Peer-Reviewed Journal Tracking and Analyzing Disease Trends

Vol.11, No.6, June 2005



Search
past issues

EID
online
www.cdc.gov/eid

Appendix Table. Clinical characteristics of patients infected with various respiratory viruses, Brazil, 2006–2007*

Virus (no. patients)	Age/sex	Status	Clinical symptoms
Single infections			
HCoV-HKU1 (1)	4 mo/F	H	Pneumonia, bronchiolitis, fever, wheezing, rhinorrhea
HCoV-NL63 (2)	2 y/M	E	Pneumonia, fever
HMPV (5)	2 y 5 mo/M	H	Pneumonia, fever, cough
	6 mo/M	E	Bronchiolitis, fever, cough, vomiting, exanthem
HBoV (5)	4 y/M	C	Fever, cough, wheezing, rhinorrhea
	5 y/M	C	Sore throat, cough, rhinorrhea, vomiting
	11 mo/F	C	Fever, cough
	7 mo/F	C	Fever, rhinorrhea, cough, wheezing, vomiting, diarrhea
	2 y 5 mo/M	C	Pneumonia, fever, cough, wheezing
RV (33)	1 y/M	C	Fever, cough, rhinorrhea, wheezing, vomiting
	?/M	C	Sore throat, cough, rhinorrhea
	?/M	C	Fever, rhinorrhea
	1 y 8 mo/F	H	Pneumonia, fever, cough, wheezing
	1 y/F	C	Cough, rhinorrhea
	10 y/F	C	Fever, vomiting, headache
	3 y/F	E	Fever, cough, rhinorrhea, wheezing
	5 y/F	H	Fever, sore throat, cough, nasal congestion, asthma
	7 y/F	E	Fever, sore throat
	10 y/F	H	Rhinorrhea, asthma
	2 mo/F	C	Cough, rhinorrhea, nasal congestion, wheezing
	5 y/F	C	Cough, nasal congestion, wheezing, asthma
	2 mo/F	E	Fever, nasal congestion
10 mo/F	E	Fever, nasal congestion, cough, rhinorrhea	
1 y 4 mo/F	C	Pneumonia, bronchiolitis, fever, nasal congestion, cough, wheezing	
1 y/F	E	Pneumonia	
4 y/F	C	Fever, cough, nasal congestion, vomiting	
1 y/F	C	Cough, rhinorrhea, wheezing	
7 mo/F	C	Bronchiolitis, cough, rhinorrhea, wheezing	
8 mo/F	C	Fever, cough, rhinorrhea	
2 y/M	C	Rhinorrhea, vomiting, diarrhea	
6 mo/M	E	Broncospasm	
7 mo/M	H	Fever, cough, nasal congestion, rhinorrhea	
3 mo/M	H	Pneumonia, fever	
9 y/M	E	Cough, rhinorrhea, asthma	
12 y/M	H	Pneumonia, cough, wheezing, rhinorrhea	
1 y/M	C	Pneumonia, cough, rhinorrhea	
7 y/M	C	Pneumonia, cough	
6 y/M	H	Pneumonia, fever, rhinorrhea	
9 mo/M	E	Cough, rhinorrhea	
1 mo/M	H	Pneumonia, broncospasm, fever, rhinorrhea	
9 mo/M	H	Broncospasm, fever, rhinorrhea	
4 mo/M	H	Broncospasm, cough, rhinorrhea, nasal congestion	
10 mo/M	E	Pneumonia, fever, rhinorrhea	
3 y/M	C	Bronchitis, wheezing, cough	
7 mo/M	C	Bronchitis, wheezing, cough, rhinorrhea	
6 y/M	C	Fever, cough, rhinorrhea	
?/M	C	Fever, cough, rhinorrhea	
FluV A (5)	15 y/F	C	Sore throat, rhinorrhea, nasal congestion
RSV (3)	1 y/F	C	Fever, cough, rhinorrhea, nasal congestion, wheezing
	9 y/F	C	Sore throat, rhinorrhea
	10 y/F	H	Pneumonia, fever, cough
	2 y/M	H	Sore throat, rhinorrhea, nasal congestion
HCoV-OC43 (1)	9 mo/M	E	Fever, rhinorrhea
	4 mo/M	E	Fever, rhinorrhea
	5 mo/M	E	Fever, cough, rhinorrhea, nasal congestion
AdV (1)	3 mo/F	H	Bronchiolitis
AdV (1)	5 mo/F	C	Fever, cough, nasal congestion, wheezing
Co-infections			
HCoV-NL63 + HMPV	6 y/F	C	Sore throat, cough
HCoV-NL63 + RV	8 y/F	C	Pneumonia, fever, cough
HMPV + RV + HCoV-OC43 + AdV	11 y/F	E	Fever, rhinorrhea

HMPV + KIPyV	4 y/M	E	Fever, cough, rhinorrhea, wheezing
HBoV + RV	1 y/F	C	Pneumonia, fever, cough, wheezing, rhinorrhea
HBoV + WUPyV	10 mo	E	Cough, rhinorrhea, laryngomalacia
FluV A + RV	1 y 4 mo	C	Fever, rhinorrhea, diarrhea, ear pain
RSV + HCoV-OC43	5 mo/M	E	Bronchiolitis, rhinorrhea

*HCoV, human coronavirus; H, hospitalized; E, emergency department; HMPV, human metapneumovirus; C, walk-in clinics; HBoV, human bocavirus; ?, unknown; RV, rhinovirus; FluV A, influenza virus A; RSV, respiratory syncytial virus; AdV, adenovirus; KIPyV and WUPyV, human polyomaviruses.